Celecoxib and Cisplatin Synergistically Inhibit Oral Cancer Cell Proliferation Via Modulating the NOTCH 1 Signalling Pathway

Zuhair M. Mohammedsaleh1*, Mamdoh S. Moawadh1, Mohammed M. Jalal1, Abdulaziz S Al-Otaibi1, Nizar H. Saedi1, Rathinasamy Baskaran3, Chih-Yang Huang4 and V. Bharath Kumar5*

1Department of Medical Laboratory Technology, Faculty of Applied Medical Sciences, University of Tabuk, Tabuk 71491, Saudi Arabia.
2Department of Medical Microbiology, Faculty of Medicine, University of Tabuk, Tabuk 71491, Saudi Arabia.
3Department of Bioinformatics and Medical Engineering, Asia University, Taichung, Taiwan.
4Department of Medical Research, China Medical University Hospital, China Medical University, Taichung 404, Taiwan.
5Department of Medical Laboratory Science and Biotechnology, Asia University, Taichung 413, Taiwan.

ABSTRACT

Oral squamous cell carcinoma is the sixth most common cancer worldwide. Though there are significant improvements in diagnosis and treatment, the outcome is still very low. Thus, the development of novel therapeutic approaches is needed. In this study, we aimed to identify the inhibitory effect of celecoxib (CXB), cisplatin (Cis) and the combination of CXB and Cis and to explore the potential molecular mechanisms involved in oral cancer cells (OC-3 and SCC-9). MTT assay was used to check the cell viability, cell migration and invasion was performed to evaluate the metastasis and western blot to check the protein expression. The results showed that the combination of CXB and Cis synergistically inhibited cell proliferation, migration and invasion of OC-3 cells. The notch signalling pathway plays an oncogenic role in tongue squamous cell carcinoma. We observed that treatment with CXB and Cis decreased NOTCH1, JAG1, Pre 2, and ADAM9 expression and increased Numb protein expression. Our study demonstrated that Notch signalling is dysregulated in human OSCC and plays a role in cell proliferation.

INTRODUCTION

Oral squamous cell carcinoma (OSCC) is the sixth most common cancer worldwide, and accounts for 90% of all oral cancer types found in the mouth, tongue, and lips (Markopoulos, 2012; Suresh et al., 2019). The risk factors of OSCC include smoking, alcohol consumption, betel nut chewing and human papillomavirus (HPV) infection (Hashibe et al., 2009). Despite recent improvements in chemotherapy and surgery, the survival rate is still low as a result of the emergence of treatment resistance and recurrence (Hsu et al., 2019). Cisplatin, cis-diamminedichloroplatinum (II) Pt (NH₃)₂Cl₂ is widely used anticancer drug, used in the treatment of various cancer types, including OSCC (Grosch et al., 2001). The development of platinum-resistant OSCC is an important factor that leads to treatment failure.

CXB, a cyclooxygenase 2 (COX-2) inhibitor is an effective non-steroidal anti-inflammatory drug (NSAIDs). Targeted inhibition of COX-2 may be an effective strategy for treating several cancer types like prostate, colon, lung and liver cancer (Xu, 2002; Liu et al., 2017). CXB is also said to reverse the epithelial-to-mesenchymal transition (EMT), reduce cell motility, and restrict proliferation in oral cancer cells (Chiang et al., 2017). Furthermore, COX-2 expression was significantly downregulated after treatment with CXB alone or in combination with Cis compared with Cis alone in oral cancer cells (Li et al., 2010). Previous studies have suggested that a combination...
of the Cis with CXB exerts a synergistic anti-proliferative effect (Liu et al., 2017). Numerous studies have shown that combining CXB with additional medications may have synergistic anti-cancer benefits in a variety of cancer types (Qian et al., 2014; Gowda et al., 2017; Velmurugan et al., 2020; Srivastava et al., 2021).

Notch controls how cancer stem cells (CSCs) develop and helps cells acquire EMT phenotype, both of which are crucially linked to treatment resistance (Wang et al., 2009, 2010). The Notch signalling pathway may play dual roles such as tumour suppressor/ oncogene in different cancers (Gan et al., 2019), role of Notch signalling in OSCC remains controversial. It has been shown that OSCC expressed jagged canonical Notch ligand 1 (JAG1), a canonical notch ligand (Hijioka et al., 2010). Notch signalling inhibition has been shown to inhibit cancer cell proliferation/cell cycle progression, reduce cancer cell viability, and increase cell apoptosis in various cancer types (Purow, 2012). An earlier study hypothesized that COX-2 could accelerate the progression of gastric cancer by activating the neurogenic locus notch homolog protein 1 (NOTCH1) signal pathway (Yeh et al., 2009). Nevertheless, the combinatory effect of CXB and Cis on Notch activity in oral cancer cells has not yet been studied. Our study demonstrated that Notch signalling is dysregulated in human OSCC and plays a significant role in cell proliferation and migration.

MATERIALS AND METHODS

Chemicals and antibodies
Celecoxib (Pfizer) was purchased from a local pharmacy and dissolved in DMSO (Sigma Chemical Co). Cisplatin was purchased from local pharmacy and dissolved in normal saline (2 mg/ml) as a stock solution in 4 °C. CXB and Cis were added at various concentrations to cells in 10% fetal bovine serum (FBS)-containing DMEM. Antibodies NOTCH1 (#3268), Jag1 (#2620), Jag2 (#2210), Presenilin-1 (#5643) and 2 (#9979), Nicastrin (#3632) and Numb (#2756), A disintegrin and a metalloprotease 9 (ADAM9) (#4151), and RBPSUH (#5442) were purchased from cell signalling technology (Danvers, MA). β-actin (MABT825), GAPDH (AB2302) was purchased from Millipore (Billerica, MA). Secondary antibodies were obtained from Santa Cruz Biotechnology, Inc (Santa Cruz, CA, USA).

OSCC cell lines
SCC-9 and OC-3 cell lines were purchased from Food Industry Research and Development Institute, Hsinchu, Taiwan. SCC-9 cells were cultured in Dulbecco’s modified Eagle’s medium (DMEM-F12) supplemented with 40 ng/ml hydrocortisone containing 10% fetal bovine serum. OC-3 cells were cultured in a 1:2 mixture of DMEM and Keratinocyte Serum Free Medium (KSF) with 10% FBS (Gibco, Grand Island, NY, USA). All culture medium were supplemented with 1% penicillin and streptomycin and incubated in a 5% CO2, 37°C humidified incubator.

Cell viability assay
OC-3 cells were seeded into 96-well plates and incubated overnight at 37°C, which was followed by treatments CXB (100µM), Cis (20 µM) and in combination for 24 h. Cell growth was measured using 0.5 mg/ml 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (Sigma) colorimetric method. The blue MTT formazan precipitate was then dissolved in 200 µl of DMSO and absorbance was measured on a multiwell plate reader.

Migration and invasion assay
Cell migration and invasion was performed at a 24-well Transwell chamber with a pore size of 8 µm (Corning, Bedford, MA, USA). SCC-9 cells were mixed with cultured medium containing 0.5% FBS and seeded into the upper chambers of the insert coated with or without 100 µl Matrigel (dilution at 1: 2; Corning, Bedford, MA, USA). The inserts were placed in 24-well plates containing complete medium with CXB (100µM), Cis (20 µM) and in combination for 24 h in lower wells, respectively.

Western blot analysis
Cells were collected after treatment, twice washed with cold PBS, and then lysed in lysis buffer. NativePAGE™ 4-16% Bis-Tris gels from Thermofisher Scientific were used to produce whole cell lysates and fractionate them for SDS-PAGE. After that, proteins were transported to an Immobilon-P polyvinylidene difluoride membrane (Millipore, Bedford, MA, USA). SCC-9 cells were mixed with cultured medium containing 0.5% FBS and seeded into the upper chambers of the insert coated with or without 100 µl Matrigel (dilution at 1: 2; Corning, Bedford, MA, USA). The inserts were placed in 24-well plates containing complete medium with CXB (100µM), Cis (20 µM) and in combination for 24 h in lower wells, respectively.

Statistical analysis
The student t-test and one-way ANOVA were used to assess the experimental data from the in vitro experiments. P value <0.05 was considered statistically significant. All statistical analyses were performed using Prism 6.0 (GraphPad, San Diego, CA, USA).
RESULTS

Celecoxib and Cisplatin synergistically reduced SCC-9 cell proliferation on oral cancer cells

The effects of CXB combined with Cis on the viability of OC-3 cells were detected by MTT assay. After treated with 2.5–25 μM Cis (Fig. 1A) for 24 h, cell viability decreased significantly ($p < 0.001$) in a dose dependent manner. The IC$_{50}$ of treatment with Cis alone was 25 μM and CXB concentration was chosen from previous studies (Chiang et al., 2017a). As shown in Figure 1B, combination treatment with both drugs greatly decreased cell growth compared to CXB and Cis treatment alone (Fig. 1A). Our findings suggest that CXB combined with Cis had a synergistic anti-proliferation effect on OC-3 cells.

Celecoxib combined with Cisplatin inhibits migration and invasion of OC-3 cells

We then tested the ability of CXB, Cis, or CXB+Cis treated OC-3 cells to proliferate for 24 h. Compared to control cells, cells treated to CXB or Cis alone or in combination displayed a decreased potential for migration. In contrast to CXB or Cis treated cells, the combination group significantly inhibited the migration of OC-3 cells (Fig. 2A). The impact of CXB and Cis on the propensity of OC-3 cells to invade was then examined. In comparison to the control group, cell invasion was reduced in the treatment groups (Fig. 2B). The combo group showed much less OC-3 cell invasion than the CXB or Cis groups. The outcomes shown that CXB and Cis can prevent OC-3 cells from migrating and invading.

We then evaluated the expression difference of epithelial-mesenchymal transition (EMT) markers between control and treatment group. We found no significant changes in E-cadherin, Vimentin and Claudin expression (Fig. 2C).

Effect of CXB combined with Cis on NOTCH1 signalling related molecules in OC-3 cells

We then determined whether NOTCH1 signalling-related molecules are expressed in OSCC cell lines- OC-3, SCC-9, and the cell lysates were analysed using western blotting. These results demonstrate that NOTCH1, Jag1, Pre-1 and 2, Nic, Numb, RBPSUH and ADAM9 proteins were upregulated and Jag2 protein were downregulated in OC-3 cells (Fig. 3A). Based on Notch1 expression, we chose OC-3 cells to study the molecular mechanism. Our data indicated that combination group decreased NOTCH1, Pre-2, and ADAM9 expression and the expression of Numb was increased (Fig. 3B).
DISCUSSION

Accumulating evidence suggests that the anticancer activity of known chemotherapeutic drugs can be enhanced by using CXB (Irie et al., 2007). The proliferation of non-small cell lung cancer (NSCLC) cells was dramatically suppressed in vitro, and the growth of the tumour in vivo when low dosages of sorafenib (SOR) and CXB were combined (Zhang et al., 2014). In our previous study combination of CLA (1 nM) and CXB (50 µM) significantly inhibited cell viability, and migration and invasion of oral cancer cells (Velmurugan et al., 2020). Jeon and Suh (2013) demonstrated that a combination of CXB and luteolin greatly inhibited breast cancer cell growth than either CXB or luteolin treatment. In the present study, we provide clear evidence that the combined tumour treatment with Cis and CXB significantly decreased oral squamous cell carcinoma cell proliferation, migration, and invasion in vitro, compared with the single treatment.

To observe molecular target, this study targeted NOTCH1 as a key protein that are responsible for pathogenesis of several different types of cancer (Arai et al., 2018). Previous research emphasized that NOTCH1 acts as an oncogene in tongue cancer and induced tongue cancer cell proliferation and migration and inhibit cell apoptosis (Gan et al., 2018). Our recent finding showed that NOTCH1 plays an oncogenic role in oral squamous cell carcinoma (Data not published). Therefore, we first screened for NOTCH1 related molecular markers- Jag1, 2, Pres-1 and 2, Nic, Numb, RBPSUH and ADAM9 in OC-3 and SCC-9 cell lines. In the present study, OC-3 cells showed increased NOTCH1, Jagged1 (JAG1), Presenilin 1,2, Nicastrin, and RBPSUH expression. However, NOTCH1 was weakly expressed and Jagged 2 (JAG2), Numb was detected in SCC-9 cells. Based on NOTCH1 expression we chose to use OC-3 cell line to reveal the potential association between NOTCH1 signalling and oral cancer cell proliferation.

Notch receptors are activated by Notch ligands, Jag1 is a NOTCH1 ligand that triggers Notch signalling through cell-cell interactions (Xiu et al., 2020). The Notch receptor is cleaved by gamma secretase in the third cleavage (S3). A presenilin-dependent -secretase protease complex, consisting of presenilin 1 (PSEN1) or PSEN2, nicastrin, presenilin enhancer 2 (PEN2), and anterior pharynxdefective1 (APH1), regulates S3. Notch intracellular domain (NICD) would be released from the membrane to the cytoplasm following gamma secretase proteolysis of the Notch receptor (Fortini, 2002; Kopan and Ilagan, 2009; You et al., 2023). Previous studies indicated that ADAM9 mediates cancer progression via regulating EMT (Sarkar et al., 2015). However, our results indicating that Cis decreased NOTCH1, Pre2 and ADAM9 expression by increasing Numb expression. Numb over expression in human esophageal squamous cell carcinoma led to a decrease in cell proliferation, migration, and invasion (Hong et al., 2014). In this study we observed that CXB and combination treatment was not able to inhibit JAG1 expression, however Cis treatment alone decreased Jag1 expression compared to control cells. Without binding to NOTCH receptors, the JAG1 intracellular domain can stimulate tumour growth and epithelial-mesenchymal transition (Pelullo et al., 2019).

The current study only found that CXB and Cis combination therapy improved antitumour activity in the OC-3 cell line more than CXB or Cis treatment alone; this discovery was not confirmed in other oral cancer cell lines and requires further investigation.

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Data availability statement
The data that support the findings of this study are available from the corresponding author, Z.M.M, upon reasonable request.

Statement of conflicts of interest
The authors have declared no conflict of interest.

REFERENCES


