



Identification of Novel Biomarkers *MCM2* and *GINS2* for Cervical Cancer

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Key words

GINS2, *MCM2*, Cervical cancer, Prognosis, Differentially expressed genes

ABSTRACT

Cervical cancer (CC) is the most common malignant tumor in women, and its prognosis is poor. The key genes and pathways of CC need to be further discovered. GEO2R was used to identify differentially expressed genes (DEG), GO and KEGG enrichment were analyzed by DAVID. Then, the PPI network is constructed with STRINGS. The HUB gene and module of DEGS were obtained by Cytoscape. Finally, GEPIA also analyzed the differential expression and survival of key genes. 234 DEG were extracted from GSE9750. The uterus is the fourth organ highlighted in the concentrated analysis. The functional changes of DEGS are mainly related to cell cycle progression, cell cycle, helicase activity, DNA helicase activity, exosome and p53 signal pathway. In addition, five HUB genes and one key module were identified. Survival analysis showed that *MCM2* and *GINS2* were significantly correlated with overall survival. Expression analysis showed that *MCM2* and *GINS2* were highly expressed in cancer tissues, but low in normal tissues, which was consistent with the results of GEO analysis. Correlation analysis showed that there was a significant positive correlation between *MCM2* and *GINS2*. This study suggests that *MCM2* and *GINS2* may be new biomarkers to predict the prognosis of CC.

INTRODUCTION

Cervical cancer (CC) is a common gynecological malignant tumor, it is the fourth leading cause of cancer-related death in women all over the world and ranks eighth among the most common cancers (Cosper *et al.*, 2020). In recent decades, the incidence of CC in young women is on the rise (Lin *et al.*, 2019). There were an estimated 527,600 new cases and 265,700 deaths worldwide in 2012. The global death toll in 2018 was 311,000. Although the morbidity and mortality of CC have decreased due to the improvement of diagnosis and treatment in recent years,

the prognosis of secondary metastatic cancer and tumor recurrence is very poor (Nambaru *et al.*, 2009).

Although human papillomavirus (HPV) is a prerequisite for CC (Yuan *et al.*, 2018), only a few women infected with the virus develop cancer (Huijismans *et al.*, 2016). Therefore, other risk factors should be considered as auxiliary factors leading to the progression of CC (Luyten *et al.*, 2014). Abnormal regulatory genes play an important role in the occurrence and development of cervical squamous cell carcinoma (Li *et al.*, 2017; Alifu *et al.*, 2018). Many studies have identified the key genes in cervical squamous cell carcinoma and normal cervical tissues through gene expression profile technology, and a large number of differentially expressed genes (DEG) have been detected (Shah *et al.*, 2020). However, DEGs reported in different studies varies greatly, and only part of them are consistently detected. Thus, it is urgent to find new and effective targets for anti-CC therapy.

In this study, we selected the following microarray dataset GSE9750 from the Gene Expression Omnibus (GEO) database to identify DEGs. DEGs was analyzed through Kyoto Encyclopedia of Genes and Genomes

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(KEGG). A protein-protein interaction (PPI) network was constructed to clarify the important relationship between DEGs and to find HUB genes. In addition, the differential expression and survival analysis of HUB genes were carried out on Gene Expression Profiling Interactive Analysis (GEPIA). The purpose of this study is to better understand the characteristics of the genes and signal transduction pathways related to CC through bioinformatics analysis.

MATERIALS AND METHODS

Microarray data

The gene expression profiles of GSE9750 were downloaded from GEO database. GSE9750, which was based on GPL96 platform (Affymetrix Human Genome U133A Array). The GSE9750 dataset contained 33 tumor samples and 24 normal cervical samples.

Identification of DEGs

The online tool, GEO2R was applied to determine DEGs in normal cervical tissues and CC tissues. Adjusted P-values were used to reduce the false positive rate using the Benjamini and Hochberg false discovery rate method by default. Adjusted $P \leq 0.05$ and $|\log \text{fold change (FC)}| \geq 1.5$ were set as cut-off values. A total of 234 DEGs were then identified, including 55 up-regulated and 179 down-regulated genes. Eventually, the top 5 genes were determined as hub genes ranked by the Degree method in cytoHubba, a plugin in Cytoscape 3.6.0 software.

GO and KEGG enrichment analysis

The database for annotation, visualization and integrated discovery is a public online bioinformatic database which helps to identify the most significant enriched functional genes and biological pathways. To further analyze the DEGs, GO and KEGG enrichment analyses were performed by using the DAVID. GO analysis was used to annotate biological process (BP), cytological component (CC), and molecular function (MF) of genes, and KEGG enrichment analysis was used to understand the relevant signaling pathways, p value < 0.05 was considered to be statistically significant.

PPI network and key module analysis

We constructed the protein-protein interaction network (PPI) of DEGs by using Search Tool for the Retrieval of Interacting Genes database based on the confidence scores. What's more, we further visualized the PPI by Cytoscape. And the Molecular Complex Detection (MCODE) plugin in Cytoscape was used to filter the key modules in the network with degree cutoff = 2, node score cutoff = 0.2, k-core = 2, and max. depth = 100. The criteria

were set as follows: MCODE scores > 3 and number of nodes > 4 .

Key genes screening and analysis

The genes with degree ≥ 10 in the network were identified as key genes, Gene Expression Profiling Interactive Analysis (GEPIA) is an interactive web application for gene expression analysis. We visualized the expression of key genes in CC tissues and normal tissues by box plots in GEPIA, and the overall survival analysis and correlation analysis of key genes was also performed.

RESULT

Identification of DEGs

There were 33 CC tissues and 24 normal cervical tissue samples analysed in this study. Firstly, the GEO2R tool was employed to identify DEGs using the following cut-off values: Adjusted $P \leq 0.05$ and $|\log \text{FC}| \geq 1.5$. As a result, a total of 234 DEGs were identified, including 55 up-regulated and 179 down-regulated genes (Fig. 1).

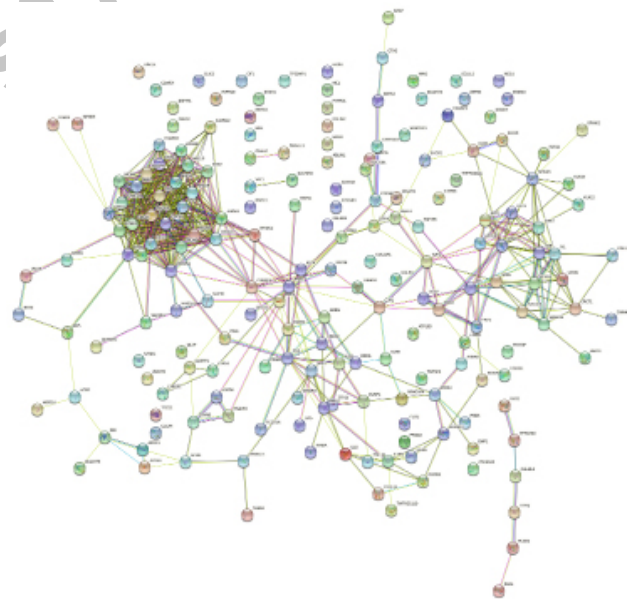


Fig. 1. Protein-protein interaction network of 234 DEGs.

GO enrichment and KEGG pathway enrichment

We uploaded all DEGs to DAVID to identify overrepresented GO categories and KEGG pathways. The enriched results of tissue expression were revealed, the screened DEGs were enriched in tissues including Foreskin, Keratinocyte, Skin, Tongue, Esophagus, Bone marrow, Plasma, Uterus, Placenta and Epithelium (Table I). In the GO analysis, the screened DEGs mainly participate

Table I. Enrichment analysis of 234 differentially expressed genes in different tissues.

Category	Term	Count	%	P-value	FDR
UP_TISSUE	Foreskin	6	3.14	1.84E-04	0.219073889
UP_TISSUE	Keratinocyte	8	4.19	4.55E-04	0.540274684
UP_TISSUE	Skin	33	17.3	0.002057593	2.424061768
UP_TISSUE	Tongue	10	5.24	0.026234963	27.14758832
UP_TISSUE	Esophagus	3	1.57	0.033587692	33.43796859
UP_TISSUE	Bone marrow	15	7.85	0.034025143	33.79604522
UP_TISSUE	Plasma	7	3.66	0.067794744	56.67257722
UP_TISSUE	Uterus	25	13.09	0.074480313	60.23314132
UP_TISSUE	Placenta	43	22.51	0.085638329	65.58380022
UP_TISSUE	Epithelium	35	18.32	0.092928299	68.71433346

in the biological process (BP) of epithelial cell differentiation, epidermis development, skin development, epidermal cell differentiation, mitotic cell cycle process, keratinocyte differentiation, epithelium development, cell cycle process, cell cycle and mitotic cell cycle. As for the molecular function (MF), DEGs mainly involved in serine-type peptidase activity, serine hydrolase activity, serine-type endopeptidase activity, structural molecule activity, helicase activity, DNA helicase activity and calcium ion binding. The cell component (CC) of DEGs includes extracellular region part, extracellular exosome, extracellular vesicle, extracellular organelle, membrane-bounded vesicle, extracellular region, cornified envelope and apical plasma membrane. In the KEGG pathway analysis found that DEGs significantly enriched in Gap junction, Estrogen signaling pathway and p53 signaling pathway (Table II).

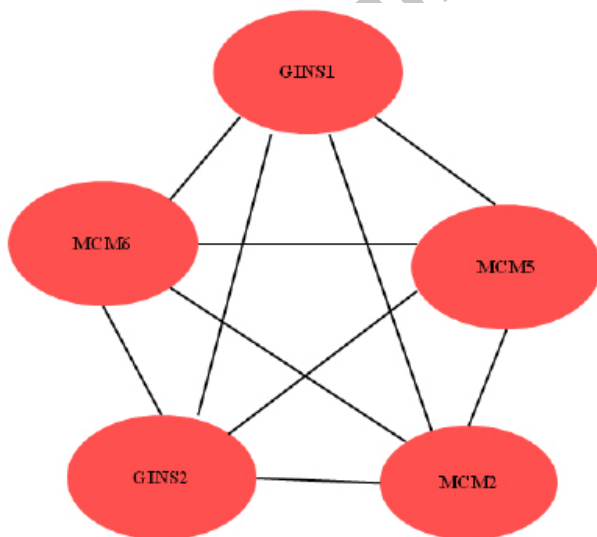


Fig. 2. Up-regulate the key modules of gene composition. The red nodes represent up-regulated genes.

PPI network construction and analysis

PPI network of 234 DEGs was constructed in STRING, the network visualized by Cytoscape. MOCD plug-in screened out one key modules, which were composed by up-regulated genes (Fig. 2). Red nodes represent up-regulated genes. Degree ≥ 10 as screening criteria, 5 key genes were screened to form key modules; Their names are shown in Table III.

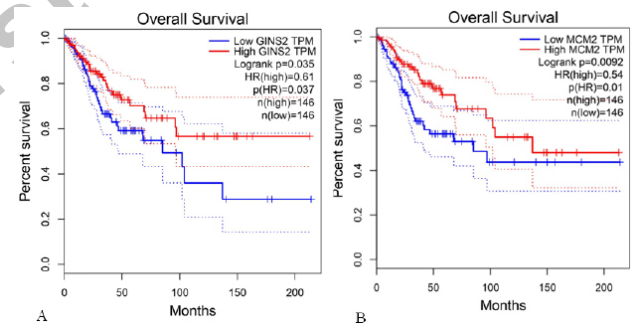


Fig. 3. Survival analyses of *GINS2* and *MCM2*. *GINS2* (A) and *MCM2* (B) were significantly associated with OS.

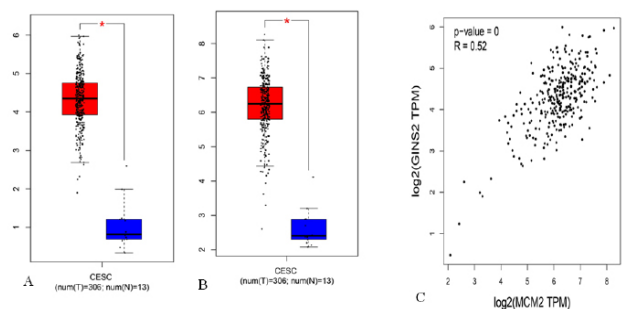


Fig. 4. Expression levels and Pearson correlation analyses between *GINS2* and *MCM2*. Both *GINS2* (A) and *MCM2* (B) presented high expression levels in CC tissues, but exhibited low expression levels in normal cervical tissues. (C) *GINS2* was positively correlated with *MCM2*.

Table II. GO and KEGG pathway enrichment analysis of DEGs.

Category	Term	Count	P value
Biological processes			
GO:0030855	epithelial cell differentiation	28	3.93E-11
GO:0008544	epidermis development	21	5.41E-11
GO:0043588	skin development	19	5.81E-11
GO:0009913	epidermal cell differentiation	16	6.22E-10
GO:1903047	mitotic cell cycle process	32	3.38E-09
GO:0030216	keratinocyte differentiation	13	4.45E-09
GO:0060429	epithelium development	34	1.56E-08
GO:0022402	cell cycle process	39	1.82E-08
GO:0007049	cell cycle	44	2.06E-08
GO:0000278	mitotic cell cycle	32	2.40E-08
Cell component			
GO:0044421	extracellular region part	80	2.97E-08
GO:0070062	extracellular exosome	64	6.07E-08
GO:1903561	extracellular vesicle	64	7.35E-08
GO:0043230	extracellular organelle	64	7.45E-08
GO:0031988	membrane-bounded vesicle	74	2.20E-07
GO:0005576	extracellular region	87	3.49E-07
GO:0001533	cornified envelope	8	1.01E-06
GO:0016324	apical plasma membrane	15	9.63E-06
Molecular function			
GO:0008236	serine-type peptidase activity	12	2.35E-04
GO:0017171	serine hydrolase activity	12	2.57E-04
GO:0004252	serine-type endopeptidase activity	11	4.33E-04
GO:0005198	structural molecule activity	20	7.30E-04
GO:0004386	helicase activity	8	0.001640
GO:0003678	DNA helicase activity	5	0.003037
GO:0005509	calcium ion binding	17	0.004460
KEGG pathway			
hsa04540	Gap junction	5	0.018922
hsa04915	Estrogen signaling pathway	5	0.027773
hsa04115	p53 signaling pathway	4	0.04265

Key gene analysis

All aforementioned 5 hub genes were analyzed using the prognostic values of OS and DFS via the GEPIA website. *GINS2* and *MCM2* were significantly associated with OS (log-rank $P=0.035$ and 0.0092 (Fig. 3A and B)). The analysis of these two genes revealed that low expression levels led to better survival status. The other hub genes did not exhibit statistical significance. The genes *GINS2* and

MCM2 were then subjected to further analysis. Expression levels of these two genes are displayed in Figure 4A, B. Both *GINS2* and *MCM2* presented high expression levels in CC tissues, but exhibited low expression levels in normal cervical tissues. In addition, Pearson correlation analyses between the genes are presented in Figure 4C. Results revealed that *GINS2* was positively correlated with *MCM2* ($R=0.52$, $P=0$).

Table III. Functional roles of 5 key genes with degree ≥ 10 .

Gene symbol	Degree	Full name
<i>MCM2</i>	26	minichromosome maintenance complex component 2
<i>MCM5</i>	26	minichromosome maintenance complex component 5
<i>GINS1</i>	24	GINS complex subunit 1
<i>GINS2</i>	24	GINS complex subunit 2
<i>MCM6</i>	24	minichromosome maintenance complex component 6

DISCUSSION

In the study, we investigated the potential prognostic association between CC and DEGs in GSE9750. The results showed that there were 234 DEGs in 24 normal cervical tissues and 33 CC tissues, including 55 up-regulated genes and 179 down-regulated genes. Both up-regulated and down-regulated genes were enriched in multiple organs. Notably, the uterus was the fourth organ highlighted in the enrichment analysis. Five hub genes were screened and one module was identified. *GINS2* and *MCM2* genes have potential prognostic value in patients with CC.

Although the morbidity and mortality of CC have declined in recent years due to improvements in diagnosis and treatment, the clinical outcome of advanced diseases is still very dim (Cuschieri *et al.*, 2014). In addition, lymph node metastasis can lead to higher mortality and recurrence rates (Liu *et al.*, 2020), even in patients with early CC (Li *et al.*, 2016), and exact lymph node status information is essential for tailored adjuvant therapy (Zhang *et al.*, 2014). However, so far, there are no sensitive biomarkers that specifically reflect the indications of lymph node metastasis, as well as the early detection and prognosis of CC.

GINS complex 2 (*GINS2*), also known as *PSF2*, encodes a protein with a molecular weight of about 21 kDa (Ye *et al.*, 2019). *GINS2* belongs to the *GINS*

complex family, which also includes *GINS2*, *GINS3* and *GINS4*. *GINS* complex plays an important role in initiating DNA replication and cell cycle (Chi *et al.*, 2020). The *GINS* family is involved in the maintenance of micro chromosomes (*MCM*) 2-mel-7 complex and *cdc45* maintain stable interaction, which can correctly establish and maintain DNA replication bifurcations (Peng *et al.*, 2016). In addition, the *GINS* component may play a role in cell division or, more accurately, in chromosome segregation. Ouyang *et al.* (2017) found that *GINS2* gene knockout inhibits the proliferation, tumorigenicity, migration and invasion of CC cells (Ouyang *et al.*, 2017). This is consistent with the results of this study, high expression of *GINS2* in CC, low expression in normal cervical tissues, therefore, low expression of *GINS2* leads to better survival. These results suggest that *GINS2* may be a new index to identify high-risk patients and may be used as a clinical biomarker to predict the prognosis of patients with early CC.

In addition, *GINS2* is reported to be associated with several other types of cancer (Yan *et al.*, 2018). Such as, genome-wide gene expression profile analysis shows that *GINS2* is highly expressed in lung cancer (Liu *et al.*, 2019). Zheng *et al.*, (2014) believe that *GINS2* is related to the invasiveness of breast cancer and speculate that it is related to lung metastasis (Zheng *et al.*, 2014). Besides, the increased expression of *GINS2* can promote the proliferation of leukemic cells and reduce the sensitivity of leukemic cells to apoptosis (Gao *et al.*, 2013). These findings suggest that *GINS2* plays an important role in cancer progression.

A series of events during HPV infection can cause host cells to experience an unplanned cell cycle (Hernadi *et al.*, 2003). This phenomenon leads to cell division out of control, promotes cell proliferation, and then leads to cancer. In HPV-related CC, cancer cells up-regulate specific genes that control several steps of DNA replication. Microchromosome maintenance complex (*MCM*) is an essential protein in DNA replication and cell division. Kaur *et al.* (2019) showed that the expression of *MCM* gene was up-regulated during the carcinogenesis of CC, which once again proved that *MCM* is a proliferation marker in the DNA replication pathway, which makes dysplasia and cancer cell proliferation more and more out of control (Kaur *et al.*, 2019). *MCMs* is a candidate marker of cell proliferation. The increase of *MCMs* level indicates the proliferation of malignant cells. This is consistent with the results of this study, the expression of *MCM2* is high in CC tissue and low in normal cervical tissue.

In addition, there is some evidence that *MCM* can predict tumor progression. Studies have shown that *MCM* protein is highly expressed in several other types of

cancers (Pruitt *et al.*, 2007), such as lung cancer (Cheung *et al.*, 2017), breast cancer (Yousef *et al.*, 2017; Issac *et al.*, 2019), colon cancer (Burger, 2009), and other cancers (Deng *et al.*, 2019) *MCM* protein may also be used as a potential diagnostic or prognostic marker for them.

CONCLUSION

In this study, 234 DEGs and 5 hub genes were identified in CC patients. Only *MCM2* and *GINS2* genes have prognostic value in patients with CC. This study suggests that *MCM2* and *GINS2* may be potential prognostic biomarkers of CC. In addition, *MCM2* and *GINS2* may play a carcinogenic role in cervical tumors. These genes may play a role through cell cycle process, cell cycle and helicase activity, DNA helicase activity, exosome and p53 signal pathway. Further research is needed to explore the functional role of these genes, especially in metastasis and cancer progression, in order to guide the clinical direction. The purpose of this study is to better understand the characteristics of cervical cancer-related genes and signal transduction pathways by means of bioinformatics, and to provide further research ideas for discovering new pathogenesis, more prognostic factors and potential therapeutic targets of CC.

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Statement of conflict of interest

The authors have declared no conflict of interest.

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