Short Communication

Protective Effects of Oviductus Ranae Mediated HPA Axis Regulation on Depressive Model of Mice

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ABSTRACT

Oviductus ranae (OR) is an animal-based traditional medicine. To explore its mechanism of antidepressant effect, healthy Institute of Cancer Research (ICR) male mice were divided into control, chronic unpredictable mild stress (CUMS), fluoxetine (3 mg/kg), OR800 (OR 800 mg/kg) and OR400 (OR 400 mg/kg) dose groups. After the last administration, the content of corticotropin-releasing hormone (CRH), adrenocorticotropic hormone (ACTH) and corticosterone (CORT) in serum were detected. Hematoxylin and Eosin (H&E) staining was used to observe the hippocampal histomorphological changes. The protein levels of glucocorticoid receptor (GR), mineralocorticoid receptor (MR), brain-derived neurotrophic factor (BDNF), tropine kinase B (TrkB) and cyclic adenosine monophosphate response element binding protein (CREB) in hippocampus were detected by western blotting. The results showed that, OR could significantly reduce the contents of CRH, ACTH and CORT in serum and improve the damage of hippocampal structure, and increase the expression levels of GR, MR, BDNF, TrkB and CREB in hippocampus. In conclusion, Oviductus ranae can reduce hypothalamic-pituitary-adrenal (HPA) axis hyperfunction induced by CUMS, increase the expression level of BDNF related signal pathway proteins, improve hippocampal tissue damage, and thus play an antidepressant role by regulating the negative feedback of HPA.

Depression is a common disease of mental disorder. Its pathological manifestations include the decrease of monoamine neurotransmitters level (Zhao et al., 2019; Naoi et al., 2018) and brain-derived neurotrophic factor (BDNF) (Phillips, 2017), the increase of inflammatory factors (Shelton et al., 2011), the hyperfunction of hypothalamic pituitary adrenal (HPA) axis (Keller et al., 2017), and the injury of hippocampal tissue (Pei et al., 2020). A large amount of cortisol secretion in the plasma of patients, accompanied by a decrease of BDNF level (Katz et al., 2017), which leads to reduction of dendritic complexity and changes in synaptic plasticity, thus affecting the normal physiological functions of the central system (Roversi et al., 2019). The regulatory target of HPA axis and a large amount BDNF can complete the negative feedback regulation of HPA axis through two corticosterone receptors, GR and MR, in hippocampus (Meyer et al., 2001). CREB and TrkB are proteins closely related to BND. CREB modified by phosphoric acid can promote the expression of BDNF, so as to promote neuron growth and protecting neurons. TrkB binds to BDNF as a receptor to promote BDNF for nerve cell survival, neurogenesis, regeneration and repair of nerve injury, and the role of plasticity synapses (Ge et al., 2015; Shirayama et al., 2020). Some existing drugs may cause side effects such as diarrhea or constipation, sexual problems, etc. (Uddin et al., 2017; Kikuchi et al., 2013). Thus, finding more effective and reliable new antidepressants has become a research hotspot.

Oviductus ranae (OR), the dried oviduct of mature female Rana temporaria chensinensis David, is an animal-based crude drug. It contains a variety of nutrients, including polyunsaturated fatty acids (PUFA), proteins and vitamins (Guo et al., 2019), thus is widely used as tonic (Xu et al., 2018). OR has effects of enhancing immunity, anti-aging, reducing blood lipid and antidepressant. Some fatty acids could inhibit the activation of HPA axis by regulating intestinal microorganisms and promote the expression of...
BDNF in Müller glia cells (Suzumura et al., 2020), these may be the biological basis of OR on antidepressant. However, its mechanism of antidepressant is still unclear.

In this study, the chronic unpredictable mild stress (CUMS) was used to establish the depression model. This study aims to provide a theoretical and experimental basis for a safe, effective and rational clinical application of OR.

Materials and methods

Oviductus ranae was provided by Tonghua Dingshen Pharmaceuticals Company (Tonghua, China). Fluoxetine hydrochloride capsules were purchased from Sinochem Pharmaceutical Industry Co., Ltd. (Suzhou, China). Institute of Cancer Research (ICR) male mice were purchased from Yisi Experimental Animal Technology Co., Ltd (Changchun, China). The other main reagents and equipment are listed in Supplementary Table SI.

Fifty male ICR mice were housed for 7 days under standard conditions at temperature 24 ± 1°C, 12 h:12 h light: dark cycles, with normal diet and drinking water. The mice were randomly divided into 5 groups (10 mice/group): control (equal volume of distilled water), CUMS (fluoxetine 3 mg/kg), OR800 (800 mg/kg) and OR400 (400 mg/kg) dose groups, and administered intragastrically.

The CUMS model was established and evaluated accordingly (Su et al., 2017; Liu et al., 2018). The control group was normally fed, while the other groups received 28 days of chronic unpredictable stressors. The details including: electric foot-shock (24 V for 5 min), fasting and water deprivation (24 h), tilting the cage at 45° (24 h), force swimming in ice water at 4°C (3 min), shaking the cage horizontally (10 min), hot environment at 45°C (5 min), noise stimulation (10 min) and tail-clamping (5 min). To prevent mice from adapting, 1 to 2 different stressors listed above were given a day.

Mice in each group were anesthetized, their blood samples were taken from eyeballs and centrifuged 3000 r/min at 4°C for 10 min. The upper serum was collected and placed in a 2 mL EP tube, and stored at -80 °C. ELISA was used to detect the contents of CRH, ACTH and CORT in serum (Song et al., 2018).

After the mice were sacrificed, the brain tissue was collected, and the hippocampal tissues were quickly separated on ice and put into 4% paraformaldehyde fixative. After dehydration, paraffin embedding and sectioning, the hippocampal tissue was sliced into paraffin sections and the changes of tissue structure were observed by H&E staining under a microscope (Song et al., 2018).

Hippocampal tissue samples were placed in tube, lysed in lysis buffer, homogenized on ice, centrifuged to obtain the supernatant and stored at -80°C for later use. The concentration of protein in hippocampus was measured by bicinchoninic acid (BCA) assay. Proteins were resolved on SDS-PAGE by electrophoresis and are transferred to PVDF membrane (Mishra et al., 2017). After incubation with specific antibody at 4°C overnight, the membrane was exposed to secondary antibody for 1 h, washed with TBST for 3 times. Then, chemiluminescence and gel imaging analyses were performed. The ratio of target protein to internal reference GAPDH was used as the relative protein expression.

All data were presented as the mean of three samples with standard deviation. One-way analysis of variance (ANOVA) and Tukey’s range test were used to determine differences between groups, \( p < 0.05 \) was considered as statistically significant.

Results

The contents of CRH, ACTH and CORT in serum are shown in Figure 1A. In CUMS group, the contents of CRH, ACTH, and CORT were increased significantly compared with that of the control group \( (p < 0.01) \). Compared with the CUMS group, the contents of serum CRH, ACTH and CORT in fluoxetine group, OR800 and OR400 groups were significantly reduced \( (p < 0.01) \). Long-term stress can cause HPA axis hyperfunction and increase the contents of CRH, ACTH and CORT.

The pathological changes in hippocampal tissues were shown in Figure 1B. The hippocampal CA1 cells in control group were normal, plump, dense, arranged neatly and no obvious atrophy, while that of the CUMS group showed cells atrophy, decreased density, disordered arrangement, large gaps in the middle, and obvious degree of damage. Compared with CUMS group, the structure of cells in the OR800 and OR400 groups became normal gradually, density increased, the space between the cells decreased, the overall arrangement was more orderly, and the damage was significantly alleviated.

The expression of BDNF, TrkB, CREB, GR and MR proteins in hippocampal tissues were shown in Figure 1C. Compared with control group, the expression of BDNF, TrkB, CREB, GR, and MR proteins in CUMS group decreased significantly \( (p<0.01) \). While the fluoxetine group and OR800 and OR400 groups were significantly up-regulated \( (p<0.01) \), compared with that of CUMS group. The results showed that OR could enhance the inhibition of HPA axis by increasing the cortical hormone receptor, reduce the damage and repair the hippocampal tissue by increasing the neurotrophic factors and receptors.

Discussion

When exposing under long-term chronic pressure, the HPA axis of the body will be activated. CRH secreted by hypothalamus can stimulate the pituitary gland to secrete ACTH, then stimulated adrenal glands to release a large amount of glucocorticoids, resulting in the increase of...
glucocorticoids in body. Hippocampus plays an important role in the condition of HPA axis. Glucocorticoids in body will first bind to MR in hippocampus at a low level; only when at a high level, it will bind to GR. Meanwhile, hippocampus regulation inhibits the excessive secretion of HPA axis, maintains it to a steady state, and normalize glucocorticoid levels in vivo (Chen et al., 2016). The results in this study indicated that can increase the contents of hippocampal GR and MR receptors, enhance the hippocampus’ negative feedback regulation of the HPA axis, reduce the body’s glucocorticoids and relieve the hippocampal tissue damage caused by excessive glucocorticoids.

BDNF exists in hippocampus and cortex, which participates in the growth and development of neurons, and also protects the physiological functions of neurons from being damaged. The increase of glucocorticoids leads to the low expression of BDNF and damages the nervous system from being repaired, resulting in depression (Serra et al., 2018). As a receptor with high affinity with BDNF, TrkB can activate downstream signaling pathways, such as mitogen-activated protein kinase (MAPK), phosphoinositide 3 kinase (PI3K), etc., and play biological effects after binding to BDNF (Li et al., 2018). Phosphorylated CREB can combine with CRE of downstream BDNF sequence, play biological functions and promote the expression of BDNF (Peng et al., 2018). The results showed that the contents of CREB, BDNF, TrkB in CUMS group decreased significantly, however, their contents in OR800 and OR400 groups increased significantly after administration. H&E staining showed that the mice in CUMS group had obvious structural damages such as cell atrophy, reduced density and more disordered arrangement, while that in OR800 and OR400 groups showed a gradually normal in cell structures, an increase in the density and significantly reduced damage, compared with CUMS group.

Studies showed that oleic acid and linolenic acid were two fatty acids in OR. It was found that linolenic acid could reduce the incidence of female depression (Lucas et al., 2011), oleic acid can play an antidepressant role as an inhibitor of autoinducer-2 (AL-2) (Medina-Rodriguez et al., 2020). These fatty acids may be the material basis for the antidepressant effect of OR, which need to be further studied.

Conclusion
Oviductus ranae has an antidepressant effect. Its mechanism is to restore the excessive activation of the HPA axis caused by stress and reduce glucocorticoids, alleviate the decline in BDNF caused by glucocorticoids, and promote the expression of BDNF by up-regulating CREB. Moreover, up-regulate the expression level of TrkB further for activating downstream signaling pathways, promote the neuron growth and neuron protection of neurotrophic factors, promote the repair of hippocampal tissue damage.

Acknowledgment
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Ethics statement
This study was approved by the Ethics Committee of Changchun University of Traditional Chinese Medicine (Approval No: 2020107).

Supplementary material
There is supplementary material associated with this article. Access the material online at: https://dx.doi.org/10.17582/journal.pjz/20210513050530
Statement of conflict of interest

The authors have declared no conflict of interest.

References


Supplementary Material

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Supplementary Table S1. Resources of main reagents and equipment.

<table>
<thead>
<tr>
<th>Main reagents and equipment</th>
<th>Resources</th>
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<tbody>
<tr>
<td>Hematoxylin and eosin (H&amp;E) stain solution</td>
<td>Beijing Beyotime Biotech, Beijing, China</td>
</tr>
<tr>
<td>Corticotropin-Releasing Hormone (CRH), Adrenocorticotropic Hormone (ACTH) and Corticosterone (CORT) enzyme-linked immunosorbent assay (ELISA) kits</td>
<td>Jiangsu Meimian Biotech, Jiang Su, China</td>
</tr>
<tr>
<td>BCA Protein Assay Kit</td>
<td>Solarbio, Beijing, China</td>
</tr>
<tr>
<td>BDNF, Trosine kinase B (TrkB), Cyclic adenosine monophosphate response element binding protein (CREB), Glucocorticoid Receptor (GR), Mineralcorticoid Receptor (MR) and reduced glyceraldehyde-phosphate dehydrogenase (GAPD)</td>
<td>San Ying Biotechnology, Wuhan, China</td>
</tr>
<tr>
<td>Gas chromatograph</td>
<td>Agilent Technologies, Wilmington, DE, USA</td>
</tr>
<tr>
<td>Chemiluminescence imaging system</td>
<td>TECAN, Männedorf, Switzerland</td>
</tr>
<tr>
<td>Embedding center and cooling stage, automatic water bath, automatic slide dryer</td>
<td>Taiwei Technology, Hubei, China</td>
</tr>
<tr>
<td>Slicer, Full-wavelength microplate reader</td>
<td>Thermo Fisher Scientific Inc., Waltham, MA, USA</td>
</tr>
<tr>
<td>Electrophoresis system, transfer tank</td>
<td>Bio-Rad, Hercules, CA, USA</td>
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