Effect of Diet Supplemented with Rumen-protected 5-Hydroxytryptophan on the Concentration of 5-Hydroxytryptophan and Melatonin in the Plasma of Sheep

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A B S T R A C T

This study was conducted to investigate the effect of diet supplemented with rumen-protected 5-hydroxytryptophan (RPT 5-HTP) on the concentration of 5-hydroxytryptophan (5-HTP) and melatonin (MLT) in the plasma of sheep. Eighteen sheep were assigned randomly to three diet groups (n = 6). The treatment groups included control (CT, corn-soybean meal basal diet), CT + 111 (CT + 111 mg/kg BW RPT 5-HTP), and CT + 222 (CT + 222 mg/kg BW RPT 5-HTP) groups. The experiment lasted for 16 d. On the 16th day of the experiment, blood samples from the sheep were collected at 0 h before feeding and at 1.5, 3, 4.5, 6, 8, 10, and 12 h after feeding in the morning. The plasma concentration of 5-HTP, tryptophan (Trp), 5-hydroxytryptamine (5-HT), and MLT was determined. The plasma concentration of 5-HTP at 6, 8, and 10 h after feeding in the morning increased (P < 0.05) in response to RPT 5-HTP supplementation when compared with that at 0 h before feeding. The plasma concentration of Trp remained unchanged with RPT 5-HTP supplementation. However, the plasma concentration of 5-HT increased significantly in the sheep of CT + 222 group at 3, 4.5, 6, 8, and 12 h post feeding in the morning when compared with that at 0 h before feeding. The effect of RPT 5-HTP supplementation on the plasma MLT concentration was observed. The concentration of MLT increased (P < 0.05) from 1.5 to 6 h after feeding. In conclusion, RPT 5-HTP supplementation increased the plasma 5-HTP, 5-HT, and MLT concentrations in the sheep. However, the plasma Trp concentration in the sheep remained unchanged after RPT 5-HTP supplementation.

INTRODUCTION

Melatonin (MLT), a circadian rhythm regulatory hormone (Arendt, 1998), controls the expression of clock genes at the central and peripheral tissue levels in organisms (Alonso-Vale et al., 2008; de Farias Tda et al., 2015; Vriend and Reiter, 2015). Melatonin regulates several physiological functions related to circadian alterations (Dijk and Cajochen, 1997; Miller et al., 2006; Pandi-Perumal et al., 2005; Zhdanova, 2005). It is a potent free radical scavenger and antioxidant (Reiter et al., 2016). It not only directly interacts with reactive oxygen species (ROS) and reactive nitrogen species (RNS), but also up-regulates the activities of antioxidant enzymes (Ding et al., 2014; Galano et al., 2011, 2013; Manchester et al., 2015). Furthermore, MLT (10^-7 mol/L) exhibited potent effects on porcine oocyte maturation under heat stress (Li et al., 2016). MLT also promotes bovine embryonic development under both in vitro and in vivo conditions (Wang et al., 2014). In addition, MLT advances the onset of estrus in adult ewe (English et al., 1986). Overall, MLT is secreted by the pineal gland mainly during dark period, and the changing circadian pattern of MLT secretion mediates the effect of changing photoperiod (day length) on various physiological functions in several species.

Based on the extensive biological effects of MLT, appropriately increasing the MLT content in animals might be of significance. Increased intake of MLT precursor material through diet can increase the MLT content in animal blood. Tryptophan (Trp) increases the plasma MLT level in monogastric animals, such as chicken, rat (Huether et al., 1992), and mice (Sanchez et al., 2004). In contrast, Trp supplementation in sheep is not an effective approach to increase blood 5-hydroxytryptamine (5-HT) or MLT synthesis (Sugden, 1989). In sheep, there was an interspecific difference in the plasma...
However, L-5-hydroxytryptophan (5-HTP, 200 mg/kg BW i.p.), but not Trp (500 mg/kg BW i.p.), supplementation substantially increased the serum MLT concentration in sheep (Sugden et al., 1985). The serum MLT concentration was elevated at 2–5 h after the injection of 20 or 200 mg/kg BW in sheep (Namboodiri et al., 1983).

5-HTP is synthesized from Trp by the action of Trp hydroxylase. Tryptophan hydroxylase transfers a hydroxyl group to the C5 position of the indole ring to give 5-HTP. Aromatic amino acid decarboxylase removes the side-chain carboxyl group to give 5-HT. MLT is synthesized from serotonin in a two-step process by the sequential action of pivotal enzymes, aralkylamine N-acetyltransferase (AA-NAT) and hydroxylindol-O-methyl transferase (Maronde and Stehle, 2007).

Therefore, in the present study, we hypothesized that the RPT 5-HTP can elevate the plasma 5-HT concentration, and thus, will affect the synthesis of MLT in sheep during daytime. The effects of RPT 5-HTP supplementation on the concentration of Trp and 5-HTP, which are the two major precursors of MLT synthesis, were observed.

MATERIALS AND METHODS

L-5-Hydroxytryptophan was purchased from Wuhan Yuancheng Gongchuang Technology Co. Ltd. (Wuhan, China). Additionally, RPT L-5-HTP (containing 45% L-5-HTP) was procured from Beijing Yahe Products Co. Ltd. (Beijing, China).

Animals and design

This study was conducted using 3-yr-old Kazakh sheep (female) with an average body weight of 47.79 ± 3.70 kg in the Xinjiang Huikang Animal Husbandry Biotechnology Co. Ltd. (Beijing, China).

Eighteen sheep were randomly assigned to three diet groups (n= 6). The treatments groups included CT (control group, corn-soybean meal basal diet), CT + 111 (CT + 111 mg/kg BW RPT 5-HTP), and CT + 222 groups (CT + 111 mg/kg BW RPT 5-HTP); 5-HTP content in CT + 222 was two times higher than that in CT + 111. The experimental period lasted for 16 d. Rumen-protected L-5-hydroxytryptophan was added to the concentrate diet. The levels of RPT 5-HTP were chosen according to the study of Sugden et al. (1985). Based on the differences in weight and breed, we set two levels of supplementation 111 and 222 mg/kg BW RPT 5-HTP. On the 16th day of the experiment, blood samples were drawn from the external jugular vein during daytime, at 0 h before feeding and at 1.5, 3, 4.5, 6, 8, 10, and 12 h after feeding. The samples were then centrifuged at 120 ×g for 10 min to obtain the plasma for 5-HTP, Trp, 5-HT, and MLT assays. The animal care, handing, and sampling procedures were carried out in strict accordance with the protocol approved by the Xinjiang Agricultural University Animal Care and Use Committee.

Diets and feeding

The sheep were housed in an open-sided barn in individual pens (1.0 m × 1.5 m). The powder concentrate was fed at a concentration of 1.0% of the body weight daily and divided into two equal meals at 07:30 and 19:30 h. Hay and fresh water were provided ad libitum. The formulation and nutrition composition of powder concentrate and the nutrition composition of hay are presented in Tables I and II.

Table I. Formulation and nutrition composition of powder concentrate (dry matter basis).

<table>
<thead>
<tr>
<th>Material name</th>
<th>Content (%)</th>
<th>Nutrient name</th>
<th>Content (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Corn</td>
<td>44</td>
<td>DM</td>
<td>91.28</td>
</tr>
<tr>
<td>Soybean meal</td>
<td>20</td>
<td>OM</td>
<td>84.29</td>
</tr>
<tr>
<td>Oat</td>
<td>16</td>
<td>CP</td>
<td>21.82</td>
</tr>
<tr>
<td>Barley</td>
<td>15</td>
<td>NDF</td>
<td>36.24</td>
</tr>
<tr>
<td>CaHPO4</td>
<td>3</td>
<td>ADF</td>
<td>12.97</td>
</tr>
<tr>
<td>Salt</td>
<td>1</td>
<td>Ca</td>
<td>1.54</td>
</tr>
<tr>
<td>Premix</td>
<td>1</td>
<td>P</td>
<td>0.59</td>
</tr>
<tr>
<td>Total</td>
<td>100</td>
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<td></td>
</tr>
</tbody>
</table>

ADF, acid detergent fiber; Ca, calcium; CP, crude protein; DM, dry matter; NDF, neutral detergent fiber; P, phosphorus

Table II. Nutrient composition of corn silage, alfalfa, and wheat straw (dry matter basis).

<table>
<thead>
<tr>
<th>Items</th>
<th>Corn silage</th>
<th>Alfalfa</th>
<th>Wheat straw</th>
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</thead>
<tbody>
<tr>
<td>DM</td>
<td>94.76</td>
<td>93.67</td>
<td>95.93</td>
</tr>
<tr>
<td>OM</td>
<td>84.05</td>
<td>85.30</td>
<td>82.49</td>
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<tr>
<td>CP</td>
<td>8.33</td>
<td>13.72</td>
<td>4.21</td>
</tr>
<tr>
<td>NDF</td>
<td>67.36</td>
<td>53.97</td>
<td>71.31</td>
</tr>
<tr>
<td>ADF</td>
<td>43.73</td>
<td>36.52</td>
<td>47.49</td>
</tr>
<tr>
<td>Ash</td>
<td>10.71</td>
<td>8.37</td>
<td>13.44</td>
</tr>
<tr>
<td>Ca</td>
<td>0.64</td>
<td>1.36</td>
<td>0.38</td>
</tr>
<tr>
<td>P</td>
<td>0.24</td>
<td>0.16</td>
<td>0.07</td>
</tr>
</tbody>
</table>

For abbreviation, see Table I.

Sample analysis

The concentration of 5-HTP in the plasma was
determined as described previously (Enbaek and Magnussen, 1978). The plasma (750 L) was mixed with 4 mL of acidified butanol (4 mL of 70% HClO₄ per liter of 1-butanol) and centrifuged (1960 × g for 15 min at 4°C). Subsequently, the precipitate was removed, the supernatant was mixed with 6 mL of n-hexane, and centrifuged (5600 × g for 20 min at 4°C) to obtain 400 µL of aqueous phase (containing 5-HTP), which was used for column chromatography. The organic phase was discarded. Twenty-five microliters of the extracted plasma was applied to the column and the chromatogram was developed with the column buffer at 70°C with a flow rate of 0.3 mL/min. The effluent from the column was reacted with fluorogen reagent (per liter: 3 mmol phthaldialdehyde, 20 mg FC-134, and 150 mg Brij 35, all in 9 mol/L HCl. This reagent, prepared daily, was maintained in dark) at 70°C for 8 min before measuring the fluorescence.

The plasma Trp level was measured by fluorescence spectrophotometer (SHIMADZU, Japan). The plasma (50-100 µL) was mixed with an equal volume of 0.6 N trichloroacetic acid and centrifuged at 2000 × g for 20 min at 4°C, the precipitate was removed, and the supernatant was diluted 5 times. One hundred microliters of the diluted plasma was dropped into a test tube, then added 0.9 mL distilled water, 2.5 mL 0.6 mol/L trichloroacetic acid solution, 0.2 mL 2% formaldehyde solution and 0.1 mL 6 mmol/L FeCl₃-0.6 N TCA solution, respectively. Mixed the solution and placed a boiling water bath for 90 min. The mixed solution was detected by fluorescence spectrophotometer (Excitation wavelength 304 micrometers, emission wavelength 448 micrometers).

The plasma 5-HT level was determined using an ELISA kit (Beckman-Coulter, Krefeld, Germany). The sample was diluted to 1:100 in order to ensure that it was within the range of the standard curve.

The plasma MLT concentration was measured using a commercial ELISA kit (IBL Hamburg, Germany). The detection limit was 3.0 pg/mL, intra-assay-variation was 7.5%, and inter-assay-variation was 11.3%.

Statistical analyses
The data are presented as mean ± standard deviation, and analyzed by repeated ANOVA test using the SPSS 18.0 statistical software (IBM, State of New York, Armonk USA). The results with P value < 0.05 were considered statistically significant and those with P value < 0.01 were considered significantly different.

RESULTS

Plasma 5-HTP
The supplementation of RPT 5-HTP increased the plasma 5-HTP level in the sheep of both at CT +111 and CT +222 groups (Fig. 1A). A significant increase was observed from 6 h up to 8 h post feeding in the morning (P < 0.05). In the control sheep, the 5-HTP concentration remained unchanged on all experimental days during daytime.

Plasma Trp
There were not differences in the plasma Trp content between the control, CT +111, and CT +222 groups (Fig. 1B). The plasma Trp content did not change significantly when compared with that of the control group after the administration of either 111 mg/kg BW or 222 mg/kg BW RPT 5-HTP (P > 0.05).

Plasma 5-HT
In the control group, the concentration of 5-HT was low during 0–4.5 h post feeding, and then increased to a plateau that lasted the entire activity period (Fig. 1C). There was a significant difference in the level of 5-HT with time in the CT +222 group, with a maximum level at 6 h post feeding. Moreover, the plasma 5-HT content in the CT +222 was significantly higher than that of the control at 3, 6, and 8 h post feeding in the morning (P < 0.05). However, there was no significant difference between the control and CT +111 groups (P > 0.05).

Fig. 1. Plasma 5-hydroxytryptophan (A) tryptophan (B) 5-hydroxytryptamine (C) and melatonin (D) concentrations in the sheep provided diet supplemented with 55.5 mg/kg BW RPT 5-HTP or 111 mg/kg BW RPT 5-HTP at 07:30 h. Control (♦-♦); CT+111(■-■); CT+222(▲-▲). The below abscissa shows the time after feeding. The data are presented as mean ± S.D. The means within a time point without common superscripts were significantly different (P < 0.05). *P < 0.05.
**Plasma MLT**

Because of the intake of diet at 07:30 h, the plasma MLT level increased during 1.5–3 h post feeding in the morning (Fig. 1D). The plasma MLT levels in the sheep of CT + 111 and CT + 222 groups at 1.5 and 4.5 h post feeding were consistently higher than those in the sheep of the control group ($P < 0.05$). Furthermore, there was no significant difference in the plasma MLT level between the CT + 111 and CT + 222 groups ($P > 0.05$).

**DISCUSSION**

Fifteen days pretreatment with RPT 5-HTP (111 or 222 mg/kg BW), produced a significantly increase in the accumulation of plasma 5-HTP. The results of the present study are in accordance with those of Westenberg et al. (1982) and Joseph and Baker (1976), who demonstrated the changes in the serum 5-HTP level in humans after oral administration of 5-HTP. 5-Hydroxytryptophan is well absorbed from oral dose, with approximately 70 percent ending up in the bloodstream (Magnussen et al., 1981; Magnussen and Nielsen-Kudsk, 1980). The absorption of 5-HTP is not affected by the presence of other amino acids; therefore, it can be supplemented with diet in sheep. Unlike Trp, 5-HTP cannot be shunted into niacin or protein production (Birdsall, 1998).

There was no increasing trend of sheep plasma 5-HTP with the increased level of RPT 5-HTP. 5-Hydroxytryptophan is derived from Trp via a pathway in which Trp hydroxylase (TPH) catalyzes the rate limiting step in the synthesis of 5-HTP. Furthermore, TPH uses Fe$^{2+}$ as a cofactor and O$_2$ and tetrahydrobiopterin (BH4) as co-substrates to hydroxylate Trp generating 5-HTP (Mockus and Vrana, 1998). The major metabolic pathway of 5-HTP is decarboxylation to 5-HT. The concentration of L-aromatic amino acid decarboxylase is particularly high in the kidney, liver, and gut (Sourkes, 1987). Orally administered 5-HTP might therefore be metabolized in the gastrointestinal mucosa during absorption and by the liver before the amino acid reaches the systemic circulation (Magnussen et al., 1981). Our finding suggests that a part of administered 5-HTP is metabolized in the liver and gut before entering the bloodstream.

RPT 5-HTP treatment did not significantly change the plasma Trp concentration (Fig. 1B). The intestinal absorption of 5-HTP does not require transport molecules (Birdsall, 1998), whereas Trp is absorbed by amino acid transporters (Broer, 2008a, 2008b). This indicates that 5-HTP absorption is not affected by other amino acids. Despite this, in the present study, the plasma Trp content in the control and treatment sheep increased at 3 h after feeding. The increased plasma Trp might be correlated with diet intake.

A marginal increase in the plasma 5-HT level was observed for a long duration in sheep (Fig. 1C). This suggests that 5-HTP administered is metabolized to 5-HT in the intestine or liver by aromatic amino acid decarboxylase, with the resulting product entering the blood circulation. Studies have reported contradicting results with respect to the blood 5-HT level after treatment with 5-HTP. Wa et al. (1995) reported significant increases in urinary 5-HTP and 5-HT excretion, and no significant changes in the blood 5-HT level, whereas, Kaneko et al. (1979) and Takahashi et al. (1976) have reported a significant increase in the blood 5-HT level after the oral administration of 5-HTP. However, there have been a few suggestions regarding a positive relationship between 5-HTP intake and blood 5-HT in ewes.

MLT plasma level was affected by RPT 5-HTP supplementation (111 or 222 mg/kg BW) in sheep (Fig. 1D). We believe that the marked elevation in the plasma MLT level by 5-HTP administration is due to a mass-action effect on the 5-HTP-MLT pathway in the pineal gland (Sugden et al., 1985). The three enzymatic reactions decarboxylation, N-acetylation, and O-methylation do not seem to be saturated during daytime. This is supported by three lines of evidence: 1) an increase in the concentration of 5-HT in the plasma has been observed in sheep treated with RPT 5-HTP (Fig. 1C); 2) monoamine oxidase inhibitors, which elevate 5-HT level, also increase the pineal MLT level in rat (Namboodiri et al., 1983); 3) during daytime, 5-HTP increases the level of pineal gland MLT in rat in vivo (Wurzburger et al., 1976); 4) the treatment with 5-HTP (200 mg/kg BW) elevates the pineal 5-HT, N-acetylserotonin, and MLT levels in sheep (Sugden et al., 1985). Therefore, the animals receiving RPT 5-HTP presented higher plasma concentrations of MLT during daytime, indicating the transfer of MLT from the pineal gland to the peripheral blood circulation. However, there was no dose-effect relationship between 5-HTP intake and plasma MLT level (Fig. 1D). This might depend on whether there is saturation of the enzyme during the conversion of 5-HTP to MLT, or whether there is also saturation when the synthesized MLT in pineal gland is transported to the blood circulation.

RPT 5-HTP has a significant time effect in regulating plasma 5-HTP, 5-HT and MLT in sheep. The concentration of Trp in the plasma showed negligible change at 1.5–3 h post feeding. The level of 5-HTP increased initially, and then decreased, whereas, the 5-HT content decreased initially, and then increased, suggesting a contrary tendency with the prolonging of feeding time. The 5-HT level decreased marginally. McKinney et al. (2005)
revealed that the $K_m$ value of TPH in sheep is 22 $\mu$mol/L, and at this point, the tryptophan level in the blood is 38 $\mu$mol/L. It is suggested that a part of tryptophan in diet is converted to 5-HTP, which can be converted to 5-HT and MLT. However, there is no obvious change in the content of Trp, 5-HTP, 5-HT, and MLT at 4.5–12 h post feeding, which indicates that the influence of RPT 5-HTP on these components is clearly seen from 0–4.5 h post feeding.

**CONCLUSIONS**

The present study demonstrated that the plasma 5-HTP and MLT concentrations increased at 3 h post feeding, and then presented a fluctuating trend. Dietary RPT 5-HTP can elevate the plasma 5-HTP, 5-HT, and MLT concentration, but 111 mg/kg BW RPT 5-HTP did not increase the plasma 5-HT concentration. The trend of changes in the plasma 5-HTP level was consistent with that of the plasma 5-HT level when treated with RPT 5-HTP. The plasma Trp level was not affected by RPT 5-HTP.

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Statement of conflict of interest
The authors have declared no conflict of interest.

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