Expression of the ZFX Gene in Mouse Kidney During Postnatal Development

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ABSTRACT
The relationship of cell proliferation and apoptosis is important in normal kidney development. The ZFX gene is important for regulation of growth, proliferation and differentiation in tissue cells while the Bcl-2 and BAX genes regulate apoptosis. This experiment adopted the real-time fluorescence quantitative technique to test ZFX, Bcl-2, and BAX expression in mice kidney tissue on 1 d, 7 d, 14 d, 21 d, 42 d, and 90 d after birth. The results showed that with the maturation of mouse kidney, the cell proliferation activity decreased, and the expression of ZFX, Bcl-2, and BAX were down regulated. The ZFX gene may participate in the developmental process of the kidney through the balance of cell proliferation and apoptosis regulation.

INTRODUCTION
Zinc finger protein X-linked (ZFX) is a member of the zinc finger protein super family and contains 13 C2H2-type zinc finger structures. The ZFX gene is on the x chromosome in mammals, and its expression is highly conservative. The structure of ZFX contains an acidic transcriptional activation domain (AD), a nuclear localization sequence (NLS), and a DNA binding domain (DBD). Early research suggested the ZFX gene was associated with sex determination of mammals (Schneider-Gadicke et al., 1989). In recent years, research documented that ZFX plays an important role in self-renewal and anti-apoptotic mechanisms of embryonic stem cells, adult hematopoietic stem cells, lymphoid cells and several other types of stem cells (Galan-Caridad et al., 2007; Arenzana et al., 2009). Considering the characteristics of the tumor and stem cell, oncology studies found ZFX was abnormally expressed in laryngeal carcinoma’s, glioma’s, gastric cancer, prostate cancer, breast cancer and other common human malignancies, and it may also be involved in the regulation of tumor cell proliferation and anti-apoptotic processes (Jiang et al., 2012; Nikpour et al., 2012; Zhou et al., 2011). Since the zinc finger protein is a transcription factor which has a finger-like domain, it can combine with the promoter region thereby regulating gene expression, and also plays an important role in the growth, proliferation and differentiation of tissue cells (Chandrasekharan et al., 2009; Schnidar et al., 2009). There are two categories of apoptosis regulatory genes which contain apoptosis inhibition genes and apoptosis promotion genes. Bcl-2 is the most important anti-apoptotic gene while BAX is a major pro-apoptotic gene (Kallio et al., 2004; Yamamoto et al., 2004).

The development of the mammalian kidney includes stages of pronephros, mesonephros and metanephros, and the three stages are consecutive in the timing and location (Lipschutz, 1998; Davies, 2002). The appearance of mouse pronephros is at embryonic day 8 (E8), mesonephros is initiated at E9.5, and metanephros begins at E11 with the appearance of a small epithelial buds or diverticulum from the lower end of the mesonephric duct near its entry into the cloaca; the completion of the number of glomeruli in mice is on day 7 postpartum, with completion of kidney medulla on day 21, and by day 42 the structure of kidney is the same as the adult (Nyengaard, 1993).

Real-time, reverse transcription quantitative PCR (RT-qPCR) is a highly sensitive and reproducible technology for the analysis of gene expression patterns. In this study, by quantitative real-time technique ZFX, Bcl-2 and BAX expression at different stages of kidney development in mice after birth were evaluated, to determine the expression of ZFX during kidney development.
Table I. Conditions of PCR and oligonucleotide primer pairs.

<table>
<thead>
<tr>
<th>Target gene</th>
<th>GenBank No.</th>
<th>Sequence of primer (5'-3')</th>
<th>Annealing temp.</th>
<th>Products</th>
</tr>
</thead>
<tbody>
<tr>
<td>ZFX</td>
<td>NM_011768.2</td>
<td>F: CAGATCCGTGGTAAGTGAAGTCGC</td>
<td>61°C</td>
<td>147 bp</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R: ACTGATACTTTCTCGGGCACT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bcl-2</td>
<td>NM_011768.2</td>
<td>F: GTCCGCTACCTGTGTGACCTTC</td>
<td>61°C</td>
<td>284 bp</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R: CAGACATGCACCTACCCAGC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BAX</td>
<td>NM_007527.3</td>
<td>F: CGCGAATTGGAGATGAACCTG</td>
<td>61°C</td>
<td>161 bp</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R: GCCAAGTAGAAGAGGGCAACC</td>
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</tr>
<tr>
<td>GAPDH</td>
<td>NM_008084.2</td>
<td>F: ACCCAAGAGACTGTGGATAGG</td>
<td>61°C</td>
<td>171 bp</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R: CACATTGGGGGTAGGAACAC</td>
<td></td>
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</tr>
</tbody>
</table>

MATERIALS AND METHODS

Animals
Clean adult mice were obtained from the Experimental Animal Center, Shihezi University. Mice were housed in clean, controlled environment, with food and water available ad libitum. Room conditions included maintenance of temperature at 24°C, humidity at 50%, with lighting from 8:00-20:00. After adaptation for 7 days, mice were paired and bred. The next morning females with vaginal plugs were separated for production of experimental litters. Kidneys of progeny aged 1, 7, 14, 21, 42 and 90 days were removed immediately after the animals were killed by cervical dislocation. The tissues stored in liquid nitrogen for reverse transcription-quantitative real-time polymerase chain reaction (RT-qPCR).

Primary reagents
Trizol was purchased from Invitrogen Life Technologies, Carlsbad, CA, USA, reverse transcription kits were purchased from Takara Co., Dalian, China, diethyl pyrocarbonat was purchased from Sigma (Santa Clara, CA, USA), agarose was purchased from GeneTech Co., Shanghai, China, DNA Marker was purchased from TianGen Co., Beijing, China, SYBR® Premix Ex Taq™ (TaKaRa), 96-well plates and cover films were purchased from Applied Biosystems Inc., CA, USA and SYBR Green SuperMix was acquired from Qiagen, Valencia, CA, USA.

RNA extraction and reverse transcription
Total RNA was extracted from kidneys cryopreserved with liquid nitrogen using Trizol following the manufacturer’s instructions. Reverse transcription of RNA was carried out using the gDNA Eraser PrimerScript RT reagent kit according to the manufacturer’s instructions.

Primer design
According to the principles of primer design (Rodriguez et al., 2015; Livak et al., 2001), Primer Premier 5.0 software was applied to design the primer sequences of ZFX, BAX, Bcl-2, and GAPDH. The primer sequences was synthesized by BGI (Table I).

Conventional PCR assays
To detect primer specificity, and the quality of the template, PCR was conducted in advance. The amplification system was as follows: cDNA 1 μl, PCR reaction mixture (Mix) 10μl, upstream and downstream of each primer, 0.5 μl, ddH₂O 8μl. Reaction conditions were: 94°C denaturation for 5 min; 94°C denaturation 30 s, 61°C annealing 30 s, 72°C extension 50 s, 30 cycles; 72°C extension 10 min. Finally, 10 μl of PCR products were run on 2.0% agarose gel, and then analyzed by a gel imager analysis system (need to specify the system).

RT-qPCR
Reverse transcription-quantitative real-time polymerase chain reaction (RT-qPCR) was conducted using a LightCycler 2.0 instrument (Roche, Basel, Switzerland), with SYBR Green SuperMix, following the manufacturer’s instructions. Each sample was tested three times to insure accuracy. The amplification system was as follows: cDNA 2 μl, SYBR@ Premix Ex Taq™ (2×) mixture (Mix) 10 μl, upstream and downstream of each primer, 0.4 μl, sterile distilled water to 20 μl. Reaction conditions were: initial denaturation step (30 s at 95°C), 45 cycles of 95°C for 10 s, 61°C for 30 s, 72°C extension 10 min. Finally, 10 μl of PCR products were run on 2.0% agarose gel, and then analyzed by a gel imager analysis system (need to specify the system).

Statistical analysis
Numerical analysis was done using the 2-ΔΔCT method using the Gapdh gene as an internal standard. Data were analyzed using SPSS 17.0 (city, state, USA). Tests of hypotheses were done using t-tests with P<0.05 considered statistically significant.

RESULTS

Amplified products of ZFX, Bcl2, BAX and GAPDH
The cDNA were amplified with ZFX, Bcl-2, BAX and GAPDH primers (Table I) to obtain 147 bp, 284 bp, 161 bp
and 171 bp products (Figs. 1, 2), respectively. According to the results of the gel imaging system, primers had excellent specificity and the qualities of the templates were reliable.

**Fig. 1. PCR amplification of ZFX and Bcl-2.**

**Fig. 2. PCR amplification of BAX and GAPDH.**

**Expression analysis of ZFX**

RT-qPCR was performed to determine if ZFX mRNA displayed an age-dependent expression pattern and results indicated an age-dependent change of ZFX mRNA expression in the postnatal mouse kidney. ZFX mRNA expression was greatest on the day 1, and then dropped gradually from day 1 through day 21 with the least expression on day 21 (P<0.01). Day 42 had similar expression to day 21 (P>0.05) while day 90 was slightly greater (P < 0.05) (Fig. 3A).

**Expression analysis of Bcl-2**

The mRNA level of Bcl-2 also displayed an age-dependent expression pattern. On day 1, the expression of Bcl-2 was greatest, but lesser on days 7-90 (P<0.01). The least expression occurred day 21 compared to days 1-14 (P < 0.01) with days 42 and 90 similar to day 21 (P > 0.05) (Fig. 3B).

**Expression analysis of BAX**

The mRNA level of BAX in different periods of postnatal mouse kidney showed a trend similar to Bcl-2. Expression of BAX was greatest on day 1, and then declined on day 7 (P<0.01) and day 14 (P<0.01) until reached the least expression on day 21 (P<0.01). Expression of BAX increased slightly on day 42 (P < 0.05) but expression on day 90 was similar to day 21 (P>0.05) (Fig. 3C).

![Expression analysis of ZFX, Bcl-2, and BAX](image_url)

**DISCUSSION**

Zinc finger protein plays an important role in the proliferation and differentiation of cells. The ZFX gene products are necessary for maintaining embryonic stem cells and adult stem cell self-renewal, and are a common molecular basis of embryonic stem cells development and adult stem cell regeneration (Cellot et al., 2007; Ouyang et al., 2009). Inactivation of the ZFX gene has been shown to cause increased mortality of neonatal mice and decreased reproductive capacity of surviving maternal mice (Luoh et al., 2009).
Cell proliferation and apoptosis in mouse kidney development is regulated by a number of genes (Terzi et al., 2000) and normal growth and development involves a balance relationship of cell proliferation and apoptosis. On day 1 of kidney development, the cell proliferation index of medullary collecting duct and renal tubular tissue was greater than the apoptosis index, with proliferation dominant. On day 7, cell apoptosis was dominant and cells begin to differentiate into mature cells. On day 14, the cell number and size of renal tubular and collecting duct tissue were not predominant with proliferation and apoptosis both at lesser levels. On day 21, the renal cortex and medulla were thickened with the development of the inner medulla, and cell proliferation decreased. At 42 days after birth, the renal cortex and medulla were thickened with the development of the inner medulla, and cell proliferation and apoptosis were in a relatively balanced state (Li et al., 2006).

The Bcl-2 and BAX genes belong to the Bcl-2 family. The Bcl-2 gene is an anti-apoptotic gene and BAX is a pro-apoptotic gene, and are important genes in regulating cells apoptosis. The mechanism of Bcl-2 is through production of antioxidant, inhibiting the release of pro-apoptotic protein and inhibiting the cytotoxicity of BAX to achieve an anti-apoptotic effect. The BAX gene can form homodimers, damaging mitochondrial membranes, and the appearance of apotosome in the cytoplasm, activating Caspase cascade activation, triggers the fragmentation of DNA and damage to a variety of cellular proteins components, which accelerates cell death (Saikumar et al., 1998; Yang et al., 1997). During postnatal kidney development Bcl-2 and BAX are both necessary for regulating cells apoptosis, and Bcl-2 deficient mice demonstrate polycystic kidneys (Veis et al., 1993).

SYBR Green reverse transcriptase quantitative PCR (RT-qPCR) is the most widely method used to investigate gene expression in ruminants. The increase in the fluorescent signal is directly proportional to the number of PCR product molecules generated. It possible to simultaneously characterize different genes, using small quantities of sample with high specificity, sensitivity and accuracy (Giulietti et al., 2001; Bustin, 2000).

In our study ZFX, Bcl-2, and BAX mRNA was confirmed to display an age-dependent expression pattern by RT-qPCR, mRNA expression figures were done to determine: (i) the relationship between the mRNA expression of ZFX, Bcl-2 and BAX; and (ii) whether ZFX mRNA age-dependent expression tendency was consistent with those of Bcl-2 and BAX. Results showed that mRNA expression of the ZFX, Bcl-2, and BAX genes was detected in all stages of postnatal kidney development and displayed similar age-dependent changes. On day 1 of early development of medulla ZFX was strongly expressed, and with the maturation in medulla development, ZFX expression decreased gradually until 21 days when the medulla was fully developed and the expression of ZFX reached the minimum. While the expression trend of Bcl-2 and Bax during the development of postnatal kidney showed the cell proliferation and apoptosis were have the same patterns which were consistent with Zang et al. (2007). These results suggested that during the development of postnatal kidney, ZFX might ensure cell proliferation and viability, to promote cell further maturation. However, after day 42 when the kidney is mature, ZFX expression was increased so that renal cell proliferation and apoptosis were in relative equilibrium.

It was reported that ZFX was significantly up-regulated in renal cell carcinoma (RCC) tissues (Li et al., 2015), and knockdown of ZFX suppresses renal carcinoma cell growth and induces apoptosis (Fang et al., 2014). In addition congenital anomalies of the kidney and urinary tract (CAKUT): cystic kidney disease is the first primary cause of the end-stage renal disease (ESRD) in China’s children (Tang et al., 2014). According the expression pattern of ZFX in the postnatal kidney in our research, ZFX may be involved in the process of developing renal cell proliferation and balanced regulation of apoptosis.

**CONCLUSION**

In this study, ZFX gene expression in the different maturation state of mouse kidney was tested using the real-time fluorescence quantitative technique, meanwhile Bcl-2, and BAX expression was tested. Through the comparison, it showed the ZFX gene may participate in the developmental process of the kidney through the balance of cell proliferation and apoptosis regulation. However, the role of ZFX in the balance of cell proliferation and apoptosis in kidney development needs further protein test to confirm or refute this hypothesis ,and it maybe provide some information to research on the end-stage renal disease.

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**Statement of conflict of interest**

Authors have declared no conflict of interest.
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