

Research Article



Mannheimiosis in A Rusa Deer (*Rusa timorensis*): A Case Report and A Herd Analysis

WAN-NOR FITRI¹, SALLEH ANNAS*², MUHAMMAD RASHID AZROLHARITH³, FAEZ FIRDAUS ABDULLAH JESSE¹, ZAMRI-SAAD MOHD², WAHID HARON¹

¹Department of Veterinary Clinical Studies; ²Department of Veterinary Laboratory Diagnostics; ³Department of Medicine and Surgery of Farm and Exotic Animals, Faculty of Veterinary Medicine, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia.

Abstract | Mannheimiosis and pasteurellosis is an important respiratory tract disease in cattle, sheep, goats and other ruminants. This article describes the first case of pneumonic and septicaemic mannheimiosis and pasteurellosis in a herd of *Rusa timorensis* and *Dama dama* deer. A *Rusa timorensis* stag showed constant reduction in body weight and was found dead on the 30th May 2016. Severe pulmonary hepatisation and multiple pulmonary abscessation along with emaciation and evidences of traumatic injuries were observed at post-mortem. Diagnosis of pneumonic and septicaemic mannheimiosis with concurrent colisepticaemia was made based on the bacterial culture. A retrospective study on deer from the same herd revealed occurrence of respiratory infection leading to death. Review of other cases from the animal from the same herd revealed that respiratory infections is a problem within this particular herd with transportation, bullying, poor nutrition and inclement weather being some of the important stress factors leading to respiratory infection. *Pasteurella multocida* and *Mannheimia haemolytica* were the two main important organisms in these cases, along with concurrent infection by *Escherichia coli* and *Burkholderia pseudomallei*. Other stress factors such as occurrence of El Niño and natural behaviour such as rutting in deer leading to manifestation of the disease in the herd were highlighted. Potential approaches to manage stress with the interest of rutting season were also discussed. This report is the first reported case of septicaemic mannheimiosis in a deer.

Keywords | Colisepticaemia, Deer, Mannheimiosis, Pasteurellosis, *Rusa timorensis*

Editor | Muhammad Abubakar, National Veterinary Laboratories, Islamabad, Pakistan.

Received | March 02, 2017; **Accepted** | March 24, 2017; **Published** | March 26, 2017

***Correspondence** | Annas Bin Salleh, Department of Veterinary Laboratory Diagnosis, Faculty of Veterinary Medicine, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia; **Email:** annas@upm.edu.my

Citation | Fitri WN, Annas S, Axrolharith MR, Jesse FFA, Mohd ZS, Haron W (2017). Mannheimiosis in A Rusa Deer (*Rusa Timorensis*): A Case Report and A Herd Analysis. Res. J. Vet. Pract. 5(1): 5-11.

DOI | <http://dx.doi.org/10.17582/journal.rjvp/2017/5.1.5.11>

ISSN (Online) | 2308-2798; **ISSN (Print)** | 2309-3331

Copyright © 2017 Fitri. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Mannheimiosis is an important respiratory tract disease in cattle, sheep, goats and other ruminants (Highlander, 2001). Other than mannheimiosis, pasteurellosis is also known as an important respiratory disease in livestock. Pasteurellosis is caused by *Pasteurella multocida*, which is a genus closely related to mannheimia. Mannheimiosis in animals is known to be caused by *Mannheimia haemolytica* of specific serotype A. The bacterium of this genus are known to be normal flora of many domestic and wild ani-

mals, including dog, cat, sheep, goat, and cattle (Highlander, 2001). Although the disease is seemingly prevalence in many species of animal, the nature of disease in *Rusa timorensis* is unknown and has yet to be reported. This report is an evidence that the disease is prevalence in deer herd in Malaysia and the disease should be closely monitored.

Since the bacterium is a normal flora, one or more factors could upset this balance. Like most primary respiratory tract disease, stress accounts a substantial factor in the disease progression (Annas et al., 2014; Mohamed and

Abdelsalam, 2008). Similarly, stress factor was taken into consideration in this case. Stressors that have clearly been identified in this report was transport and environmental stress among others. But apart from that, this study also report a potential physiologically induced stressor uniquely found in seasonal breeder. Breeding season, or also known as rutting season is an annual affair of stags competing for the right to mate with the hind. This includes agonistic and aggressive behavior such as sparring, antler fights, chasing and groaning between stags. A recent study found that the rutting season in deer is stressful to the stags (Pavitt et al., 2015). Thus, rutting season could be an important factor in the epidemiology of respiratory tract disease in seasonal breeder such as deer.

Mannheimiosis has been well documented in both sheep and goats, resulting in pneumonic or septicemic manifestation in sheep, goats and larger ruminants (Misra et al., 1970; Dyer and Ward, 1998; Foreyt et al., 1994). Yet only few number of cases of mannheimiosis in deer has been reported (Hattel et al., 2004), none involving *Rusa timorensis*. The disease could have been under-reported in deer since it is a semi-domesticated species. In semi-domesticated animal, behaviour is not readily observed while the sick often conceal themselves. Thus the disease could have easily been overlooked especially in a dynamic disease such as mannheimiosis. Though there has been no study on the financial burden of mannheimiosis, the disease has proven to be economically significant to domestic animal with significant mortality rate to calf (Mohamed and Abdelsalam, 2008). This present article describes the progression of mannheimiosis in *R. timorensis* – exploring the possible stressors in the particular animal as well as describing the necropsy findings of the case. Subsequently, a retrospective case review from sympatric deer species is discussed.

MATERIALS AND METHODS

ANIMAL

A *R. timorensis* stag from Small Ruminant Unit (2.99°N, 101.7°E), Taman Pertanian Universiti (TPU), Universiti Putra Malaysia (UPM) was the primary subject of the present case. This stag (ID: T521) was part of a group of deer being transferred from Dewan Bandaraya Kuala Lumpur (DBKL) to TPU. The translocation took place in 25th March 2015, involving 45 deer (8 male *R. timorensis*, 20 female *R. timorensis*, 5 male *Dama dama*, and 12 female *D. dama*). Note that no previous record was available on all animals involved. At the TPU facilities, the deer were managed by extensive system; rotated in a paddock of approximately 10,640 m² each paddock, allowed to graze on guinea grass with palm kernel cake supplementation once every two days and drinking water were provided *ad li-*

bitum. All deer in TPU were subjected to monthly body weight monitoring, performed in a dark room.

POST-MORTEM EXAMINATION

T521 was found dead on 30th May 2016 and necropsy was immediately conducted at the Post-Mortem Laboratory, Faculty of Veterinary Medicine, University Putra Malaysia (UPM). Gross lesions of various organs were detected, appropriate samples collected and subsequently sent to Bacteriology Laboratory, Faculty of Veterinary Medicine, UPM for bacterial isolation. Organs were also fixed in 10% neutral-buffered formalin, sent to Histopathology Laboratory, Faculty of Veterinary Medicine, UPM and were subjected to routine processing, stained with haematoxylin and eosin (H and E).

RETROSPECTIVE STUDY

A retrospective study was conducted by screening through the necropsy record from March 2015 to October 2016. Only data of cases involving the death of deer translocated from DBKL to TPU were included in this report. Data includes time of death, stress factors, bacterial isolation, gross lesions examination, histopathologic lesions examination, and diagnosis.

RESULTS AND DISCUSSIONS

T521 was observed to be the most dominant in the herd. The individual, T521 participated in rutting (establishment of hierarchy) and actively maintain the status in the herd by competing and restricting other individual stags from feeding through aggression. Prior to its death, its behavior was dull, depressed and isolation from the herd.

BODY WEIGHT MONITORING

Body weight monitoring of T521 revealed slight but consistent reduction of body weight from February to March (Figure 1). Remarkable body weight reduction was observed between (April to mid May 2016), during which the antler drop occurred.

GROSS LESIONS EXAMINATION

At necropsy, multifocal areas of alopecia was noted at the left flank of the animal, with circular contusions with diameter of approximately 7 cm at the left dorsal part of thorax cranial to the diaphragm border. Poor hair coat was noted, with dehydration as evidenced by sunken eyes. Ribs and prominent body protuberances were noted, suggesting very poor body condition score of the animal at 1/5 score (Audige et al., 1998). Subcutaneous fat was minimal, with most areas showing serous atrophy of the fat. The general musculature was moderately reddened.

Examination of the respiratory tract revealed bilateral, dry, severe pulmonary hepatitis at the cranioventral



Figure 1: T521 body weight (kg) from February to May 2016. Note the severe reduction of body weight from 4th May 2016 with 30th May 2016

aspect of the lungs (Figure 2). Multiple firm nodules were palpated at the affected areas of the lungs. Gross sectioning of the lungs revealed multiple pulmonary abscesses of various sizes, and various thickness of fibrous tissues capsules containing whitish caseation (Figure 3). Prominent thickening of interlobular septa was also noted. The other part of the lungs showed generalised, mild to moderate pulmonary congestion.

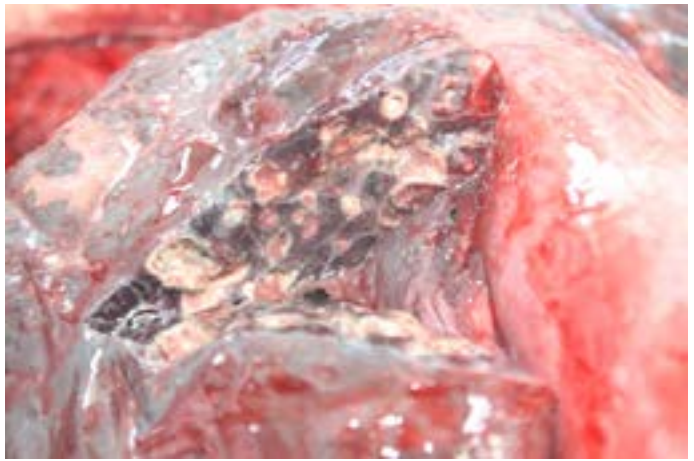


Figure 3: Multiple pulmonary abscesses with thickening of interlobular septa

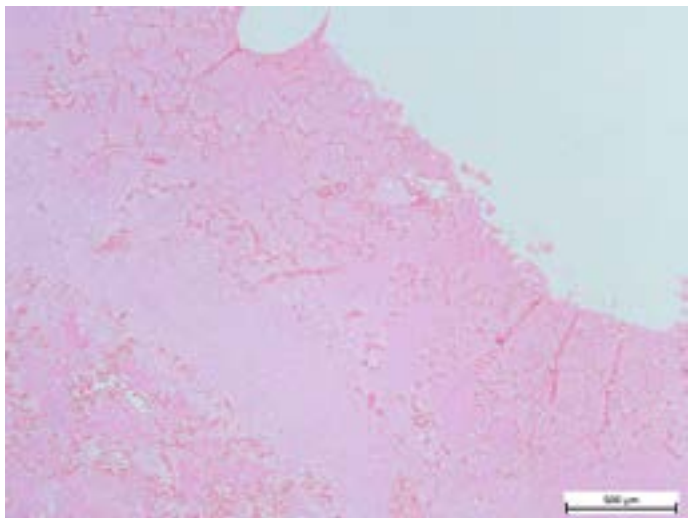


Figure 4: Fibrous tissue capsule in the lungs, with congested remnant of alveoli. (H and E, bar = 500µm)

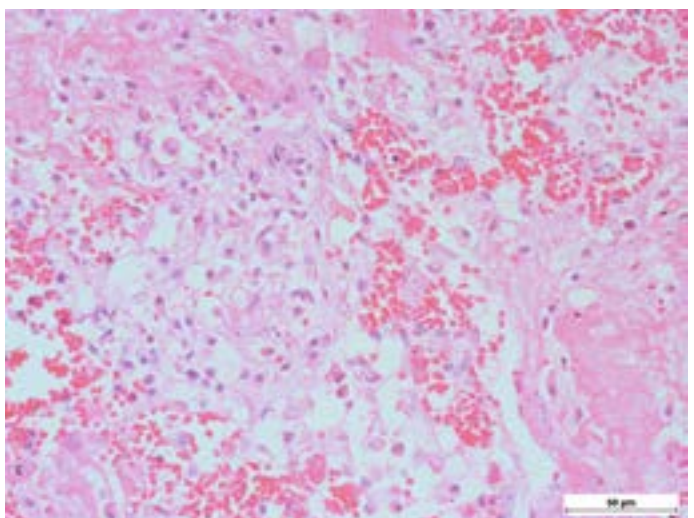


Figure 5: Presence of alveolar macrophages and neutrophils, intermixed with fibrin along with haemorrhage in the lungs

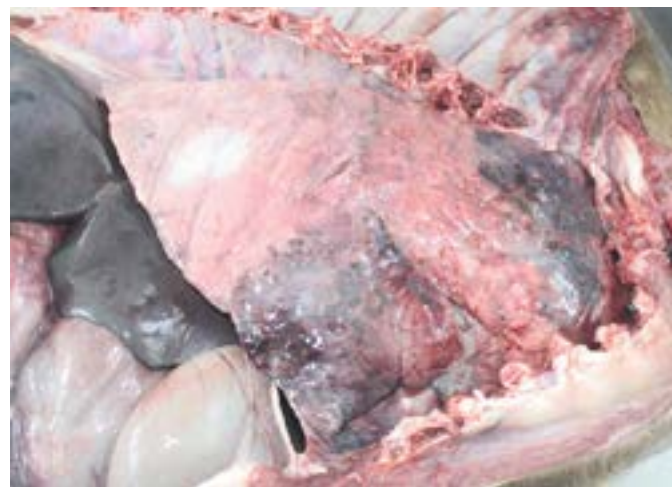


Figure 2: Severe pulmonary hepatisation at the cranioventral aspect of the lungs

In the gastrointestinal tract, foreign body concretion made up of plastic bags was found in the rumen. Severe congestion of the blood vessels, accompanied by haemorrhages was noted at the the serosa surface of the stomach. Multi-focal areas of congestion and petechiation was also noted at the small intestinal serosa. No lesion was noted in the small intestinal mucosa.

In the kidneys, bilateral, generalised, severe congestion was observed. Severe splenic congestion was also observed.

HISTOPATHOLOGIC LESIONS EXAMINATION

Histopathologic examination of the lungs showed multiple areas of nodules, surrounded by fibrous tissues of various thickness. Remnant of alveoli may be observable inside

the nodules, adjacent to the fibrous capsule (Figure 4). The content of the nodules were mainly caseous, with mixture of proteinaceous fluid, and degenerated and necrotic macrophages and neutrophils intermixed within fibrin matrix (Figure 5). These findings were consistent with chronic infection of the respiratory tract. Purplish bacterial colonies were frequently observed in the alveolar spaces. The interalveolar septa of the capillaries was observed to be moderately congested, with thickening of the interalveolar septa due to this congestion. Disseminated intravascular coagulopathy (DIC) was noted mainly in the small capillaries of the lungs. Moderate to severe pulmonary haemorrhage was occasionally observed. These findings were suggestive of acute septicaemia.

In the liver, there was mild disarrangement of architecture of the liver. Generally, most hepatocytes were either swollen or shrunken, with some individual hepatocytes showing cytoplasmic eosinophilia suggesting of severe degeneration (Figure 6).

Severe congestion was noted in the kidneys, with the presence of DIC.

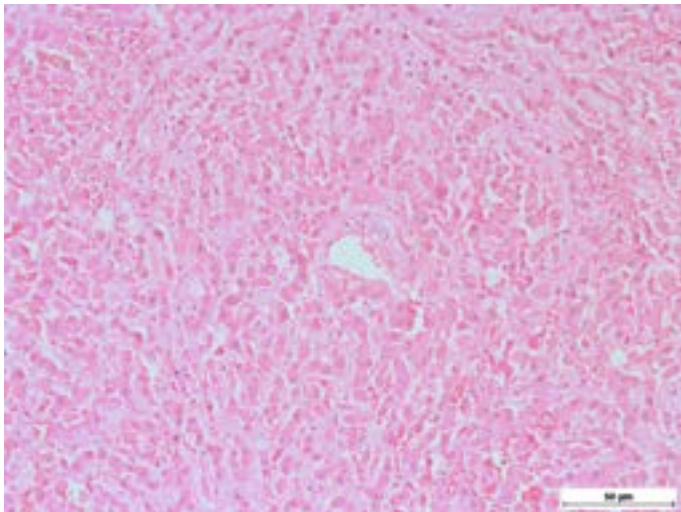


Figure 6: Swollen and shrinking of hepatocytes, with occasional cytoplasmic eosinophilia, Note that the sinusoids were widened

BACTERIAL ISOLATION

Concurrent septicemia was suspected on post-mortem examination. Thus, lungs, liver and kidneys samples were collected and sent for bacterial isolation. *M. hemolytica* and *Escherichia coli* were isolated from the lungs, liver, and kidneys of T521.

DIAGNOSIS

Based on the laboratory findings, the final diagnosis made was pneumonic and septicaemic manheimiosis with concurrent colisepticaemia.

RETROSPECTIVE STUDY

Time of Death: Earliest time of death occur at 65 days

post translocation from the previous facility, involving animal D502. Subsequently, on 76 days post-translocation, another animal died (D507). No death due to pasteurellosis was recorded for the next 357 days, until another deer from the same herd died on day 432 post-translocation (T521). This is then followed by a deer succumbed at 514 days post-translocation (missing ID).

Possible stress factors: Based on records, several possible factors leading to pasteurellosis in the individual deer were noted. Animal D502 was observed to be involved in transportation stress, translocation to quarantine, being bullied, and suffered from traumatic injuries. Animal D507 was also experiencing transportation stress, and being bullied especially during feeding time thus leading to malnutrition. Animal T521 was observed to be bullied after antler casting, suffered from poor nutrition due to poor quality grass, and inclement weather. The animal with missing ID was observed to be bullied.

Bacterial isolation: Bacterial isolation from various vital organs of the animals revealed that *P. multocida*, *E. coli*, *M. haemolytica*, and *Burkholderia pseudomallei* were isolated in various organs.

In animal D502, *P. multocida*, and *E. coli* were isolated from the lungs, and *B. pseudomallei* was isolated from the spleen and kidneys. In T507, *P. multocida* was isolated from the lungs, *E. coli* was isolated from the lungs and liver, and *B. pseudomallei* isolated from the liver and kidneys.

Gross Lesions Examination: In animal D502, severe dehydration, abrasion on the right antler, generalised musculature congestion, severe generalised pulmonary congestion, multifocal anthracosis and emphysema, generalised congestion of gastrointestinal tract (most severe in the abomasum), foreign body concretion (fabric) and phytobezoar in the rumen, bilateral moderate renal congestion, left cortico-medullary renal granulomas, and multiple splenic abscess were observed. On the other hand, in animal T507, the carcass was emaciated, with generalised severe pulmonary congestion with bronchoalveolar pattern pneumonia, presence of foreign body concretion (fabric) and multiple tricobezoar in the rumen, tearing and severe haemorrhage at the reticulo-omasal junction, empty intestine, and multiple hepatic and renal abscessation. In animal with missing ID, the carcass was autolysed.

Histopathologic Lesions Examination: In animal D502, severe pulmonary atelectasis, DIC, generalised severe suppurative pneumonia, suppurative nephritis with multifocal renal fibrosis and abscessation, splenic haemosiderosis, with bacterial colonies, splenic fibrosis and splenic abscess were observed. On the other hand, in T507, generalised severe suppurative pneumonia, severe pulmonary congestion,

severe haemorrhagic reticulitis and omasitis were noted.

Diagnosis: D502 was diagnosed with pneumonic pasteurellosis, with concurrent melioidosis and septicaemia. T507 was diagnosed with pneumonic pasteurellosis, with concurrent melioidosis, colisepticaemia, and reticulo-omasal perforation secondary to trauma.

This article is the first report of pneumonic and septicaemic mannheimiosis and pasteurellosis in a herd of *Rusa timorensis* and *Dama dama* deer in Malaysia. Cases of pneumonic mannheimiosis and septicaemic pasteurellosis has been previously described in *D. dama* (Carrigan et al., 1991), but no cases of septicaemic mannheimiosis has been described in *R. timorensis* or *D. dama*. Typically in domestic ruminants, mannheimiosis is only restricted to pulmonary manifestation, while pasteurellosis may lead to pneumonic or septicaemic manifestation. In this case, the concurrent infection by *E. coli* may contribute to the occurrence of septicaemia by *M. haemolytica*.

In cases of respiratory problems in a herd of deer, diseases of nutritional or infectious in nature should be considered in the list of differential diagnosis. This nutritional diseases are atypical interstitial pneumonia, and bracken fern toxicosis (Bell, 2008). On the other hand, the contagious diseases are bovine corona virus (BCoV) infection (Saif, 2010), bluetongue, pneumonic mannheimiosis, pneumonic pasteurellosis, and pneumonic mycoplasmosis (Donal, 2015). Multiple infections by these agents may be termed as shipping fever. In this herd, atypical interstitial pneumonia was ruled out due to no observation of the classical hyaline membrane formation in the histopathological examination of the lungs. Bracken fern does not exist in the compound of TPU, thus it was ruled out. BCoV was never reported in Malaysia. BCoV and bluetongue were ruled out because the observed neutrophils (suppurative) infiltrations was suggestive of bacterial infections. Thus, final diagnosis were made based on bacterial culture, which yielded *M. haemolytica*, and *P. multocida* along with *E. coli* and *B. pseudomallei*.

Apart from detailed description of clinical and pathological aspect of mannheimiosis in a deer, this article highlights the occurrence of pasteurellosis in a herd of deer recently being exposed to episodes of stressors. In this particular herd, the possible stress factors include transportation, introduction into new environment, inclement weather, feeding changes, underlying disease, and hierarchical relationship within the herd (Chung et al., 2015; Zamri-Saad et al., 1994). In small and large ruminants, it has been shown that stress highly contributes to the progression of mannheimiosis and pasteurellosis (Zamri-Saad et al., 1994; Straus et al., 1998). Severe, multiple stress factors may lead to severe immunosuppression which eventu-

ally leads to rapid proliferation of the respiratory tract normal flora to cause disease to animals (Grey and Thomson, 1971; Highlander, 2001). In this case, transportation and new environment stress occurred since the animals were recently being translocated from DBKL facility to TPU facility in March 2015, leading to the death of 2 deer in 29th June and 9th May 2015. Inclement weather was apparent especially in 2016 in which extreme temperature related to El Niño was reported (Fatimah et al., 2016). This inclement weather also affect the pasture quality, which in turn leads to poor nutrition to the herd. Underlying disease was evidenced by the isolation of *B. pseudomallei* in various organs of 2 deer.

Stress in stags is apparent during the rutting season and is important for the manifestation of pasteurellosis or mannheimiosis in our case. Previous study had elaborated the activities of *R. timorensis* rutting behaviour (Mourik, 1985). This behaviour is described as a mating ritual that includes fighting between stags to obtain the dominance in the hierarchy. The status or rank is important for stags to have access to mate with hinds. The pre-rutting behaviour starts earlier during the velvet stage of the antler where the stags start challenging each other. Once the antler is fully harden, the sparring turns into full-blown antler fight to obtain the top rank in the herd. The rank of stags is dynamic, with subordinate male constantly trying to mate with hinds despite the presence of dominance stags. Hence, the dominant stag always have to be in an alert state to guard the hinds from rival stags. Once hinds expressed signs of estrous, the stags will once again compete to mate with an extended mating ritual; kissing and kicking, mounting, intercourse and refractory (Samsudewa and Capitan, 2013). This process is long, strenuous and very stressful to stags, hence it is very common for stags to lose body weight post rutting season (Savanth and Saseendran, 2012). This was most apparent in dominant and top ranking stags in their effort to establish and maintain the hierarchy. This study observed that T521 kept his harden antler the longest, portraying long exposure to stressful condition involved with establishment and maintenance of the antler.

There are various way to manage stress in deer herd during rutting season. In farm settings, it is possible to velvet the antler as recommended by Mourik (1985). However in establishment that does not practice velveting of antler, we recommend separating the stag once changed in behaviour such as distancing from the herd is observed. Rutting usually occur during the drier period in Malaysia (Zakaria et al., 2016). The fact that this coincide with dry season emphasize the need to increase feed supplementation to the stags due to poor pasture quality. While most of the maintenance feed is obtain from the pasture, it is ever more important to supplement higher quantity of concentrates to the animal during rutting season. Though stags general-

ly loose interest to feed during rutting season, observation should be made that all stags have access to supplemented feed during this poor quality pasture period. Observation should be conducted so that all animals has access to feed and not excessively bullied. Multiple feeding stations could help alleviate bullying and allow subordinate male to feed. This is more important in dominant stags since most of the energy was utilised for breeding and could lead to extreme stress condition rendering it to opportunistic immunosuppression related bacterial infection like *Mannheimia* and *Pasteurella*. At the same time, the stags could benefit to injectable prophylactic antibiotic, supplementation of vitamins and antioxidants dispensed in the water.

Hitherto, there is no available vaccine against manheimiosis and pasteurellosis specific for deer. And, there is no experimental and clinical evidence to suggest that available vaccines against these diseases in sheep and goats are useful in providing protection in deer. However, since the disease has already developed within this particular herd, usage of these vaccines may be useful. Vaccination protocols against manheimiosis and pasteurellosis in livestock is well documented, including vaccination using intranasal recombinant vaccine (Sabriet et al., 2013) or intranasal formalin-killed vaccine (Effendy et al., 1998). Vaccination should be done 2 weeks prior to expected stressful episodes. In deer management, perhaps, vaccination should be done with the consideration of the antler stages. Further research should be conducted to determine the prevalent serotype of *M. haemolytica* and *P. multocida* causing respiratory infection in deer. Subsequently, a more specific vaccine against these organisms should be produced.

Post-mortem examination of T521 which was diagnosed with manheimiosis revealed lesions resembling pneumonic manheimiosis in sheep and goats; cranioventral pulmonary hepatisation (Ekonget et al., 2015). However, unlike in most cases of manheimiosis in small ruminants, no fibrin formation and adhesion was noted in this particular deer. Possibility of species-specific differences reacting differently to *M. haemolytica* infection is an area yet to be discovered. Based on the post-mortem findings, it is believed that in this case, the particular deer was infected with *M. haemolytica*, leading to chronic pneumonic manheimiosis and abscessation. In other domestic ruminants, *M. haemolytica* leukotoxin are known to create pores in the pulmonary cell membrane, eventually leading to lysis (Clinkenbeard et al., 1989). At sub-lytic concentrations, the toxin activates neutrophils (Czuprynski et al., 1991), and induces severe inflammatory cytokine production (Yoo et al., 1995). On the other hand, its lipopolysaccharide is responsible for induction of IL-1beta and IL-8, leading to neutrophils influx and severe inflammation (Lafleur et al., 1998; Yoo, et al., 1995). Subsequently, these neutrophils will engulf and digest the agent. Degeneration and lysis of neutrophils

may occur, leading to release of the lysosomal enzyme into the lungs. The endotoxin of Gram-negative bacteria causing respiratory infection in ruminants are known to result in respiratory organ damage and increase pulmonary vascular permeability by damaging the endothelial cells (Annas et al., 2015). Collectively, these activities are thought to impair the lung immune mechanism (Highlander, 2001), and possibly allow entry of the agent into the circulation, leading to septicaemic mannaemia. In this case, complication by *Escherichia coli* infection occur via the aerogenous route, leading to concurrent acute colisepticaemia. However, further investigation on interactions between *M. haemolytica* with deer need to be established in order to fully understand the exact pathogenesis.

CONCLUSIONS

The present article reports a detailed report of manheimiosis in a deer with history of exposure to stress. Previous records revealed that the deer in the herd are also affected by other organism closely related to *M. haemolytica*, which is *P. multocida*. This respiratory diseases usually occur secondary to recent severe stress, thus, prevention steps revolves around reduction of stress by overall good animal husbandry practice. Special points to consider in deer management includes the complex interactions between multiples factors which could partially be elucidated by the patriarch-hierarchical relationship of the stags.

CONFLICT OF INTEREST

There is no conflict of interest.

AUTHOR'S CONTRIBUTION

Fitri, Wan-Nor and Annas Salleh wrote the manuscript with assistance and contributing ideas from Azrolharith Muhammad Rashid, Jesse Faez Firdaus Ariasamy, Zamri-Saad Mohd and Wahid Haron. Fitri, Wan-Nor and Azrolharith Muhammad Rashid were involved with ante mortem care, weight monitoring and provide veterinary treatment to the deer together with Wahid Haron. Annas Salleh performed post mortem examination and retrospective study. Zamri-Saad Mohd assisted in the histopathologic lesions examination.

REFERENCES

- wAnnas S Zamri-Saad M, Jesse FFA, Zunita Z (2014). New sites of localisation of *Pasteurella multocida* B: 2 in buffalo surviving experimental haemorrhagic septicaemia. BMC Vet. Res. 10 (1). 88. <https://doi.org/10.1186/1746-6148-10-88>
- Annas S, Zamri-Saad M, Jesse FFA, Zunita Z (2015). Comparative clinicopathological changes in buffalo and cattle following infection by *Pasteurella multocida*

- B: 2. Microbial pathogenesis. 88. 94-102. <https://doi.org/10.1016/j.micpath.2015.08.009>
- Audige L, Wilson PR, Morris RS (1998). BSC and its use for farmed deer. The Royal Society of New Zealand. 41. 545-553.
 - Bell TM (2008). Case I: Conference 19, Wednesday Slide Conference 2008-2009, 4 March 2009. The Armed Forces Institute of Pathology, Department of Veterinary Pathology.
 - Carrigan M, Dawkins H, Cockram F, Hansen A (1991). *Pasteurella multocida* septicaemia in fallow deer (*Dama dama*). Aust. Vet. J. 68: 201-203. <https://doi.org/10.1111/j.1751-0813.1991.tb03193.x>
 - Chung ELT, Abdullah FFJ, Abba Y, Tijjani A, Sadiq MA, Mohammed K (2015). Clinical management of pneumonic pasteurellosis in boer kids: A Case Report. Int. J. Livestock Res. 5(4). 94-99. <https://doi.org/10.5455/ijlr.20150417014307>
 - Clinkenbeard KD, Mosier DA, Confer AW (1989). Transmembraneporesizeandroleofcellswellingincytotoxicity caused by *Pasteurellahaemolytica* leukotoxin. Infect. Immun. 57. 420-425.
 - Czuprynski CJ, Noel EJ, Ortiz-Carranza O, Srikumaran S. (1991). Activation of bovine neutrophils by partially purified *Pasteurellahaemolytica* leukotoxin. Infect. Immun. 59. 3126-3133.
 - Donal OT (2015). Case II: Conference 3, Wednesday Slide Conference 2015-2016, 23 September 2015. Joint Pathology Center, Veterinary Pathology Services.
 - Dyer NW, Ward AC (1998). Pneumonic pasteurellosis associated with *Pasteurellahaemolytica* serotype A6 in American bison (Bison bison). J. Vet. Diagn. Invest. 10. 360-362. <https://doi.org/10.1177/104063879801000409>
 - Effendy AW, Zamri-Saad M, Puspa R, Rosiah S (1998). Efficacy of intranasal administration of formalin-killed *Pasteurellahaemolytica* A2 against intratracheal challenge in goats. Vet. Record. 142(16). 428-431. <https://doi.org/10.1136/vr.142.16.428>
 - Ekong PS, Akanbi BO, Odugbo MO (2015). A case report of respiratory mannheimiosis in sheep and goat complicated by Bordetellaparapertussis. Niger. Vet. J. 35(2).
 - Fatimah ZS, Fadila JF, Mat Kamaruzaman MA, Sang YW, Nursalleh KC, Fariza Y, Helmi MA (2016). Weather analysis from July until October 2015. In Technical Note No. 3/2016. Petaling Jaya: Jabatan Meteorologi Malaysia.
 - Foreyt WJ, Snipes KP, Kasten RW (1994). Fatal pneumonia following inoculation of healthy bighorn sheep with *Pasteurellahaemolytica* from healthy domestic sheep. J. Wildl. Dis. 30. 137-145. <https://doi.org/10.7589/0090-3558-30.2.137>
 - Grey CL, Thomson RG (1971). *Pasteurellahaemolytica* in the tracheal air of calves. Can. J. Comp. Med. 35. 121-128.
 - Hattel AL, Shaw DP, Love BC, Wagner DC, Drake TR, Brooks JW (2004). A retrospective study of mortality in Pennsylvania captive white-tailed deer (*Odocoileus virginianus*): 2000-2003. J. Vet. Diagn. Investig. 16(6). 515-521. <https://doi.org/10.1177/104063870401600605>
 - Highlander SK (2001). Molecular genetic analysis of virulence in *Mannheimia (Pasteurella) haemolytica*. Front. Biosci. 1. D1128-1150. <https://doi.org/10.2741/highland>
 - Lafleur RL, Abrahamsen MS, Maheswaran SK (1998). The biphasic mRNA expression pattern of bovine interleukin-8 in *Pasteurellahaemolytica* lipopolysaccharide-stimulated alveolar macrophages is primarily due to tumor necrosis factor alpha. Infect. Immun. 66. 4087-4092.
 - Misra HN, Pandurangarao CC, Khera SS (1970). An outbreak of highly fatal pneumonia in kids due to *Pasteurellahaemolytica*. Indian Vet. J. 47. 808-809.
 - Mohamed RA, Abdelsalam EB (2008). A review on pneumonic pasteurellosis (respiratory mannheimiosis) with emphasis on pathogenesis, virulence mechanisms and predisposing factors. Bulg. J. Vet. Med. 11(3): 139-160.
 - Mourik SV (1985). Expression and relevance of dominance in farmed rusa deer *Rusa timorensis*. Appl. Anim. Behav. Sci. 14: 275-287. [https://doi.org/10.1016/0168-1591\(85\)90008-5](https://doi.org/10.1016/0168-1591(85)90008-5)
 - Pavitt AT, Walling CA, Möstl E, Pemberton JM, Kruuk LEB. (2015). Cortisol but not testosterone is repeatable and varies with reproductive effort in wild red deer stags. Gen. Comp. Endocrinol. 222: 62-68. <https://doi.org/10.1016/j.yggen.2015.07.009>
 - Sabri MY, Shahrom-Salisi M, Emikpe BO (2013). Comparison prior and post vaccination of inactivated recombinant vaccine against Mannheimiosis in Boer Goats Farm in Sabah. J. Vacc. Vaccin.
 - Saif LJ (2010). Bovine respiratory coronavirus. Veterinary Clinics of North America: Food Anim. Pract. 26(2): 349-364. <https://doi.org/10.1016/j.cvfa.2010.04.005>
 - Samsudewa D, Capitan SS (2013). Reproductive behaviour of timor deer (*Rusa timorensis*). Wartazoa. 21(3). 108-113.
 - Savanth VV, Saseendran PC (2012). Impact of testosterone on the antler stages insambar deer (*Rusa unicolor*) Stags. 8(5): 279-285.
 - Straus DC, Purdy CW, Loan RW, Briggs RF, Frank GH (1998). In vivo production of neuraminidase by *Pasteurellahaemolytica* in market stressed cattle after natural infection. Curr. Microbiol. 37: 240-244. <https://doi.org/10.1007/s002849900372>
 - Yoo HS, Rajagopal S, Maheswaran SK, Ames TR (1995). Purified *Pasteurellahaemolytica* leukotoxin induces expression of inflammatory cytokines from bovine alveolar macrophages. Microb. Pathog. 18: 237-252. [https://doi.org/10.1016/S0882-4010\(05\)80001-4](https://doi.org/10.1016/S0882-4010(05)80001-4)
 - Zakaria MA, Zamri-Saad M, Hasliza AH, Wahid H (2016). Growth and reproductive performances of farmed timorensisdeer, cervustimorensis. Pertanika J. Trop. Agric. Sci. 39(1): 79-86.
 - Zamri-Saad M, Azri A, Nurida AB, Sheikh-Omar AR (1994). Experimental respiratory infection of goats with Mycoplasma arginini and *Pasteurellahaemolytica* A2. Pertanika J. Trop. Agric. Sci. 17(3): 239-242.