## **Review Article**



# Microalgae in Poultry Field: A Comprehensive Perspectives

### WAFAA A. ABD EL-GHANY\*

Poultry Diseases Department, Faculty of Veterinary Medicine, Cairo University, 12211 Giza, Egypt.

**Abstract** | Many years ago, antibiotics have been added to animals and poultry feed as growth promotor. Hazardous using of antibiotics on long run creates several health problems for animals as well as humans. Therefore, countries searched for natural alternatives that incorporate antimicrobials in livestock and poultry ration without any adverse effect on both productivity and health. One of these important and promising alternatives is microalgae. Microalgae are microscopic, uni- or multi-cellular and photosynthetic algae that grow in marine, fresh and brackish water. They considered as rich sources of proteins, essential fatty acids, carbohydrates, vitamins, minerals, pigments and antioxidants. Several types of microalgae have been used safely in human, animals and poultry. Different species of microalgae as *Spirulina, Chlorella* and others proved positive influences on poultry nutrition. Therefore, this review article aimed to show the effects of using different types of microalgae on the productive characteristics, immune response, microbial resistance and carcass trait of poultry.

Keywords | Chlorella, Immunity, Performance, Poultry, Spirulina

Received | May 07, 2020; Accepted | July 15, 2020; Published | July 25, 2020

\*Correspondence | Wafaa A. Abd El-Ghany, Poultry Diseases Department, Faculty of Veterinary Medicine, Cairo University, 12211 Giza, Egypt; Email: wafaa. ghany@yahoo.com

Citation | El-Ghany WAA (2020). Microalgae in poultry field: A comprehensive perspectives. Adv. Anim. Vet. Sci. 8(9): 888-897. DOI | http://dx.doi.org/10.17582/journal.aavs/2020/8.9.888.897

ISSN (Online) | 2307-8316; ISSN (Print) | 2309-3331

Copyright © 2020 El-Ghany. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## **INTRODUCTION**

Doultry industry is considered as one of the most important source of income all over the world. Due to continuous and extensive increase in human population, the need for safe and protein rich sources of poultry or livestock becomes very necessary. Antibiotics in feed resulted in development of drug-resistant bacteria (Sorum and Sound, 2001), antibiotic residues in products (Burgat, 1999) and decrease the numbers of intestinal microflora (Andremont, 2000). Hazardous using of antimicrobials in poultry field threatens this industry as they affect both birds and humans health. The European Union (European Commission, 2001) banned using of antibiotics growth promoters in poultry nutrition. So, many biologicals alternative strategies have been investigated to incorporate antimicrobials natural compounds without any adverse effect on both productivity and health.

Microalgae are microscopic, uni or multi-cellular and photosynthetic type of algae that present in both salty and fresh aquarium. Microalgae is regarded as a perfect

source of valuable and naturally efficient nutrients, it extensively gained particular interest, attention and application among multiple producers worldwide. They have been tested in experimental animals hundreds of years as immunomodulatory, anti-inflammatory, antioxidant, antimicrobial and antiviral and proved their efficacy (Uyisenga et al., 2010; Abdel-Daim et al., 2013; Shokri et al., 2014; Ashgan et al., 2015). Microalgae may also considered as a promising fundamental constituents of animals and poultry ration. Blue-green microalgae; Arthrospira (Spirulina species) as Spirulina platensis (S. platensis) and S. maxima are the most efficient and widely distributed edible feed additive microalgae in humans, animals and poultry (Yoshida and Hoshi, 1980; Vonshak, 2002; Meineri et al., 2009; Kanagaraju and Omprakash, 2016). It has been found that Spirulina contains 50% -70% proteins (Soni et al., 2017) as well as most essential amino acids (Anusuya et al., 1981), so it could be used as alternative to protein sources in poultry ration (Spolaore et al., 2006; Austic et al., 2013). Besides, this type of microalgae showed high contents of fatty acids, carbohydrates, vitamin B complex (thiamine, riboflavin, pyridoxine and vitamin

## <u>OPEN OACCESS</u>

B12), vitamin C, vitamin A, vitamin E, minerals (calcium, iron, zinc, manganese, magnesium, phosphorus, copper, chromium, sodium and potassium), pigments (carotenoids, phycocyanin and xantophylls) and antioxidants (Ross and Dominy, 1990; Khan et al., 2005; Cheong et al., 2010; Holman and Malau-Aduli, 2012; Beheshtipour et al., 2013; Jafari et al., 2014; AbouGabal et al., 2015; Soni et al., 2017; Hynstova et al., 2018).

*Chlorella* species as *Chlorella vulgaris* is another type of natural green microalga that showed growing importance owing to beneficial nutritional and functional characteristics (Grau and Klein, 1957; Pulz and Gross, 2004; Sugiharto and Lauridsen, 2016). *Chlorella* is rich in proteins (48%), omega 3 and 9 fatty acids and phosphorus (Tokusoglu et al., 2003; Yaakob et al., 2014; EL-Mohsnawy et al., 2020). *Chlorella* has been sold as healthy food, cosmetics, antioxidants and as animal feed (Becker, 2007; Lee et al., 2010). In addition, other types of microalgae were defined and used for nutrition of broilers and layers without adverse effect on production or health.

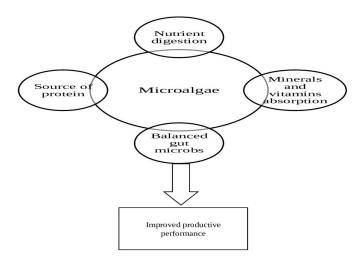
Therefore, this review article was planned to deeply investigate the effects of using different types of microalgae on the productive characteristics, immune response, microbial resistance and carcass trait of poultry.

#### INFLUENCE OF USING MICROALGAE IN POULTRY FIELD PRODUCTIVE CHARACTERISTICS

The mode of action of microalgae for improving the performance parameters is illustrated in Figure 1. Dietary incorporation of S. platensis for broilers were associated with improvement in live body weight, body weight gain and the feed conversion rate of broilers (Kharde et al., 2012; Shanmugapriya and Babu, 2014; Shanmugapriya et al., 2015; Park et al., 2018). The early study of Saxena et al. (1983) demonstrated that Spirulina at levels of 111 g/kg and 166 g/kg diet of White Leghorn chicks could replace groundnut cake with improving the weight gain of the birds. Similarly, Venkataraman et al. (1994) detected that addition of Spirulina (140 and 170 g/kg diet), without additional vitamins/minerals, could substitute groundnut cake and fishmeal and had no adverse effects on chicken's performance. Addition of Spirulina at a level of 12% to broiler ration could replace the source of protein in it as well as enhance the growing and feed conversion rate (Ross and Dominy, 1990). Spirulina showed an improvement of nutrient digestion, minerals absorption and protection from diarrhea (Gružauskas et al., 2004). According to Nikodémusz et al. (2010), feeding of pheasants on ration containing 0.3 g Spirulina/ kg diet induced superior productive performance when compared with non-fed control birds. The same results were obtained by Mariey et al. (2014) who found that supplementation of broilers with very low level of Spirulina (0.02 or 0.03%) improved the

### Advances in Animal and Veterinary Sciences

performance. Kaoud (2015) recorded similar improvement in performance parameters induced by dietary inoculation of S. platensis when compared with prebiotic treated controls. It was demonstrated that incorporation of Spirulina algae up to 16% in broiler starter ration induced no negative effects on the chick's performance (Evans et al., 2015). The authors determined that Spirulina contains high levels of both energy and proteins which representing 90% and 76%, consequently. In Japanese quails, addition of Spirulina powder in diet at level of 1% or in drinking water at 0.25% had beneficial significant effects on body weight, weight gain, conversion rate and fertility when compared with control groups (Abouelezz, 2017). Fathi et al. (2018) demonstrated that broilers diet supplemented by 0.7 and 0.9 g/kg of feed S. platensis could improve the growth performance parameters, in addition, S. platensis supplementing level (0.7g/kg) was economically recommended. Moreover, Abd El-Hady and El-Ghalid (2018) recorded that the final body weight, weekly body weight gain and feed conversion rate of broilers fed on 6% S. platensis were significantly higher than birds fed on 3% of this algae at 42 days of age. Treatments of ration with 0.25, 0.5, 0.75, or 1.0% Spirulina enhanced broilers body weight gain, feed conversion, and/or European production efficiency index during 8 to 21, 22 to 35, and overall 1 to 35 days of age (Park et al., 2018).



**Figure 1:** The mode of action microalgae to improve the production performance.

Regarding the effect of *Chlorella* on bird's performance, a very early experiment conducted in chickens using 10% dried *Chlorella* into feed revealed increased performance in birds (Combs, 1952). The authors referred this result to the presence of pigments (carotene) and vitamins (B2 and B12) in *Chlorella* if the diet of birds didn't contain these components. As well, *Chlorella* at dietary level of 5 or 10% provoked no adverse effects on broilers growth performance as algae considered as an excellent protein source (Lipstein and Hurwitz, 1983). In a subsequent study, Kang et al. (2013) concluded that addition of fresh

### Advances in Animal and Veterinary Sciences

liquid form of *Chlorella* at 1% dietary level beneficially affected body weight gain. The positive effect of feed supplementation of ducks with 0.1 or 0.2% *Chlorella* on body weight gain and feed intake was confirmed by Oh et al. (2015). Broiler chicken watered with culture of *Chlorella vulgaris* showed less consumption of food and the highest percentage of performance index and relative economic efficiency (Niamat and Ragaa, 2017).

Considering other microalgae, it was observed that addition of 6% of *Micractinium* spp. to broilers diet also induced positive effect on performance parameters (Lipstein and Hurwitz, 1981).

The improvement in performance characters may be related to the role of *Spirulina* in inducing balanced gut microbial population that enhances absorption of dietary vitamins and minerals and consequently increases the feed utilization efficiency (Tsuchihashi et al., 1987; Mariey et al., 2012).

The effect of using microalgae to improve the productive performance was contradictive, certain study showed that there was no improvement in the body production of chickens after feeding on ration containing 4 or 8% of *Spirulina* (Toyomizu et al., 2001). Zahroojian et al. (2013) also found that there was no difference in feed intake of laying hens fed on 1.5, 2 and 2.5% *Spirulina* and non-supplemented control group. Likewise, the cumulative feed intake in the 1% *Spirulina* treated birds was lesser than that of the control group (Kanagaraju and Omprakash, 2016). Water treatment of laying hens with *Chlorella* for 9 weeks induced no significant positive effect on feed intake or conversion ratio (Moradikor and Mohamadi, 2015).

Considering egg production performance parameters, it was noticed that supplementation of laying Japanese quails diet with Spirulina (1.5, 3.0, 6.0, or 12.0%) produced an increase in egg production with good egg quality (Ross and Dominy, 1985, 1990). Spirulina is considered as a very good source of pigments like carotenoids and xanthophylls which are very important of the intensity of egg yolk colour, as feeding of quails on dietary 1% Spirulina induced optimal colour of the egg yolk (Anderson et al., 1991). It was demonstrated that addition of 2.0-2.5% Spirulina to the diet of layers improved the absorption and accumulation of carotenoids and consequently increase the yolk colour intensity (Zahroojian et al., 2011, 2013). Similar experiment was carried out by Sujatha and Narahari (2011) who demonstrated that boiled eggs from Spirulina treated layer chickens exhibited sensorily more acceptable yolk colour than control birds. Increasing in egg performance, egg yolk colour and hatchability have been observed in hens fed on ration with of 0.10.2% Spirulina (Mariey et al., 2012). The effect of microalgae on fatty acids contents

2020 | Volume 8 | Issue 9 | Page 890

of eggs was also studies by Ginzberg et al. (2000) who found that addition of *Spirulina* valorized egg products by decreasing their contents of cholesterol and saturated fatty acid contents while increasing levels of omega-3 poly unsaturated fatty acids (PUFA). The same authors indicated significant increase in fertility and hatchability percentage of the produced eggs. Very recently, 29 to 40 weeks old layer chicken hens received 3 g *S. platensis* /kg produced significant higher egg number, egg weight and egg mass than control (Mobarez et al., 2018). Aljumaily and Taha (2019) reported that inoculation of quail's eggs with *Spirulina* liquid extracts in the late stages of incubation could enhance the percentages of eggs hatchability, increase the chances of chick's survival as well as strengthen their immunity and antioxidants status.

Feeding of laying hens with Chlorella has a beneficial influence egg quantity and quality as well as intestinal contents of lactic acid producing bacterial populations (Zheng et al., 2012). It was detected that Chlorella enhanced the production of lutein enriched eggs that inhibit macular destruction in elder human. In this context, supplementation of hens with conservative or Chlorella rich in lutein could effectively increase the egg quantity, lutein content and the yolk colour of eggs (An et al., 2014). Moreover, Chlorella increased lutein and zeaxanthin concentrations as well as improved the external egg quality and the oxidative constancy of fats in yolk of eggs (Englmaierova et al., 2013). Carotenes and xantophylls from the microalgae affect the egg yolk colour (Arakawa et al., 1960). Layer chickens taken ration containing Chlorella vulgaris produced more intensive yellow colour of the egg yolk as it is a rich source of carotenoid content (Lipstein et al., 1980; Batista et al., 2013). Spray dried or bullet milled and spray dried Chlorella vulgaris increased the number of the produced eggs, yolk colour and shell weight as well as hatchability performance (Halle et al., 2009). Janczyk et al. (2009) suggested that Chlorella encouraged increasing the verities of microflora in the intestinal tract which is important for the quality of eggs.

Marine microalgae (*Schizochytrium limacinum*) powder also exhibited positive effect on laying performance and egg quality as it enhanced docosahexaenoic acid (DHA) yolk concentration without alteration of sensory characteristics of eggs (Parpinello et al., 2006; Rizzi et al., 2009; Park et al., 2015). Addition of red seaweeds *C. crispus* and *Sarcodiotheca gaudichaudii* was found to be effectively act like prebiotics to improve chicken gut health, productivity, and egg quality (Kulshreshtha et al., 2014). In addition, Japanese quail layers fed on diet containing 0.5% *Schizochytrium* spp. microalgae produced eggs with high DHA concentration, low n-6/n-3 PUFA proporation and cholesterol level in the yolk fats (Gladkowski et al., 2014; Trziszka et al., 2014). As a source of n-3 PUFA, a dietary level 2.4% of *Isochrysis* 

galbana microalgae was considered as a rich source in egg yolk (Lemahieu et al., 2013, 2014). In the same context, diet containing Nannochloropsis gaditana might be used as an alternative source of omega-3 PUFA to produce DHA enriched eggs (Bruneel et al., 2013). The effect of feeding of layers on different concentrations of defatted Staurosira microalgae (soybean replacer) was tested (Leng et al., 2014) and the authors concluded that a level 7.5% of algae had no adverse effects, however, levels of 15% had bad effects on egg performance, feed intake and feed conversion rate. Two species of microalgal biomass (Desmodesmus and Staurosira) when used in laying hen ration at level up to 25% were considered as rich sources of protein with without bad effects on laying (Ekmay et al., 2015).

However, the effect of micro algae on egg performance parameters were also conflicting. The study of Moradikor and Mohamadi (2015) indicated that inoculation of different levels of *Chlorella* (0, 100, 200, 300 and 400 ppm) to the drinking water of laying hens for 9 weeks experimental period had no significant impact on egg production and egg mass. Moreover, *Spirulina* powder at level 1% in the feed and at 0.25% in the drinking water of 14-days-old Japanese quails did not show any significant effect on egg laying rate, egg weight and daily egg mass (Abouelezz, 2017).

The decrease in mortality rate of birds fed on microalgae reflects the improvement of the general health condition of birds. This speculation is supported by Cheong (2014) who noticed that supplementation of quails diet with 2% *Spirulina* resulting in significant reduction in the mortality rate.

#### **I**MMUNE RESPONSE

Enhancement of the immune response by microalgae is showed in Figure 2. The effect of supplementation with microalgae in the diet of chickens on the immune response was tested (Qureshi et al., 1994, 1996; Raju et al., 2004). The authors noticed an enhancement of phyto-haemagglutininintermediated the propagation of lymphocytes and the phagocytosis of macrophages as well as development of lymphoid organs. Birds received dietary Spirulina showed good health conditions indicating enhancement of immune response against diseases (Baojiang, 1994; Holman and Malau-Aduli, 2012). Furthermore, addition of *Spirulina* at level of (0.05%) to broiler ration could partially reduce the adverse properties of mycotoxins on the weights of bursa, thymus and spleen (Raju et al., 2005). This enhancement was expressed by increasing the ability to kill microbes, synthetization of antigens and increasing of T cell activity. Broilers treated with S. platensis algae appeared to have an improved immune system as demonstrated by a significant increase in white blood cell count and enhanced macrophage phagocytic activity (Al-Batshan et al., 2001; Mariey et al.,

#### Advances in Animal and Veterinary Sciences

2014). The health beneficial effect of S. platensis in poultry is related to the antimicrobial, immune-modulatory, antiinflammatory and antioxidant capacity potentials of algae (Farag et al., 2016). Kaoud (2015) indicated an increase in the relative and absolute thymus and bursa weights of chicken groups fed diet containing Spirulina compared to the control group. In this respect, addition of S. platensis at levels of 0.7 and 0.9 g/kg broiler ration displayed significant increase in bursa, thymus and spleen weights as well as the increasing the level of serum globulin when compared with un-treated control (Fathi et al., 2018). Lokapirnasari et al. (2016) and Widya et al. (2016) revealed that treatment with S. platensis increased the number of leukocytes and it could be recommended as feed additive to increase the immunity of infected chickens against avian influenza (AI) H5N1 virus. Zeweil et al. (2016) detected that using of S. platensis at levels (0.5 and 1 g/kg diet) and vitamin E (75 mg/kg diet) enhanced the total antibody production specific for Newcastle disease virus (NDV) vaccine in heat stressed broiler chicks. Furthermore, Mobarez et al. (2018) indicated that dietary supplementation of layer diet with 3 g S. platensis /kg provoked antibody titers against NDV, AI, sheep red blood cells (SRBCs) and increased interferon proteins concentration compared to the control group. The enhancement of cellular immunity may be correlated to an increase of Zn concentration in Spirulina (Mohamed, 1998; Abdel-Daim et al., 2013; Abou-Gabal et al., 2015).

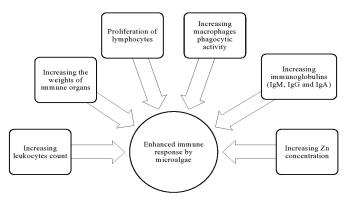


Figure 2: Enhancement of the immune response by microalgae.

The effect of different forms of *Chlorella* algae in improving of immune indices was also showed by Kang et al. (2013). The results revealed that 1% dietary level of fresh liquid form of *Chlorella* had a positive effect on the immune response of the birds and it was expressed by increasing the count of white cells and lymphocytes as well as the level of immunoglobulins (IgG, IgM and IgA) in the blood. Stimulation of immune response by dietary *Chlorella* may be due to several mode of actions, the content of fibers and presence of polysaccharide (immurella), polypeptides as well as glycoprotein. In agreement, it has been demonstrated that chickens taken ration containing 0.5% *Chlorella* showed increasing in leucocytes phagocytic activity and

lymphatic tissue development (Kotrbacek et al., 1994). Numerical rise in the reaction to phyto-hemagglutinin had been detected in broilers fed on diet with *Chlorella* (Rezvani et al., 2012). Moradikor and Mohamadi (2015) demonstrated that addition of 400 ppm of *Chlorella* to the drinking water of laying hens increase the antibody titers against SRBCs as expressed by increasing the levels of IgM and IgG.

### MICROBIAL RESISTANCE

Early findings of Tsuchihashi et al. (1987) suggested that feeding on ration containing Spirulina may boost population of Lactobacilli and increase vitamins absorbability. Concomitantly, Kulshreshtha et al. (2008) concluded that Spirulina is useful for the beneficial intestinal flora. It was demonstrated that S. platensis shared probiotics characters being possess a stimulating effect on the growth of Lactobacilli and Streptococcus thermophilus (Bhowmik et al., 2009). Dietary supplementation of broilers with Spirulina induced hypo-cholesterolmic properties due to reducing the synthesis and/or absorption of cholesterol in the intestinal tract as well as increasing the Lactobacillus population (Mariey et al., 2012). Chickens diets containing S. platensis demonstrated an increase in the number of intestinal lactic acid bacteria and a decrease in Escherichia coli (E. coli) (Shanmugapriya et al., 2015). As well, an increase in Lactobacilli count in the gut of Japanese quails after feeding with S. platensis was recorded (Yusuf et al., 2016). Recently, the results of Fathi et al. (2018) indicated significant decrease in the total intestinal bacterial and E. coli counts but significant increase in Lactobacillus count in broilers fed 0.9 and 0.7g S. platensis/kg ration. Kang et al. (2013) concluded that dietary supplementation of 1 % fresh liquid Chlorella improved body weight gain and increased the production of Lactobacillus bacteria in the intestinal microflora of broiler chickens. Inoculation of broiler diet with 1 % Spirulina powder increased the caecal Lactobacillus count (Park et al., 2018).

Feeding *Chlorella* microalgae resulted in increased *Lactobacillus* diversity in the crop or cecum or both of laying hens (Janczyk et al., 2009). Correspondingly, ducks fed on diet containing 0.1 or 0.2% *Chlorella* showed positive influence on the caecal microflora (Oh et al., 2015).

As regards the *in-vitro* antimicrobial effect of micro-algae, it was documented that *S. platensis* crude extract inhibited the growth of certain organisms including; *Klebsiella pneumoniae*, *Shigella shigae*, *Pseudomonas aeruginosa*, *E. coli*, *Proteus vulgaris*, *Salmonella typhi* (*S. typhi*) and *Staphylococcus aureus* (*S. aureus*) (Mala et al., 2009). Similar results were obtained by Kaushik and Chauhan (2008) and El-Baz et al. (2013) as they recorded antibacterial activities of *S. platensis* against *S. aureus*, *E. coli*, *S. typhi* and *Enterococcus faecalis*. *In vivo* study of Nuhu (2013) revealed *S. platensis* 

2020 | Volume 8 | Issue 9 | Page 892

### Advances in Animal and Veterinary Sciences

bacterial clearance capacity after experimental infection of chicks with E. coli and S. aureus. In Campylobacter jejuni experimentally infected broiler chickens, the efficiency of elevating the ration inoculation dose of Haematococcus pluvalis (astaxanthin source) was evaluated. The results revealed that the tissue concentrations of astaxanthin were increased by increasing the levels of dietary algae, no effect of algal meal on performance and the caecal population of Campylobacter jejuni was also not affected, but a level of dietary 0.18% of algae reduced the count of caecal Clostridium perfringens (Waldenstedt et al., 2003). Lorenz and Cysewski (2000) concluded that incorporation of poultry feed with microalgae Haematococcus spp. as a source of protein could improve their health and productivity. The antimicrobial effect of micro-algae may be related to presence of some acids like  $\gamma$  linolenic acid, active fatty acid, lauric and palmitoleic acid (El-Sheekh et al., 2014) or due to existence of alkaloids and lipopolysaccharides (Rania and Hala, 2008).

Nevertheless, results recorded in the study of Sugiharto et al. (2018) detected that treatment of broilers with *S. platensis* did not affect the ileal and caecal populations of *Lactobacilli*.

#### **CARCASS TRAIT**

It was shown that replacement of groundnut protein or fish meal by up to 170 or 140 g/kg of broiler ration Spirulina induced deeper color pigmentation of skin, breast, and thigh muscles (Venkataraman et al., 1994). Addition of Spirulina in broiler ration influenced the yellow and red colour of meat (Toyomizu et al., 2001). Accumulation of yellow pigmentation in flesh is related to presence of zeaxanthin pigments of Spirulina. Mariey et al. (2014) found an increase in dressing percentage, meat colour score and decreased relative abdominal fat weight after raising of birds on dietary 0.02 or 0.03% Spirulina. Similarly, feeding of ducks on dietary 0.1 or 0.2% Chlorella was able to improve the meat quality (Oh et al., 2015). The improvement of meat quality may be related to antioxidants activity of Spirulina which enhances integrity of muscle fibers and consequently capability of muscles to retain water (Dal Bosco et al., 2014). Bonos et al. (2016) documented that dietary Spirulina (5 g/kg of ration) influenced improving in the meat quality of broiler chicks through increasing the contents of eicosapentaenoic, docosapentaenoic and docosahexaenoic acids in the thigh muscles of birds. Lately, Altmann et al. (2018) concluded that soybean meal in broiler ration could be replaced safely by Spirulina without deteriorating meat quality.

To study the effect of *Crypthecodinium cohnii* microalgae on the carcass traits of Muscovy ducks, Schiavone et al. (2007) found that addition of 0.5% of this algae induced positive effect on the contents of fats in the breast muscles

#### Advances in Animal and Veterinary Sciences

## OPEN OACCESS

with no bad effects on production, carcass quality, and biochemical characteristics like pH, colour, oxidative constancy as well as sensory features of the breast muscles. Another study on *Schizochytrium* microalgae demonstrated that incorporation of this algae in broiler diet at levels 0.1 or 0.2% enriched the contents of fatty acids in the lipid of breast muscles with no adverse effect on the body weight gain (Yan and Kim, 2013). Supplementation of 3% green seaweed *Ulva lactucain* to broiler chickens increased the yield of breast muscle in comparison with birds fed corn diet only (Abudabos et al., 2013).

Some studies should that addition of microalgae has no influence on the carcass trait. Cheong et al. (2016) found that 15-35 days old Japanese quails supplemented with up to 8% of *Spirulina* in feed exhibited no significant effects on carcass composition or meat quality by decreasing the drip loss. In the same context, *Spirulina* in diet or in the drinking water of Japanese quails did not display significant differences considering the dressing percentage, the relative weights of heart, gizzard, liver, abdominal fat, testes, spleen and ovary as well as the length of caeca, small intestine and oviduct (Abouelezz, 2017). There was no significant effect of 1% *Spirulina* on relative organ weight and breast meat quality of broilers fed with diets (Park et al., 2018).

### CONCLUSION

From above mentioned, it could be concluded that different types microalgae could be used as useful and safe alternatives to many constituents of poultry ration like proteins, fatty acids and other essential elements. Microalgae proved great efficacy in improving the productive performance of broilers and layers, enhancement of cellular and humoral immunity, maintaining the beneficial intestinal bacteria and destroy the pathogenic ones as well as improving the carcass traits. So, in the near future, there is a great demand to use microalgae in a large scale of poultry industry.

### **AUTHOR CONTRIBUTION**

Wafaa A. Abd El-Ghany collected the data, wrote and prepared the manuscript.

### **CONFLICT OF INTEREST**

The author have declared no conflict of interest.

### **REFERENCES**

- Abdel-Daim MM, Abuzead SM, Halawa SM (2013). Protective role of *Spirulina platensis* against acute deltamethrininduced toxicity in rats. PLoS One. 8: e72991. https://doi. org/10.1371/journal.pone.0072991
- •Abd El-Hady AM, El-Ghalid OAH (2018). Spirulina platensis

Algae (SPA): A novel poultry feed additive. Effect of SPA supplementation in broiler chicken diets on productive performance, lipid profile and calcium-phosphorus metabolism. World's Poult. Sci. J. 7498: 1-7.

- Abouelezz FMK (2017). Evaluation of *Spirulina* algae (*Spirulina platensis*) as a feed supplement for Japanese quail: Nutritional effects on growth performance, egg production, egg quality, blood metabolites, sperm-egg penetration and fertility. (2017). Egypt. Poult. Sci. J. 37: 707-719. https://doi.org/10.21608/epsj.2017.5390
- AbouGabal A, Aboul-Ela HM, Ali E, Ahemd E, Shalaby OK (2015). Hepatoprotective, DNA damage prevention and antioxidant potential of *Spirulina platensis* on CCl4-induced hepatotoxicity in mice. Am. J. Biomed. Res. 3: 29-34. http:// pubs.sciepub.com/ajbr/3/2/3
- Abudabos AM, Aly BO, Riyadh SA, Emad MS, Kalid AA, Ahmed AA (2013). Nutritional value of green seaweed (*Ulva Lactuca*) for broiler chickens. Ital. J. Anim. Sci. 12: 177-181. https://doi.org/10.4081/ijas.2013.e28
- Al-Batshan HA, Al-Mufarrej SI, Homaidan AA, Qureshi MA (2001). Enhancement of chicken macrophage phagocytic function and nitrite production by dietary *Spirulina platensis*. Immunopharmacol. Immunotoxicol. 23: 281-289. https:// doi.org/10.1081/IPH-100103866
- Aljumaily TKH, Taha AT (2019). Effects of Spirulina platensis algae extract early feeding on Japanese quail embryos. Adv. Anim. Vet. Sci. 7: 30-37. https://doi.org/10.17582/journal. aavs/2019/7.1.30.37
- Altmann BA, Neumann C, Velten S, Liebert F, Mörlein D (2018). Meat quality derived from high inclusion of a microalga or insect meal as an alternative protein source in poultry diets: A Pilot Study Foods. 7: 34. https://doi.org/10.3390/ foods7030034
- An BK, Jeon JY, Kang CW, Kim JM, Hwang JK (2014). The tissue distribution of lutein in laying hens fed lutein fortified *Chlorella* and production of chicken eggs enriched with lutein. Korean J. Food Sci. Anim. Res. 34: 172-177. https:// doi.org/10.5851/kosfa.2014.34.2.172
- Anderson DW, Tang CS, Ross E (1991). The xanthophylls of Spirulina and their effect on egg yolk pigmentation. Poult. Sci. 70: 115-119. https://doi.org/10.3382/ps.0700115
- Andremont A (2000). Consequences of antibiotic therapy to the intestinal ecosystem. Ann. Fr. Anesth. Reanim. 19: 395-402. https://doi.org/10.1016/S0750-7658(00)90209-0
- Anusuya DM, Subbulakshimi G, Madhavi Devi K, Venkataram LV (1981). Studies on the proteins of masscultivated. Bluegreen alga (*Spirulina platensis*). J. Agric. Food Chem. 29: 522-525. https://doi.org/10.1021/jf00105a022
- Arakawa S, Tsurumi N, Murakami K, Muto S, Hoshino J, Yagi T (1960). Experimental breeding of white leghorn with the *Chlorella*-added combined feed. Jpn. J. Exp. Med. 30: 185-192.
- Ashgan AA, Haiam MA, Eman MA, Khaled AEM, Ola KS (2015). Hepato-protective, DNA damage prevention and antioxidant potential of *Spirulina platensis* on CCl4induced hepatotoxicity in mice. Am. J. Biomed. Res. 2: 29-34.
- Austic RE, Mustafa A, Jung B, Gatrell S, Lei XG (2013). Potential and limitation of a new defatted diatom microalgal biomass in replacing soybean meal and corn in diets for broiler chickens. J. Agri. Food Chem. 61: 7341-7348. https://doi.org/10.1021/jf401957z
- •Baojiang G (1994). Study on effect and mechanism of polysaccharides of *Spirulina* on body immune function



improvement. Second Asia-Pac. Conf. Algal Biotechnol. nu

- Singapore.
  Batista AP, Gouveia L, Bandarra NM (2013). Franco JM, Raymundo A. Comparison of microalgal biomass profiles as novel functional ingredient for food products. Algal Res. 2: 164-173. https://doi.org/10.1016/j.algal.2013.01.004
- •Becker EW (2007). Micro-algae as a source of protein. Biotechnol. Adv. 25: 207-210. https://doi.org/10.1016/j. biotechadv.2006.11.002
- Beheshtipour H, Mortazavian AM, Mohammadi R, Sohrabvandi S, Khosravi-Darani K (2013). Supplementation of *Spirulina platensis* and *Chlorella vulgaris* algae into probiotic fermented milks. Compr. Rev. Food Sci. Food Saf. 12: 144-154. https:// doi.org/10.1111/1541-4337.12004
- Bhowmik D, Dubey J, Mehra S (2009). Probiotic efficiency of *Spirulina platensis*-stimulating growth of lactic acid bacteria. American-Eurasian J. Agric. Environ. Sci. 6: 546-549. http://www.idosi.org/aejaes/jaes6(5)/9.pdf
- Bonos E, Kasapidou E, Kargopoulos A, Karampampas A, Christaki E, Florou-Paneri P, Nikolakakis I (2016). *Spirulina* as a functional ingredient in broiler chicken diets. S. Afr. J. Anim. Sci. 46: 94-102. https://doi.org/10.4314/ sajas.v46i1.12
- Bruneel C, Lemahieu C, Fraeye I, Ryckebosch E, Muylaert K, Buyse J, Foubert I (2013). Impact of microalgal feed supplementation on omega-3 fatty acid enrichment of hen eggs. J. Funct. Foods. 5: 897-904. https://doi.org/10.1016/j. jff.2013.01.039
- Burgat V (1999). Residues of drugs of veterinary use in food. La Revue du Praticien, 41: 985-990.
- •Cheong DSW (2014). Effect of supplementing microalgae diet on growth performance and carcass characteristic of Japanese quail. M. Sc. thesis submitted to the University Putra, Malaysia.
- Cheong DSW, Kasim A, Sazili AQ, Omar H, Teoh JY (2016). Effect of supplementing *Spirulina* on live performance, carcass composition and meat quality of Japanese quail. Walailak J. Sci. Technol. 13: 77–84.
- Cheong SH, Kim MY, Sok DE, Wang SYH, Kim JH, Kim HR, Lee JH, Kim YB, Kim MR (2010). *Spirulina* prevents atherosclerosis by reducing hypercholesterolemia in rabbits fed a high-cholesterol diet. J. Nutr. Sci. Vitaminol. 56: 34– 40. https://doi.org/10.3177/jnsv.56.34
- Combs GF (1952). Algae (*Chlorella*) as a source of nutrients for the chick. Science. 116: 453-454. https://doi.org/10.1126/ science.116.3017.453
- Dal Bosco A, Gerencsér Z, Szendro Z, Mugnai C, Cullere M, Kovács M, Ruggeri S, Matti SO, Castellini C, Dalle Zotte A (2014). Effect of dietary supplementation of *Spirulina (Arthrospira platensis)* and Thyme (*Thymus vulgaris*) on rabbit meat appearance, oxidative stability and fatty acid profile during retail display. Meat Sci. 96: 114-119. https:// doi.org/10.1016/j.meatsci.2013.06.021
- Ekmay RD, Chou K, Magnuon A, Lei XG (2015). Continual feeding of two types of microalgal biomass affected protein digestion and metabolism in laying hens. J. Anim. Sci. 93: 287-297. https://doi.org/10.2527/jas.2014-7506
- El-Baz FK, El-Senousy WM, El-Sayed AB, Kamel MM (2013). *In vitro* antiviral and antimicrobial activities of *Spirulina platensis* extract. J. Appl. Pharm. Sci. 3: 52-56.
- EL-Mohsnawy E, El-Sheekh MM, Mabrouk MEM, Zohir WF (2020). Enhancing accumulation of omega 3 and 9 fatty acids in *Chlorella vulgaris* under mixotrophic

### Advances in Animal and Veterinary Sciences

nutrition. J. Anim. Plant Sci. 30: 485-492. https://doi. org/10.36899/JAPS.2020.2.0047

- El-Sheekh MM, Daboor SM, Swelim MA, Mohamed S (2014). Production and characterization of antimicrobial active substance from *Spirulina platensis*. Iran. J. Microbiol. 6: 112-119. https://www.ncbi.nlm.nih.gov/pubmed/25705362
- Englmaierova M, Skrivan M, Bubancova I (2013). A comparison of lutein, spray-dried *Chlorella*, and synthetic carotenoids effects on yolk colour, oxidative stability, and reproductive performance of laying hens. Czech J. Anim. Sci. 58: 412-419. http://agriculturejournals.cz/web/cjas.htm https://doi. org/10.17221/6941-CJAS
- European commission (2001). 2<sup>nd</sup> opinion on anti-microbial resistance [cited 2009 Feb 11]. Available from: http://ec.europa.eu/food/fs/sc/ssc/ out203\_en.pdf.
- Evans AM, Smith DL, Moritz JS (2015). Effects of algae incorporation into broiler starter diet formulations on nutrient digestibility and 3 to 21 d bird performance. J. Appl. Poult. Res. 24: 206-214. https://doi.org/10.3382/japr/ pfv027
- Farag MR, Alagawany M, Abd El-Hack ME, Dhama K (2016). Nutritional and healthical aspects of *Spirulina (Arthrospira)* for poultry, animals and human. Inter. J. Pharmacol. 12: 36-51. https://doi.org/10.3923/ijp.2016.36.51
- Fathi MA, Namra MMM, Ragab MS, Aly MMM (2018). Effect of dietary supplementation of algae meal (*Spirulina platensis*) as growth promoter on performance of broiler chickens. Egypt. Poult. Sci. J. 38: 375-389.
- Ginzberg A, Cohen M, Sod-Moriah UA, Shany S, Rosenshtrauch A, Arad SM (2000). Chickens fed with biomass of the red microalga *Porphyridium* sp. have reduced blood cholesterol level and modified fatty acid composition in egg yolk. J. Appl. Phycol. 12: 325-330. https://doi. org/10.1023/A:1008102622276
- Gladkowski W, Kiełbowicz G, Chojnacka A, Bobak Ł, Spychaj R, Dobrzanski Z, Trziszka T, Wawrzenczyk C (2014). The effect of feed supplementation with dietary sources of n-3 polyunsaturated fatty acids, flaxseed and algae *Schizochytrium* sp., on their incorporation in to lipid fractions of Japanese quail eggs. Int. J. Food Sci. Technol. 49: 1876-1885. https:// doi.org/10.1111/ijfs.12497
- Grau CR, Klein NW (1957). Sewage-grown algae as feed stuff for chicks. Poult. Sci. 36: 1046-1051. https://doi. org/10.3382/ps.0361046
- Gružauskas R, Lekavičius R, Racevičiūtė-Stupelienė A, Šašytė V, Tėvelis V, Švirmickas GJ (2004). Viščiukų broilerių virškinimo procesų optimizavimas simbiotiniais preparatais. Vet. Irzoote. 28: 51-56.
- Halle I, Janczyk P, Freyer G, Souffrant WB (2009). Effect of microalgae *Chlorella vulgaris* on laying hen performance. Archiva Zootech. 12: 5-13.
- Holman BWB, Malau-aduli AEO (2012). Spirulina as a livestock supplement and animal feed. J. Anim. Physiol. Anim. Nutr. 97: 4. https://doi.org/10.1111/j.1439-0396.2012.01328.x
- Hynstova V, Sterbova D, Klejdus B, Hedbavny J, Huska D, Adam V (2018). Separation, identification and quantification of carotenoids and chlorophylls in dietary supplements containing *Chlorella vulgaris* and *Spirulina platensis* using High Performance Thin Layer Chromatography. J. Pharm. Biomed. Anal. 148: 108-118. https://doi.org/10.1016/j. jpba.2017.09.018
- •Janczyk P, Halle B, Souffrant WB (2009). Microbial community composition of the crop and ceca contents of



laying hens fed diets supplemented with *Chlorella vulgaris*. Poult. Sci. 88: 2324-2332. https://doi.org/10.3382/ps.2009-00250

- Jafari SMA, Rabbani M, Emtyazjoo M, Piryaei F (2014). Effect of dietary *Spirulina platensis* on fatty acid composition of rainbow trout (*Oncorhynchus mykiss*) fillet. Aquacult. Int. J. 22: 1307-1315. https://doi.org/10.1007/s10499-013-9748-0
- Kang HK, Salim HM, Akter N, Kim DW, Kim JH, Bang HT, Na JC, Hwangbo J, Choi HC, Kim MJ, Suh OS (2013). Effect of various forms of dietary *Chlorella* supplementation on growth performance, immune characteristics, and intestinal microflora population of broiler chickens. J. Appl. Poult. Res. 22: 100-108. https://doi.org/10.3382/japr.2012-00622
- •Kanagaraju P, Omprakash AV. (2016). Effect of *Spirulina platensis* algae powder supplementation as a feed additive on the growth performance of Japanese quails. Indian Vet. J. 93: 31-33.
- •Kaoud HA (2015). Effect of *Spirulina platensis* as a dietary supplement on broiler performance in comparison with prebiotics. J. Biol. Sci. 1: 1-6.
- Kaushik P, Chauhan A (2008). In vitro antibacterial activity of laboratory grown culture of Spirulina platensis. Indian J. Microbiol. 48: 348-352. https://doi.org/10.1007/s12088-008-0043-0
- Khan Z, Bhadouria P, Bisen PS (2005). Nutritional and therapeutic potential of *Spirulina*. Curr. Pharm. Biotechnol. 6: 373-379. https://doi.org/10.2174/138920105774370607
- •Kharde S, Shirbhate R, Bahiram K, Nipane S (2012). Effect of *Spirulina* supplementation on growth performance of broilers. Indian J. Vet. Res. 21: 66-69. http://iaavr.org/
- Kotrbacek V, Halouzka R, Jurajda V, Knotkova Z, Filka J (1994). Increased immune response in broilers after administration of natural food supplements. Vet. Med. (Parah). 39: 321-328.
- Kulshreshtha A, Jarouliya U, Bhadauriya P, Prasad G, Bisen P (2008). *Spirulina* in health care management. Curr. Pharm. Biotechnol. 9: 400-405. https://doi. org/10.2174/138920108785915111
- Kulshreshtha G, Rathgeber B, Stratton G, Thomas N, Evans F, Critchley A, Hafting J, Prithiviraj B (2014). Feed supplementation with red seaweeds, *Chondrus crispus* and *Sarcodiotheca gaudichaudii*, affects performance, egg quality, and gut microbiota of layer hens. Poult. Sci. 93: 2991-3001. https://doi.org/10.3382/ps.2014-04200
- Lee SH, Kang HJ, Lee HJ, Kang MH, Park YK (2010). Sixweek supplementation with *Chlorella* has favorable impact on antioxidant status in Korean male smokers. Nutrition. 26: 175-183. https://doi.org/10.1016/j.nut.2009.03.010
- Lemahieu C, Bruneel C, Termote-Verhalle R, Muylaert K, Buyse J, Foubert I (2013). Impact of feed supplementation with different omega-3 rich microalgae species on enrichment of eggs of laying hens. Food Chem. 141: 4051-4059. https:// doi.org/10.1016/j.foodchem.2013.06.078
- Lemahieu C, Bruneel C, Termote-Verhalle R, Muylaert K, Buyse J, Foubert I (2014). Effect of different microalgal n-3 PUFA supplementation doses on yolk color and n-3 LC-PUFA enrichment in the egg. Algal Res. 6: 119-123. https://doi.org/10.1016/j.algal.2014.10.007
- •Leng X, Hsu KN, Austic RE, Lei XG (2014). Effect of dietary defatted diatom biomass on egg production and quality of laying hens. J. Anim. Sci. Biotechnol. 6: 1-7.

### Advances in Animal and Veterinary Sciences

https://doi.org/10.1186/2049-1891-5-3

- Lipstein B, Hurwitz S (1981). The nutritional value of sewagegrown, alum-flocculated *Micractinium* algae in broiler and layer diets. Poult. Sci. 60: 2628-2638. https://doi. org/10.3382/ps.0602628
- Lipstein B, Hurwitz S (1983). The nutritional value of sewagegrown samples of *Chlorella* and *Micractinium* in broiler diets. Poult. Sci. 62: 1254-1260. https://doi.org/10.3382/ ps.0621254
- Lipstein B, Hurwitz S, Bornstein S (1980). The nutritional value of algae for poultry. Dried *Chlorella* in layer diets. Br. Poult. Sci. 21: 23-27. https://doi.org/10.1080/00071668008416631
- Lokapirnasari WP, Yulianto AB, Legowo D, Agustono (2016). The effect of *Spirulina* as feed additive to myocardial necrosis and leukocyte of chicken with avian influenza (H5N1) virus infection. Proc. Chem. 18: 213-217. http://repository. unair.ac.id/id/eprint/88517 https://doi.org/10.1016/j. proche.2016.01.033
- Lorenz RT, Cysewski GR (2000). Commercial potential for *Haematococcus* microalgae as a natural source of astaxanthin. Trends Biotechnol. 18: 160-167. https://doi.org/10.1016/ S0167-7799(00)01433-5
- •Mala R, Sarojini M, Saravanababu S, Umadevi G (2009). Screening for antimicrobial activity of crude extracts of *Spirulina platensis.* J. Cell Tissue Res. 9: 1951-1955.
- Mariey YA, Samak HR, Ibrahem MA (2012). Effect of using Spirulina platensis algae as a feed additive for poultry diets: 1- Productive and reproductive performances of local laying hens. Egypt. Poult. Sci. 32: 201-215.
- Mariey Y, Samak H, Abou-Khashba H, Sayed M, Abou-Zeid A (2014). Effect of using *Spirulina platensis* algae as a feed additives for poultry diets: 2 productive performance of broiler. Egypt. Poult. Sci. 34: 245-258.
- •Meineri G, Ingravalle F, Radice E, Aragno M, Peiretti PG (2009). Effects of high fat diets and *Spirulina platensis* supplementation in New Zealand white rabbits. J. Anim. Vet. Adv. 8: 2735-2744.
- Mobarez SM, Rizk AM, Abdel latif AM, Osama AE (2018). Effect of supplementing diet with *Spirulina platensis* algae or turmeric on productive and reproductive performance of golden Montazah layers. Egypt. Poult. Sci. J. 38: 109-125.
- Mohamed F (1998). Nutrition and Immunity on poultry. Egypt. Poult. Sci. J. 18: 443-448.
- Moradikor N, Mohamadi N (2015). The effects of different levels *Chlorell* microalgae on performance and immune response of laying hens under heat stress condition. Int. J. Life Sci. 9: 71-74. https://doi.org/10.3126/ijls.v9i2.12058
- Niamat ME, Ragaa AH (2017). Improved productivity and health of broiler chicken by micro green alga *Chlorella* vulgaris. Asian J. Poult. Sci. 11: 57-63. https://doi. org/10.3923/ajpsaj.2017.57.63
- Nikodémusz E, Paskai P, Tóth L, Kozák J (2010). Effect of dietary *Spirulina* supplementation on the reproductive performance of farmed pheasants. Technical Articles-Poultry Industry: 1-2. of some natural and commercial food products based on *Spirulina*. Analysis. 27: 533-540. https:// doi.org/10.1051/analusis:1999130
- Nuhu AA (2013). *Spirulina (Arthrospira*): An important source of nutritional and medicinal compounds. J. Mar. Biol. 1: 1-8. https://doi.org/10.1155/2013/325636
- Oh ST, Zheng L, Kwon HJ, Choo YK, Lee KW, Kang CW, An BK (2015). Effects of dietary fermented *Chlorella* vulgaris (CBT<sup>®</sup>) on growth performance, relative organ

#### **Advances in Animal and Veterinary Sciences**

## **OPEN OACCESS**

weights, cecal microflora, tibia bone characteristics, and meat qualities in Pekin ducks. Asian Australas. J. Anim. Sci. 28: 95-101. https://doi.org/10.5713/ajas.14.0473

- Park JH, Lee SI, Kim IH (2018). Effect of dietary Spirulina (Arthrospira) platensis on the growth performance, antioxidant enzyme activity, nutrient digestibility, cecal microflora, excretanoxious gas emission, and breast meat quality of broiler chickens. Poult. Sci. 97: 2451-2459. https://doi.org/10.3382/ps/pey093
- Park JH, Upadhaya SD, Kim IH (2015). Effect of dietary marine microalgae (*Schizochytrium*) powder on egg production, blood lipid profiles, egg quality, and fatty acid composition of egg yolk in layers. Asian-Australas. J. Anim. Sci. 28: 391-397. https://doi.org/10.5713/ajas.14.0463
- Parpinello GP, Meluzzi A, Sirri F, Tallarico N, Versri A (2006). Sensory evaluation of egg products and eggs laid from hens fed diets with different fatty acid composition and supplemented with antioxidants. Food Res. Int. 39: 47-52. https://doi.org/10.1016/j.foodres.2005.05.010
- Pulz O, Gross W (2004). Valuable products from biotechnology of microalgae. Appl. Microbiol. Biotechnol. 65: 635-648. https://doi.org/10.1007/s00253-004-1647-x
- •Qureshi M, Garlich D, Kidd M, Ali R (1994). Immune enhancement potential of *Spirulina platensis* in chickens. Poult. Sci. 73: 46.
- Qureshi M, Garlich J, Kidd M (1996). Dietary Spirulina platensis enhances humoral and cell-mediated immune functions in chickens. Immunopharmacol. Immunotoxicol. 18: 465-476. https://doi.org/10.3109/08923979609052748
- Raju MVLN, Rao SV, Radhika K, Chawak MM (2004). Effects of *Spirulina platensis* or furazolidone on the performance and immune response of broiler chickens fed with aflatoxin contaminated diet. Indian J. Anim. Nutr. 21: 40-44.
- Raju MVLN, Rao SV, Radhika K, Chawak MM (2005). Dietary supplementation of *Spirulina* and its effects on broiler chicken exposed to aflatoxicosis. Indian J. Poult. Sci. 40: 36-40.
- Rania MA, Hala MT (2008). Antibacterial and antifungal activity of cyanobacteria and green microalgae. Evaluation of medium components by placket-burman design for antimicrobial activity of *Spirulina platensis*. Glob. J. Biotechnol. Biochem. 3: 22-31.
- Rezvani M, Zaghari M, Moravej H (2012). A survey on *Chlorella vulgaris* effect's on performance and cellular immunity in broilers. Int. J. Agric. Sci. Res. 3: 9-15.
- Rizzi L, Bochocchio D, Bargellini A, Parazza P, Simiolo M (2009). Effects of dietary microalgae, other lipid sources, inorganic selenium and iodine on yolk n-3 fatty acid composition, selenium content and quality of eggs in laying hens. J. Sci. Food Agric. 89: 1775-1781. https://doi. org/10.1002/jsfa.3655
- Ross E, Dominy W (1985). The effect of dehydrated *Spirulina platensis* on poultry. Poult. Sci. 64: 173.
- Ross E, Dominy W (1990). The nutritional value of dehydrated, blue-green algae (*Spirulina plantensis*) for poultry. Poult. Sci. 69: 794-800. https://doi.org/10.3382/ps.0690794
- Saxena PN, Ahmad MR, Shyam R, Amla DV (1983). Cultivation of *Spirulina* in sewage for poultry feed. Experientia. 39: 1077–1083. https://doi.org/10.1007/BF01943117
- Schiavone A, Chiarini R, Marzoni M, Castillo A, Tassone S, Romboli I (2007). Breast meat traits of Muscovy ducks fed on a microalgae *Crypthecodinium cohnii* meal supplemented diet. Br. Poult. Sci. 48: 573-579. https://

#### doi.org/10.1080/00071660701615796

- Shanmugapriya B, Babu SS (2014). Supplementary effect of Spirulina platensis on performance, hematology and carcass yield of broiler chicken. Indian Streams Res. J. 4: 1-7.
- Shanmugapriya B, Babu SS, Hariharan T, Sivaneswaran S, Anusha MB (2015). Dietary administration of *Spirulina platensis* as probiotics on growth performance and histopathology in broiler chicks. Int. J. Recent Sci. Res. 6: 2650-2653.
- Shokri H, Khosravi A, Taghavi M (2014). Efficacy of *Spirulina platensis* on immune functions in cancer mice with systemic candidiasis. J. Mycol. Res. 1: 7-13.
- Soni R, Sudhakar K, Rana R (2017). Spirulina From growth to nutritional product: A review. Trends Food Sci. Technol. 69: 157-171. https://doi.org/10.1016/j.tifs.2017.09.010
- Sørum H, Sunde M (2001). Resistance to antibiotics in the normal flora of animals. Vet. Res. 32: 227-241. https://doi. org/10.1051/vetres:2001121
- Spolaore P, Joannis-Cassan C, Duran E, Isambert A (2006). Commercial applications of microalgae. J. Biosci. Bioeng. 101: 87-96. https://doi.org/10.1263/jbb.101.87
- Sugiharto S, Lauridsen C (2016). Dietary *Chlorella* supplementation effect on immune responses and growth performances of broiler chickens exposed to post hatch holding time. Livest. Res. Rural Dev. 28: 7.
- Sugiharto S, Yudiarti T, Isroli I, Widiastuti E (2018). Effect of feeding duration of *Spirulina platensis* on growth performance, haematological parameters, intestinal microbial population and carcass traits of broiler chicks. S. Afr. J. Anim. Sci. 48: 98-107. https://doi.org/10.4314/sajas. v48i1.12
- Sujatha T, Narahari D (2011). Effect of designer diets on egg yolk composition of 'White Leghorn' hens. J. Food Sci. Technol. 48: 494-497. https://doi.org/10.1007/s13197-010-0132-z
- Tokusoglu Ö, Üunal MK (2003). Biomass nutrient profiles of three microalgae: Spirulina platensis, Chlorella vulgaris, and Isochrisis galbana. J. Food Sci. 68: 1144-1148. https://doi. org/10.1111/j.1365-2621.2003.tb09615.x
- Toyomizu M, Sato K, Taroda H, Kato T, Akiba Y (2001). Effects of dietary *Spirulina* on meat colour in muscle of broiler chickens. Br. Poult. Sci. 42: 197-202. https://doi. org/10.1080/00071660120048447
- Trziszka T, Łukaszewicz E, Bobak Ł, Kowalczyk A, Adamski M, Dobrzański Z (2014). Effect of enriching feeds with algae marine and linseed on morphological composition and physical and chemical characteristics of Japanese quail eggs. Żywność. Nauka. Technologia. Jakość. 97: 138-149. https:// doi.org/10.15193/zntj/2014/97/138-149
- Tsuchihashi N, Watanabe T, Takai Y (1987). Effect of *Spirulina platensis* on caecum content in rats. Bull. Chiba Hyg. College. 7: 27-30.
- Uyisenga JP, Nzayino P, Seneza R, Hishamunda L, Uwantege K, Gasana N, Emmanuel SB (2010). *In vitro* study of antibacterial and antifungal activity of *Spirulina platensis*. Int. J. Ecol. Dev. 16: 80-88.
- Venkataraman LV, Somasekaran T, Becker EW (1994). Replacement value of blue green alga (*Spirulina platensis*) for fishmeal and a vitamin-mineral premix for broiler chicks. Br. Poult. Sci. 35: 373-381. https://doi. org/10.1080/00071669408417702
- •Vonshak A. (2002). Spirulina platensis (Arthrospira) physiology, cell-biology and biotechnology. Taylor and

### Advances in Animal and Veterinary Sciences

# OPENOACCESS

Francis Press.

- Waldenstedt L, Inborr J, Hannson I, Elwinger K (2003). Effects of astaxanthin-rich algal meal (*Haematococcus pluvalis*) on growth performance, caecal *Campylobacter* and Clostridial counts and tissue astaxanthin concentration of broiler chickens. Anim. Feed Sci. Technol. 108: 119-132. https:// doi.org/10.1016/S0377-8401(03)00164-0
- Widya PL, Andreas BY, Djoko LA (2016). The effect of Spirulina as feed additive to mocardial necrosis and leukocyte of chicken with avian influenza (H5N1) virus infection. Procedia Chem. 18: 213-217. https://doi.org/10.1016/j. proche.2016.01.033
- Yaakob Z, Ali E, Zainal A, Mohamad M, Takriff MS (2014). An overview: Biomolecules from microalgae for animal feed and aquaculture. J. Biol. Res. Thessalon. 21: 1. https://doi. org/10.1186/2241-5793-21-6
- Yan L, Kim IH (2013). Effects of dietary ω-3 fatty acid-enriched microalgae supplementation on growth performance, blood profiles, meat quality, and fatty acid composition of meat in broilers. J. Appl. Anim. Res. 41: 392-397. https://doi.org/10 .1080/09712119.2013.787361
- Yoshida M, Hoshi H (1980). Nutritive value of *Spirulina*, green algae, for poultry feed. Japn. Poult. Sci. 17: 27-30. https:// doi.org/10.2141/jpsa.17.27
- •Yusuf MS, Hassan MA, Abdel-Daim MM, El Nabtiti AS,

Ahmed AM, Moawed SA, El-Sayed, AK, Cui H (2016). Value added by *Spirulina platensis* in two different diets on growth performance, gut microbiota, and meat quality of Japanese quails. Vet. World. 9: 1287-1293. https://doi. org/10.14202/vetworld.2016.1287-1293

- Zahroojian N, Moravej H, Shivazad M (2011). Comparison of marine algae (*Spirulina platensis*) and synthetic pigment in enhancing egg yolk colour of laying hens. Br. Poult. Sci. 52: 584-588. https://doi.org/10.1080/00071668.2011.610779
- Zahroojian N, Moravej H, Shivazad M (2013). Effects of dietary marine algae (*Spirulina platensis*) on egg quality and production performance of laying hens. J. Agric. Sci. Technol. 15: 1353-1360.
- Zeweil H, Abaza, IM, Zahran SM, Ahmed MH, Haiam M. AboulElaand, Asmaa AS (2016). Effect of *Spirulina platensisas* dietary supplement on some biological traits for chickens under heat stress condition. Asian J. Biomed. Pharma. Sci. 6: 8-12.
- Zheng L, Oh ST, Jeon JY, Moon BH, Kwon HS, Lim SU, An BK, Kang CW (2012). The dietary effects of fermented *Chlorella vulgaris* (CBT) on production performance, liver lipids and intestinal microflora in laying hens. Asian-Australas. J. Anim. Sci. 25: 261-266. https://doi. org/10.5713/ajas.2011.11273