Neonatal Gnotobiotic Pig Models for Studying Viral Pathogenesis, Immune Responses, and for Vaccine Evaluation

Xingdong Yang and Lijuan Yuan*

Department of Biomedical Sciences and Pathobiology, Virginia–Maryland Regional College of Veterinary Medicine, Virginia Polytechnic Institute and State University, Blacksburg, VA 24061-0913, USA.

Abstract | The definition of gnotobiotic pig is a pig in which only certain known strains of virus or bacterium are present through deliberate inoculation with a particular virus strain or bacterial species and being maintained free of unwanted microbiota in sterile isolators. The sterile surgical derivation and gnotobiotic status of the pigs allow studies of the infection, disease and immune responses caused by a specific pathogen in the absence of interfering maternal antibodies, other maternal immune regulators, and intestinal and environmental microbes. Due to the similarities between pigs and humans in terms of genetics, physiology, intestinal anatomy, and immune system, pig is particularly a suitable animal species for the modeling of human enteric pathogen infections, immune responses and for vaccine evaluations. These are the basis for the successful establishment of the infection and disease models for human rotavirus, human norovirus and human enterovirus 71 using neonatal gnotobiotic pigs. The differences between germ-free and normal animals in the maturation status of immune systems caused by the lack of gut microbiota can be minimized by establishing human-gut-microbiota transplanted gnotobiotic pig models. Given the advantages of gnotobiotic pig models, it is expected that they will be used more widely in biomedical research for studies of human viruses and other human diseases.

Editor | Muhammad Munir, The Pirbright Institute, Compton Laboratory, UK Received | July 29, 2014; Accepted | August 7, 2014; Published | September 7, 2014 *Correspondence | Lijuan Yuan, Virginia Polytechnic Institute and State University, USA; E-mail | lyuan@vt.edu Citation | Yang, X. and Yuan, L. (2014). Neonatal gnotobiotic pig models for studying viral pathogenesis, immune responses, and for vaccine evaluation. *British Journal of Virology*, 1(3): 87-91.

In a recent study published in the May issue of the Emerging Microbes and Infections journal (Yang, et al., 2014) we demonstrated that neonatal gnotobiotic (Gn) pigs infected orally or combined oral-nasally with non-pig adapted human enterovirus 71 strain BJ110 (C4 genotype) resulted in the virus shedding pattern, clinical signs, pathology, and immune responses similar to those seen in human patients. Virus shedding in the fecal samples was detected from post-inoculation day (PID) 1 to PID 21. Fever, one of the most common clinical signs in human patients, was induced in the combined oral-nasally infected pigs on PID 4 and PID 6. High titers of neutraliz-

ing serum antibodies against EV71, and strong IFN- γ producing CD4+ and CD8+ T cell responses were generated. Although no severe pathology was observed in tissues of intestines, respiratory and central nervous systems, notable respiratory and neurological signs were present in the infected neonatal Gn pigs, especially those infected through combined oral-nasal infection route. In particular, in contrast to other animal models for the human enterovirus 71 infections in mice and monkeys, Gn pigs were infected through the natural route of infection, namely, oral route, by an original human enterovirus 71 strain isolated from human patients and produced the clinical signs seen only in mice or monkeys infected through non-natural routes of infections (Zhang, et al., 2011), in immunocompromised animals (Caine, et al., 2013), transgenic animals (Fujii, et al., 2013; Lin, et al., 013), or using adapted virus strains (Wang, et al., 2011; Wang, et al., 2004). Based on our results, we concluded that neonatal Gn pig model for human enterovirus 71 represents an excellent alternative animal model to the current mice and monkey models for virus pathogenesis study and vaccine development.

Pigs are increasingly being used to study various human diseases. Given their similarities to humans in terms of genetics, physiology, intestinal anatomy, and immune system, pigs are particularly good animal species for the modeling of human enteric pathogen infections and immune responses (Meurens, et al., 2012). The sterile surgical derivation and gnotobiotic status of the pigs allow studies of the infection, disease and immune responses caused by a specific enteric pathogen in the absence of interfering maternal antibodies, other maternal immune regulators, and intestinal and environmental microbes. These are the basis for us to successfully establish the pig models for human rotavirus, human norovirus and human enterovirus 71 infection and disease using neonatal gnotobiotic pigs (Bui, et al., 2013; Kocher, et al., 2014; Ward, et al., 1996; Yang, et al., 2014; Yuan, et al., 2002). Gn pigs are excellent model for the study of viral pathogenesis. Due to the similarities of pigs and humans as well as the lack of confounding commensal microbes that are present in all specific pathogen free (SPF) animals, Gn pigs can be used to identify the pathological changes directly as the result of a particular virus infection. Virus replication, spread and the resulting tissue damages and clinical signs can be more accurately recapitulated compared to the SPF animal models. The mechanisms of the host responses and the pathology associated with the virus infection can also be more clearly identified. Thus, Gn pigs are ideal for the identification of causative agents of viral diseases and mechanisms of viral pathogenesis.

The results from Gn pig models are generally comparable to SPF animal models, even though some discrepancies do exist. The presence of certain microbial species in the intestinal environment may impact the infectivity of viruses. We found that human rotavirus replicates to higher titers in Gn pigs colonized with *Lactobacillus acidophilus and L. reuteri* compared to germ-free pigs (Zhang, et al., 2008a; 2008b). Sever-

al other studies have also shown that gut microbiota enhanced the replication or virus entry of enteric viruses and their pathogenesis (Kane, et al., 2011; Kuss, et al., 2011; Uchiyama, et al., 2014) and that elimination of microbiota delayed rotavirus infection and significantly reduced rotavirus infectivity in mice (Uchiyama, et al., 2014). On the other hand, certain species of the normal gut microbes can be protective against virus-induced tissue damages and clinical signs. *Lactobacillus rhamnosus* GG (LGG) has been found to reduce human rotavirus diarrhea in Gn pigs by protecting against virus induced tissue injury in the intestine (Liu, et al., 2013).

Discrepancies between the Gn pigs and SPF animal models mainly results from the lack of gut microbiota. It is well known that gut microbiota interact with host immune system and modulate their responses to viral infections. Such discrepancies can be minimized by establishing human gut microbiota (HGM) transplanted Gn pig models (Wen, et al., 2014). Similar to rotavirus infection of the Lactobacillus spp monoassociated Gn pigs, HGM Gn pigs shed higher titer of the virus after human rotavirus inoculation compared to the germ-free pigs (Wen, et al., 2014). Because gut microbiota play significant roles in shaping neonatal immune system and host susceptibility to enteric pathogens (Chung, et al., 2012), HGM transplanted Gn pigs may be a better animal model for certain human infectious diseases than germ-free and SPF animal models.

Gn pigs are particularly suited for the study of specific immune responses to virus stimulus. Gn pigs have been widely used to study immune responses to a variety of enteric viruses, including rotavirus (Azevedo, et al., 2012; Liu, et al., 2014; Nguyen, et al., 2003; Nguyen, et al., 2006a; Nguyen, et al., 2006b, Parreno, et a., 1999, Wen, et al., 2009; Wen et al., 2012a; 2012b; Wen, et al., 2014; Yuan, et al., 2005; 2004a; 2001; 2004b; 2002; 2008; 2013; Zhang, et al., 2008a; 2008b; 2008c) norovirus (Kocher, et al., 2014; Souza, et al., 2007), and enterovirus 71 (Yang, et al., 2014). These models have been extremely useful in understanding both innate and adaptive immune responses and for evaluations of the immunogenicity and protective efficacy of various vaccine formulations, adjuvants and immunization routes. Again, HGM transplanted Gn pigs can be used to introduce the effects of gut microbiota on the host immune responses to viruses and vaccines. Using HGM Gn pigs, our previous study



has shown that transplanted human gut bacteria can modulate host immune responses to human rotavirus infection and vaccination (Wen, et al., 2014).

In conclusion, given the advantages of Gn pig models, it is expected that Gn pigs will be used more widely for the study of viral pathogenesis and immune responses that are specific to a virus. Similarly, Gn pigs can also be used to study pathogenesis or immune responses to other types of stimulus, be it microbes, chemical, or physical. Currently, the applications of Gn pigs in modeling human diseases are limited by the cost and relative lack of research infrastructure and reagents. However, these limitations are becoming less prominent as the scientific community is increasingly interested in the Gn pig models and these models become more commonly used in biomedical researches.

References

- Azevedo, M. S., W. Zhang, K. Wen, A. M. Gonzalez, L. J. Saif, A. E. Yousef, and L. Yuan. 2012. Lactobacillus acidophilus and Lactobacillus reuteri modulate cytokine responses in gnotobiotic pigs infected with human rotavirus. Benef Microbes 3:33-42.
- Bui, T., J. Kocher, Y. Li, K. Wen, G. Li, F. Liu, X. Yang, T. LeRoith, M. Tan, M. Xia, W. Zhong, X. Jiang, and L. Yuan. 2013. Median infectious dose of human norovirus GII.4 in gnotobiotic pigs is decreased by simvastatin treatment and increased by age. J Gen Virol 94:2005-2016.
- Caine, E. A., C. D. Partidos, J. D. Santangelo, and J. E. Osorio. 2013. Adaptation of enterovirus 71 to adult interferon deficient mice. PloS one 8:e59501.
- Chung, H., S. J. Pamp, J. A. Hill, N. K. Surana, S. M. Edelman, E. B. Troy, N. C. Reading, E. J. Villablanca, S. Wang, J. R. Mora, Y. Umesaki, D. Mathis, C. Benoist, D. A. Relman, and D. L. Kasper. 2012. Gut immune maturation depends on colonization with a host-specific microbiota. Cell 149:1578-1593.
- Fujii, K., N. Nagata, Y. Sato, K. C. Ong, K. T. Wong, S. Yamayoshi, M. Shimanuki, H. Shitara, C. Taya, and S. Koike. 2013. Transgenic mouse model for the study of enterovirus 71 neuropathogenesis. Proceedings of the National Academy of Sciences of the United States of America 110:14753-14758.
- Kane, M., L. K. Case, K. Kopaskie, A. Kozlova, October 2014 | Volume 1 | Issue 3 | Page 89

C. MacDearmid, A. V. Chervonsky, and T. V. Golovkina. 2011. Successful transmission of a retrovirus depends on the commensal microbiota. Science 334:245-249.

- Kocher, J., T. Bui, E. Giri-Rachman, K. Wen, G. Li, X. Yang, F. Liu, M. Tan, M. Xia, W. Zhong, X. Jiang, and L. Yuan. 2014. Intranasal P particle vaccine provided partial cross-variant protection against human GII.4 norovirus diarrhea in gnotobiotic pigs. J Virol.
- Kuss, S. K., G. T. Best, C. A. Etheredge, A. J. Pruijssers, J. M. Frierson, L. V. Hooper, T. S. Dermody, and J. K. Pfeiffer. 2011. Intestinal microbiota promote enteric virus replication and systemic pathogenesis. Science 334:249-252.
- Lin, Y.-W., S.-L. Yu, H.-Y. Shao, H.-Y. Lin, C.-C. Liu, K.-N. Hsiao, E. Chitra, Y.-L. Tsou, H.-W. Chang, C. Sia, P. Chong, and Y.-H. Chow. 2013. Human SCARB2 transgenic mice as an infectious animal model for enterovirus 71. PLoS One 8:e57591.
- Liu, F., Li, G., Wen, K., Wu, S., Zhang, Y., Bui, T., Yang, X., Kocher, J., Sun, J., Jortner, B., Yuan, L., 2013. Lactobacillus rhamnosus GG on rotavirus-induced injury of ileal epithelium in gnotobiotic pigs. J Pediatr Gastroenterol Nutr 57, 750-758.
- Liu, F., K. Wen, G. Li, X. Yang, J. Kocher, T. Bui, D. Jones, K. Pelzer, S. Clark-Deener, and L. Yuan. 2014. Dual Functions of Lactobacillus acidophilus NCFM as Protection Against Rotavirus Diarrhea. J Pediatr Gastroenterol Nutr 58:171-178.
- Meurens, F., A. Summerfield, H. Nauwynck, L. Saif, and V. Gerdts. 2012. The pig: a model for human infectious diseases. Trends Microbiol 20:50-57.
- Nguyen, T. V., C. Iosef, K. Jeong, Y. Kim, K. O. Chang, K. Lovgren-Bengtsson, B. Morein, M. S. Azevedo, P. Lewis, P. Nielsen, L. Yuan, and L. J. Saif. 2003. Protection and antibody responses to oral priming by attenuated human rotavirus followed by oral boosting with 2/6-rotavirus-like particles with immunostimulating complexes in gnotobiotic pigs. Vaccine 21:4059-4070.
- Nguyen, T. V., L. Yuan, M. S. Azevedo, K. I. Jeong, A. M. Gonzalez, C. Iosef, K. Lovgren-Bengtsson, B. Morein, P. Lewis, and L. J. Saif. 2006a. High titers of circulating maternal antibodies suppress effector and memory B-cell responses induced by an attenuated rotavirus priming and rotavirus-like particle-immunostimulating complex boosting

vaccine regimen. Clin Vaccine Immunol 13:475-485.

- Nguyen, T. V., L. Yuan, P. A. MS, K. I. Jeong, A. M. Gonzalez, C. Iosef, K. Lovgren-Bengtsson, B. Morein, P. Lewis, and L. J. Saif. 2006b. Low titer maternal antibodies can both enhance and suppress B cell responses to a combined live attenuated human rotavirus and VLP-ISCOM vaccine. Vaccine 24:2302-2316.
- Parreno, V., D. C. Hodgins, L. de Arriba, S. Y. Kang, L. Yuan, L. A. Ward, T. L. To, and L. J. Saif. 1999. Serum and intestinal isotype antibody responses to Wa human rotavirus in gnotobiotic pigs are modulated by maternal antibodies. J Gen Virol 80 (Pt 6):1417-1428.
- Souza, M., V. Costantini, M. S. P. Azevedo, and L. J. Saif. 2007. A human norovirus-like particle vaccine adjuvanted with ISCOM or mLT induces cytokine and antibody responses and protection to the homologous GII.4 human norovirus in a gnotobiotic pig disease model. Vaccine 25:8448-8459.
- Uchiyama, R., B. Chassaing, B. Zhang, and A. T. Gewirtz. 2014. Antibiotic treatment suppresses rotavirus infection and enhances specific humoral immunity. J Infect Dis 210:171-82.
- Wang, W., J. Duo, J. Liu, C. Ma, L. Zhang, Q. Wei, and C. Qin. 2011. A mouse muscle-adapted enterovirus 71 strain with increased virulence in mice. Microbes and infection / Institut Pasteur 13:862-870.
- Wang, Y. F., C. T. Chou, H. Y. Lei, C. C. Liu, S. M. Wang, J. J. Yan, I. J. Su, J. R. Wang, T. M. Yeh, S. H. Chen, and C. K. Yu. 2004. A mouse-adapted enterovirus 71 strain causes neurological disease in mice after oral infection. Journal of virology 78:7916-7924.
- Ward, L. A., B. I. Rosen, L. Yuan, and L. J. Saif. 1996. Pathogenesis of an attenuated and a virulent strain of group A human rotavirus in neonatal gnotobiotic pigs. J Gen Virol 77 (Pt 7):1431-1441.
- Wen, K., M. S. Azevedo, A. Gonzalez, W. Zhang, L. J. Saif, G. Li, A. Yousef, and L. Yuan. 2009. Toll-like receptor and innate cytokine responses induced by lactobacilli colonization and human rotavirus infection in gnotobiotic pigs. Vet Immunol Immunopathol 127:304-315.
- Wen, K., G. Li, T. Bui, F. Liu, Y. Li, J. Kocher, L. Lin, X. Yang, and L. Yuan. 2012a. High dose and low dose Lactobacillus acidophilus exerted differential immune modulating effects on T

cell immune responses induced by an oral human rotavirus vaccine in gnotobiotic pigs. Vaccine 30:1198-1207.

- Wen, K., G. Li, X. Yang, T. Bui, M. Bai, F. Liu, J. Kocher, and L. Yuan. 2012b. CD4+ CD25-FoxP3+ regulatory cells are the predominant responding regulatory T cells after human rotavirus infection or vaccination in gnotobiotic pigs. Immunology 137:160-171.
- Wen, K., C. Tin, H. Wang, X. Yang, G. Li, E. Giri-Rachman, J. Kocher, T. Bui, S. Clark-Deener, and L. Yuan. 2014. Probiotic Lactobacillus rhamnosus GG Enhanced Th1 Cellular Immunity but Did Not Affect Antibody Responses in a Human Gut Microbiota Transplanted Neonatal Gnotobiotic Pig Model. PLoS ONE 9:e94504.
- Yang, X., G. Li, K. Wen, T. Bui, F. Liu, J. Kocher, B. Jortner, M. Vonck, K. Pelzer, J. Deng, R. Zhu, Y. Li, Y. Qian, and L. Yuan. 2014. A neonatal gnotobiotic pig model of human enterovirus 71 infection and associated immune responses. Emerging Microbes & Infections 3:e35. doi:10.1038/ emi.2014.35.
- Yuan, L., M. S. Azevedo, A. M. Gonzalez, K. I. Jeong, T. Van Nguyen, P. Lewis, C. Iosef, J. E. Herrmann, and L. J. Saif. 2005. Mucosal and systemic antibody responses and protection induced by a prime/boost rotavirus-DNA vaccine in a gnotobiotic pig model. Vaccine 23:3925-3936.
- Yuan, L., S. Honma, S. Ishida, X. Y. Yan, A. Z. Kapikian, and Y. Hoshino. 2004a. Species-specific but not genotype-specific primary and secondary isotype-specific NSP4 antibody responses in gnotobiotic calves and piglets infected with homologous host bovine (NSP4[A]) or porcine (NSP4[B]) rotavirus. Virology 330:92-104.
- Yuan, L., C. Iosef, M. S. Azevedo, Y. Kim, Y. Qian, A. Geyer, T. V. Nguyen, K. O. Chang, and L. J. Saif. 2001. Protective immunity and antibody-secreting cell responses elicited by combined oral attenuated Wa human rotavirus and intranasal Wa 2/6-VLPs with mutant Escherichia coli heat-labile toxin in gnotobiotic pigs. J Virol 75:9229-9238.
- Yuan, L., S. Ishida, S. Honma, J. T. Patton, D. C. Hodgins, A. Z. Kapikian, and Y. Hoshino. 2004b. Homotypic and heterotypic serum isotype-specific antibody responses to rotavirus nonstructural protein 4 and viral protein (VP) 4, VP6, and VP7 in infants who received selected live oral rotavirus vaccines. J Infect Dis 189:1833-1845.



- Yuan, L., and L. J. Saif. 2002. Induction of mucosal immune responses and protection against enteric viruses: rotavirus infection of gnotobiotic pigs as a model. Vet Immunol Immunopathol 87:147-160.
- Yuan, L., K. Wen, M. S. Azevedo, A. M. Gonzalez, W. Zhang, and L. J. Saif. 2008. Virus-specific intestinal IFN-gamma producing T cell responses induced by human rotavirus infection and vaccines are correlated with protection against rotavirus diarrhea in gnotobiotic pigs. Vaccine 26:3322-3331.
- Yuan, L., K. Wen, F. Liu, and L. G. 2013. Dose Effects of LAB on Modulation of Rotavirus Vaccine Induced Immune Responses. *In* J. M. Kongo (ed.), Lactic Acid Bacteria - R & D for Food, Health and Livestock Purposes. InTech, http:// www.intechopen.com/books/lactic-acid-bacteria-r-d-for-food-health-and-livestock-purposes/ dose-effects-of-lab-on-modulation-of-rotavirus-vaccine-induced-immune-responses.
- Zhang, W., M. S. Azevedo, A. M. Gonzalez, L.J. Saif, T. Van Nguyen, K. Wen, A. E. Yousef, andL. Yuan. 2008a. Influence of probiotic Lactoba-

cilli colonization on neonatal B cell responses in a gnotobiotic pig model of human rotavirus infection and disease. Vet Immunol Immunopathol 122:175-181.

- Zhang, W., M. S. Azevedo, K. Wen, A. Gonzalez, L. J. Saif, G. Li, A. E. Yousef, and L. Yuan. 2008b. Probiotic Lactobacillus acidophilus enhances the immunogenicity of an oral rotavirus vaccine in gnotobiotic pigs. Vaccine 26:3655-3661.
- Zhang, W., K. Wen, M. S. Azevedo, A. Gonzalez, L. J. Saif, G. Li, A. E. Yousef, and L. Yuan. 2008c. Lactic acid bacterial colonization and human rotavirus infection influence distribution and frequencies of monocytes/macrophages and dendritic cells in neonatal gnotobiotic pigs. Vet Immunol Immunopathol 121:222-231.
- Zhang, Y., W. Cui, L. Liu, J. Wang, H. Zhao, Y. Liao, R. Na, C. Dong, L. Wang, Z. Xie, J. Gao, P. Cui, X. Zhang, and Q. Li. 2011. Pathogenesis study of enterovirus 71 infection in rhesus monkeys. Laboratory investigation; a journal of technical methods and pathology 91:1337-1350.

