

Detection of some Pathogenic Bacteria Causing Sinusitis in Turkeys by Using Multiplex PCR

HANAN SAAD EL-SAMAHY, AMANI ABD EL-NABY HAFEZ, MOHAMED TALAT RAGAB, DISOUKY MOHAMED MOURAD*

Department of Animal and Poultry Health, Division of Animal and Poultry Production, Desert Research Center, Ministry of Agriculture, 1-Mathaf ElMateria Street, Cairo, 2633759, Egypt.

Abstract | In Egypt, turkey sinusitis is a major phenomenon that make the turkey farmers take far away from the turkey production as a result of difficult treatment, medication cost, difficult isolation of causative bacterial agents, weight loss, reduced fertility, and hatchability, so this study aimed to determine the bacterial causative agents using multiplex PCR as an accurate rapid diagnostic technique. Exudates of swollen infra-orbital sinus were aspirated and aseptically collected from affected turkeys of seventeen flocks and examined by multiplex PCR against *Escherichia coli (E. coli)*, *Mycoplasma gallisepticum (M. gallisepticum)*, *Mycoplasma meleagridis (M. meleagradis)*, *Ornithobacterium rhinotracheale* (ORT), *Pasteurella multocida (P. multocida)*, and *Pseudomonas aeruginosa (P. aeruginosa)*. The most prevalent detected bacteria were *E coli* (13) followed by *M. gallisepticum* (12), ORT (12) and *M. meleagridis* (7). Two farms reported single infection, one with *M. gallisepticum* and one with *E. coli*. Only one farm had no infection while mixed infection recorded in 14 farms. *P. multocida* and *P. aeruginosa* were not detected. It was concluded that Balady breed had a higher resistance than other breeds, *E. coli* and/or *M. gallisepticum* were the primary cause of turkey sinusitis, *M. meleagridis* recorded the lowest incidence and frequency in examined turkey flocks, while ORT had a higher incidence and frequency similar to *M. gallisepticum*. It was necessary to apply further molecular studies on these pathogens to control and avoid their spread in turkey and other poultry flocks.

Keywords | Bacteria, Multiplex PCR, Sinusitis, Turkey

Received | April 25, 2022; Accepted | May 30, 2022; Published | August 20, 2022

*Correspondence | Disouky Mohamed Mourad, Department of Animal and Poultry Health, Division of Animal and Poultry Production, Desert Research Center, Ministry of Agriculture, 1-Mathaf ElMateria Street, Cairo, 2633759, Egypt; Email: dismou235@hotmail.com

Citation | El-Samahy HS, Hafez AAN, Ragab MT, Mourad DM (2022). Detection of some pathogenic bacteria causing sinusitis in turkeys by using multiplex pcr. Adv. Anim. Vet. Sci. 10(9): 1962-1968.

DOI | http://dx.doi.org/10.17582/journal.aavs/2022/10.9.1962.1968 ISSN (Online) | 2307-8316



Copyright: 2022 by the authors. Licensee ResearchersLinks Ltd, England, UK. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons. org/licenses/by/4.0/).

INTRODUCTION

A vian respiratory lesion including sinusitis, air saculitis, tracheitis and pneumonia was considered a complex problem occurred where different microorganisms coupled with the sublevel of hygienic measures and simultaneously interacted (OIE, 2008). In turkeys the respiratory lesions were restricted to swollen infra-orbital sinuses contained mucoid to caseous exudate (OIE, 2018) causing several

economic losses with reduced feed intake, body weight, egg production and hatchability (Ali and Youssef, 2003; Alessandri et al., 2005; Bradbury, 2006, Büyüktanir et al., 2008; Nicholas et al., 2009).

Sometimes, infectious sinusitis in turkey accompanied by occasional movement disorders due to the inflammation of ankle and shoulder joints (Hafez 2002).

<u>open∂access</u>

Advances in Animal and Veterinary Sciences

Table 1: History of collected samples from turkey sinusitis

Farm No.	Area	Governorates	species	Breed	date
1	Kilo28 North coast	Alexanderia	Turkey	White Nicholas	11/5/2020
2	Hawaria	Alexanderia	Turkey	balady	22/5/2020
3	Naseria-1	Alexanderia	Turkey	Bronze	27/8/2020
4	Amria-K	Alexanderia	Turkey	balady	13/6/2020
5	Abees-1	Alexanderia	Turkey	Balady	9/8/2020
6	Kilo21 North coast	Alexanderia	Turkey	Balady	9/6/2020
7	Amria-A	Alexanderia	Turkey	Bronze	25/7/2020
8	Nobaria	Alexanderia	Turkey	Balady	20/10/2020
9	Abees-2	Alexanderia	Turkey	Bronze	15/11/2020
10	Naseria-2	Alexanderia	Turkey	Bronze	6/12/2020
11	Nobaria	Alexanderia	Turkey	Bronze	23/12/2020
12	Kilo35Desert road	Alexanderia	Turkey	Bronze	28/12/2020
13	Kilo48North coast	Alexanderia	Turkey	Balady	3/1/2021
14	Kilo59Desert road	Alexanderia	Turkey	White Nicholas	12/1/2021
15	Maryout-1	Alexanderia	Turkey	Bronze	15/1/2021
16	Maryout-2	Alexanderia	Turkey	Bronze	15/1/2021
17	Maryout-3	Alexanderia	Turkey	Bronze	15/1/2021

Not only *Mycoplasma* spp. but also other pathogens as *E. coli*, *P. aeruginosa*, *Staphylococcus aureus* (*S. aureus*) and *Salmonella gallinarun pullorum* (*S. gallinarun pullorum*) were isolated from infectious sinusitis in chicken and turkey farms particularly in endemic areas (Abd El-Hameed, 2000, Nascimento et al., 2005; Abd El-Hameed, 2006).

ORT, firstly isolated in 1981 from the respiratory tract of turkeys affected with fibrinopurulent airsacculitis while its characterization was done in 1993 (Bordoloi et al., 2020).

Micro-organisms like *Mycoplasma* and *Ornithobacterium* are delicate fastidious ones and difficult to be cultivated with the ordinary methods so in this study, multiplex PCR was used as rapid diagnostic approach help in pathogens detection and control of turkey sinusitis.

MATERIALS AND METHODS

SAMPLES COLLECTION AND PROCESSING

Seventeen turkey farms (2 White Nicholas, 6 Balady and 9 Bronze) at Alexandria governorate were manifested by runny eyes, swelling of infra-orbital sinus either unilateral or bilateral with viscous, slimy or caseated exudates, loss of body weight, high morbidity, and low mortality (Table 1). Exudates of swollen infra-orbital sinus were aspirated and aseptically collected by syringes then transmitted into laboratory for application of multiplex PCR against *E. coli*, *M. gallisepticum*, *M. meleagridis*, ORT, *P. multocida*, and *P. aeruginosa*.

DNA EXTRACTION

QIAamp DNA Mini kit (Qiagen, Germany, GmbH) was used. 200 μ l of the sample suspension was incubated with 10 μ l of proteinase K and 200 μ l of lysis buffer at 56°C for 10 min. and mixed with 200 μ l of 100% ethanol then washed and centrifuged following the manufacturer's recommendations. Nucleic acid was eluted with 100 μ l of elution buffer.

OLIGONUCLEOTIDE PRIMERS

Used Primers were supplied from Metabion (Germany) and listed in Table (2).

PCR AMPLIFICATION

Primers were utilized in a 25 μ l reaction containing 12.5 μ l of EmeraldAmp Max PCR Master Mix (Takara, Japan), 1 μ l of each primer of 20 pmol concentration, 4.5 μ l of water, and 6 μ l of DNA template. The reaction was performed in an Applied biosystem 2720 thermal cycler.

ANALYSIS OF THE PCR PRODUCTS.

 $20 \ \mu$ l of the PCR products was loaded in each gel slot of 1.5% agarose gel (Applichem, Germany, GmbH). Generuler 100 bp ladder (Fermentas, Germany) was used to determine the fragment sizes. The gel was photographed by a gel documentation system (Alpha Innotech, Biometra) and the data was analyzed through computer software.

ETHICAL APPROVAL

The present study was affirmed by the Ethics of Animal Health Committee, Desert Research Center, Egypt.

Table 2: Pr	imers sequences, target g	enes, amplicon sizes and cycling conditions.		J
Multiplex	Target gene	Primers sequences	Amplified segment (bp)	References
	E. coli phoA	CGATTCTGGAAATGGCAAAAG	720	Hu et al.,2011
		CGTGATCAGCGGTGACTATGAC		
А	<i>M. gallisepticum</i> mgc2	CGCAATTTGGTCCTAATCCCCAACA	300	Lysnyansky et al., 2005
11		TAAACCCACCTCCAGCTTTATTTCC		
	P. multocida	P. multocida ATCCGCTATTTACCCAGTGG		Oie, 2012
	Kmt1	GCTGTAAACGAACTCGCCAC		
	P. aeruginosa 16S rDNA	GGGGGATCTTCGGACCTCA	956	Spilker et al., 2004
D		TCCTTAGAGTGCCCACCCG		
D	ORT	TGGCATCGATTAAAATTGAAAG	625	Doosti et al., 2011
	16S rRNA	CATCGTTTACTGCGTGGACTAC		
	M. meleagridis	CGA GCG AAG TTT TTC GGA AC	422	Lierz et al., 2008
1	16S rRNA			

GGTACC GTC AGG ATA AAT GC

RESULTS

RESULTS OF MULTIPLEX PCR

Regarding the results of multiplex PCR used for detection of causative agents of turkeys' sinusitis, Figure 1 exhibited all tested samples were negative for P. aeruginosa and P. multocida at 956, and 460 bp fragment respectively and positive for E. coli, ORT, M. meleagridis, and M. gallisepticum at 720, 625, 422, and 300 bp fragment respectively. Table 3 & 4 revealed out of 17 investigated turkey flocks, one Balady breed flock recorded no infection, two flocks of Balady breed had single infection (one affected with E. coli and the other affected with M. gallisepticum) while the remaining 14 flocks were affected with mixed infection as follow, Three flocks of Bronze breed were affected with the four investigated M.Os, E. coli, M. gallisepticum, M. meleagridis, and ORT, five flocks of 3 Bronze and 2 White Nicholas breed were affected with three M.Os, E. coli, M. gallisepticum, and ORT, two flocks of 1 Bronze and 1 Balady breed were affected with three M.Os, E. coli, M. meleagridis, and ORT, one flock of Balady breed was affected with three M.Os, E. coli, M. gallisepticum, and M. meleagridis, another one Balady breed flock was affected with M. gallisepticum, and ORT. Finally, two flocks of Bronze breed, one of them was affected with E. coli, and M. gallisepticum while the other was affected with M. meleagridis, and ORT.



Figure 1: Multiplex PCR showed samples were positive for *E. coli*, ORT, *M. meleagradis*, and *M. gallisepticum* at the specific amplification of 720, 625, 422 and 300 bp fragment, respectively. All samples were negative for *P. aeruginosa* and *P. multocida* at the specific amplification of 956 and 460 bp fragment, respectively.

INCIDENCE OF DIFFERENT BACTERIA IN TURKEY SINUSITIS

Table 5 showed each of *E. coli*, *M. gallisepticum*, and ORT affected White Nicholas turkey flocks with the same percent 100%. Simultaneously, each of *E. coli*, and *M. gallisepticum* affected Balady turkey flocks with the same percent 50% while each of *M. meleagridis*, and ORT simultaneously found in 33.3% of the same breed. Also with the equal percent, 88.9%, each of *E. coli*, and ORT affected Bronze breed, while 77.8%, and 55.6% of the same breed were affected with *M. gallisepticum*, and *M. meleagridis* respec

OPEN BACCESS

Table 3: PCR results of investigated turkey farms

Farm No.	Breed	E. coli	M. gallisepticum	P. multocida	P. aeruginosa	ORT	M. meleagridis
1	White Nicholas	+	+	-	-	+	-
2	Balady	-	+	-	-	+	-
3	Bronze	+	+	-	-	+	-
4	Balady	+	-	-	-	+	+
5	Balady	-	+	-	-	-	-
6	Balady	-	-	-	-	-	-
7	Bronze	+	+	-	-	-	-
8	Balady	+	-	-	-	-	-
9	Bronze	+	+	-	-	+	-
10	Bronze	+	+	-	-	+	-
11	Bronze	+	+	-	-	+	+
12	Bronze	+	-	-	-	+	+
13	Balady	+	+	-	-	-	+
14	White Nicholas	+	+	-	-	+	-
15	Bronze	+	+	-	-	+	+
16	Bronze	-	-	-	-	+	+
17	Bronze	+	+	-	-	+	+

Table 4: Different bacteria isolated from different turkey breeds affected with sinusitis

Number of affected flocks	Breed	No infection	Single infection	Mixed infection
1	Balady	A		
1	Balady		▲ E. coli	
1	Balady		▲ M. gallisepticum	
3	Bronze			▲ E. coli, M. gallisepticum, M. meleagridis, ORT
5	3 Bronze & 2 White Nicholas			▲ E. coli, M. gallisepticum, ORT
2	1 Balady & 1 Bronze			▲ E. coli, M. meleagridis, ORT
1	Balady			▲ E. coli, M. gallisepticum, M. meleagridis
1	Balady			▲ M. gallisepticum, ORT
1	Bronze			▲ E. coli, M. gallisepticum
1	Bronze			▲ M. meleagridis, ORT

Table 5: Incidence of different bacteria in turkey sinusitis of variable turkey breeds

	· · · · · ·								
Turkey flocks	Total number	E. coli		M. gallisepticum		M. meleagridis		ORT	
		NO.	%	NO.	%	NO.	%	NO.	%
White Nicholas	2	2	100	2	100	0	0	2	100
balady	6	3	50	3	50	2	33.3	2	33.3
Bronze	9	8	88.9	7	77.8	5	55.6	8	88.9
Total number	17	13	76.5	12	70.6	7	41.2	12	70.6

Table 6: Incidence of single and mixed infection

	Micro-organism	E. coli	M. gallisepticum	M. meleagridis	ORT	Total number
Single	-	1 (Balady)	1 (Balady)	-	-	2

September 2022 | Volume 10 | Issue 9 | Page 1965

OPEN OACCESS Advances in Animal and Veterinary Scien								
	E. coli -		10 (1 Balady, 2 White Nicholas, 7 Bronze)	6 (2 Balady, 4 Bronze)	10 (1 Balady, 2 White Nicholas, 7 Bronze)	26		
Mixed	M. gallisepticum	10	-	4 (1 Balady, 3 Bronze)	9 (1 Balady, 2 White Nicholas, 6 Bronze)	23		
	M. meleagridis	6	4	-	6 (1 Balady, 5 Bronze)	16		
	ORT	10	9	6	-	25		

tively. In general *E. coli* isolates were represented 76.5%, each of *M. gallisepticum*, and ORT represented the same percent, 70.6%, while *M. meleagridis* represented 41.2% of turkey flocks.

INCIDENCE OF SINGLE AND MIXED INFECTION

There was one out of 17 turkey flocks negative for multiplex PCR. Furthermore, two flocks were recorded with a single bacterial infection, one flock with *E. coli*, and the other flock with *M. gallisepticum*. Mixed infection with *E. coli* repeated 26 times (10 times mixed with *M. gallisepticum*, 10 with ORT, and 6 with *M. meleagridis*), *M. gallisepticum* repeated 23 times (10 times mixed with *E. coli*, 9 with ORT, and 4 with *M. meleagridis*), *M. meleagridis* repeated 16 times (6 times mixed with *E. coli*, 6 with ORT, and 4 with *M. gallisepticum*), and ORT repeated 25 times (10 times mixed with *E. coli*, 9 with *M. gallisepticum*, and 6 with *M. meleagridis*) (Table 6).

DISCUSSION

Turkeys were considered as competitive protein source in developing countries, so it was necessary to recognize their major pathogens and problems to control and never ignore them long-term.

Swollen infra-orbital sinus is a common respiratory disease which rapidly spread in turkey breeds. So that, in this study multiplex PCR were used to rapidly identify the causative bacteria particularly more delicate and fastidious ones like *Mycoplasma* and ORT.

There were no results appeared with *P. aeruginosa* and *P. multocida* primers, while on investigation of other bacteria, the most prevalent detected one were *E. coli* in 76.5% of affected turkeys followed by *M. gallisepticum* and ORT in 70.6% of cases simultaneously and *M. meleagridis* in 41.2% of cases.

One Balady flock had no infection (5.88%), two Balady

flocks reported single infection (11.76%), one with M. gallisepticum and one with E. coli while mixed infection recorded in 14 farms, 82.35% (100% of each White Nicholas and Bronze and 50% of Balady flocks). from the field surveyed results, E. coli and M. gallisepticum were identified either alone or together with other bacteria, so they might be responsible for turkeys' sinusitis while M. meleagridis and ORT were not identified alone, also E. coli, M. gallisepticum, and ORT were more frequently mixed with each other than M. meleagridis, these results matched with those of Abd El-Hameed et al. (2009) who detected M. gallisepticum, either alone or coupled with other pathogens (P. multocida and E. coli) was the predominant etiologic agent responsible for the respiratory problems including infra-orbital sinusitis of the infected turkey flocks, Sokkar et al. (1986) identified M. gallisepticum and M. meleagridis either alone or mixed with E. coli, P. aeruginosa, S. aureus and S. gallinarun pullorum from turkeys located in different areas of Upper Egypt. Abd El-Rahman (1995) determined serologically M. gallisepticum, M. meleagridis, M. synoviae, and M. Iowa and concluded that M. gallisepticum was the predominant detected mycoplasmas, 90 % of the examined turkeys' flocks referring to the spread of M. gallisepticum rather than M. meleagridis among turkey's farms. Sokker et al. (1986); KeBin (2003) and Moustafa (2005) were naturally and experimentally reported synergistic action between M. gallisepticum and E. coli either alone or associated with other pathogens as a major etiologic agents responsible for swollen head syndrome in turkeys, chickens and ostriches. On the contrary, Eissa et al. (2000) detected M. gallisepticum and M. synoviae in 6.66 % and 13.33 % of examined turkey flocks, respectively indicating that M. synoviae was a prominent cause responsible for turkey's paranasal sinusitis rather than M. gallisepticum. Pan et al.. (2012) and Bordoloi et al. (2020) were reported that ORT had an important role in respiratory affections of broiler chicken and turkey flocks in association with E. coli, M. gallisepticum, M. synoviae, Chlamydophila psittaci, Bordetella avium, and Streptococcus zooepidemicus, also Welchman et al. (2013) and Kursa et al. (2021) mentioned that ORT was a part of the complex with other pathogens that syn-

OPEN OACCESS

ergize to induce the respiratory infection.

CONCLUSION

Turkey Balady breed had a higher resistance to infection than other breeds where it was recorded no or single infection. *E. coli* was the most predominant agents isolated from turkey sinusitis followed by *M. gallisepticum* and ORT with equal percent then *M. meleagridis* had the lowest incidence and frequency. *E. coli* or *M. gallisepticum* was the primary cause responsible for turkey sinusitis as each of them was present in single infection. In this study, a complex of bacterial respiratory pathogens including *E. coli*, *M. gallisepticum*, ORT, and *M. meleagridis* were identified in swollen infra-orbital sinus of turkeys, therefore, Further continuous molecular studies on these delicate pathogens were needed to help in their control.

ACKNOWLEDGMENTS

The authors thank the members of Maryout Research Station and Department of Animal and Poultry Health, Desert Research Center, Ministry of Agriculture, Egypt.

CONFLICT OF INTEREST

The authors declare that they do not have any Competing interests.

AUTHORS' CONTRIBUTION

Hanan El-Samahy designed, and revised the article; Amani Hafez helped laboratory analyses, and tabulation of experimental data; Mohamed Talaat helped in manuscript writing and Disouky Mourad helped field study, collected data, and conducted statistical analysis. All authors have read and approved the final manuscript.

ETHICAL CONSIDERATIONS

Ethical issues including plagiarism, consent to publish, misconduct, data fabrication and/or falsification, double publication and/or submission, and redundancy have been checked by the authors.

REFERENCES

- Abd El-Hameed F (2000). Control of *Mycoplasma gallisepticum* in commercial chicken flocks via vaccination. MVSci Thesis (Poultry Diseases). Faculty of Vet. Med. Assiut University, Egypt.
- Abd El-Hameed F (2006). Control of *Mycoplasma gallisepticum* in broiler breeder flocks using different methods. Ph D Thesis (Poultry Diseases) Faculty of Vet. Med. Beni-Suef

University, Egypt.

- Abd El-Hameed F, El-Shafey DYH, Abd El-Dayem NS, Tantawi LA (2009). Role of various species of mycoplasma and bacteria in turkey's sinusitis with description of pathological picture. Assiut Vet. Med. J. 55: 121.
- Abd El-Rahman FI (1995). Incidence of mycoplasmal infections in turkeys. Veterinary Med. J. Giza. 43 (2): 201-206.
- Alessandri E, Massi P, Paganelli F, Prandini F, Saita M (2005). Field trials with the use of a live attenuated temperaturesensitive vaccine for the control of *Mycoplasma gallisepticum* infection in meat type turkeys. Italian J. Anim. Sci. 4 (3): 282-286. https://doi.org/10.4081/ijas.2005.282
- Ali AR, Youssef AE (2003). Bacteriological studies and biochemical parameters of respiratory infection in ostriches. Vet. Med. J. Giza. 51 (2): 189-203. https://www.cabi.org/isc/ abstract/20033123604
- Bordoloi S, Nayak A, Jogi J, Shakya P, Rai A, Sharma S (2020) Ornithobacterium rhinotrachalae : An emerging poultry pathogen. J. Entomol. Zool. Stud. 8(2): 92-97.
- Bradbury JM (2006) Mycoplasmas ever present pathogens? Int. Hatchery Pract. 20 (7): 21-23.
- Büyüktanir Ö, Yıldırim T, Yakicier C, Genc O, Yurdusev N (2008). A recombinant PvpA protein-based diagnostic prototype for rapid screening of chicken Mycoplasma gallisepticum infections. Vet. Microbiol. 129: 139-149. https://doi.org/10.1016/j.vetmic.2007.11.028
- Doosti A, Sharifzadeh A, Ghasemi H, Vaez J (2011). Molecular identification of Ornithobacterium rhinotracheale in turkeys in Isfahan province of Iran. African J. Biotechnol.10 (40): 7911-7914. https://doi.org/10.5897/AJB11.1077
- Eissa SI, Dardeer MA, Abo-Norag MA (2000). Application of sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) for identification of mycoplasma infection in turkeys with special reference to treatment. Vet. Med. J. Giza. 48 (2): 197-206.
- Hafez HM (2002). Diagnosis of Ornithobacterium rhinotracheale. Int. J. Poult. Sci. 1 (5): 114-118. https://doi. org/10.3923/ijps.2002.114.118
- Hu Q, Tu J, Han X, Zhu Y, Ding C, Yu S (2011). Development of multiplex PCR assay for rapid detection of *Riemerella anatipestifer*, *Escherichia coli*, and *Salmonella enterica* simultaneously from ducks. J. Microbiolog. Methods. 87 (1): 64-69. https://doi.org/10.1016/j.mimet.2011.07.007
- KeBin L (2003). Diagnosis and control of the mixed infection of *Mycoplasma gallisepticum* with *Escherichia coli*. Chinese J. Vet. Sci. Technol. 33 (9): 67-68.
- Kursa O, Tomczyk G, Sawicka-Durkalec A, Giza A, Słomiany-Szwarc M (2021). Bacterial communities of the upper respiratory tract of turkeys. Sci. Rep. 11, 2544.
- Lierz M, Hagen N, Lueschow D, Hafez HM (2008). Use of polymerase chain reactions to detect Mycoplasma gallisepticum, Mycoplasma imitans, Mycoplasma iowae, Mycoplasma meleagridis and Mycoplasma synoviae in birds of prey. Avian Pathol. 37(5): 471-476. https://doi.org/ 10.1080/03079450802272952
- Lynsyansky I, Garcia M, Levisohn S (2005) Use of mgc2-Polymerase chain Reaction-Restriction fragment length polymorphism for rapid differentiation between field isolates and vaccine strains of *Mycoplasma gallisepticum* in Israel. Avian dis. 49 (2):238-245. https://doi.org/10.1637/7285-10020R
- Moustafa FA (2005). Some studies on bacterial causes associated with cases of swollen head syndrome in chickens. Assiut

OPEN OACCESS

Advances in Animal and Veterinary Sciences

Vet. Med. J. 51 (104): 1-16. https://doi.org/10.21608/ avmj.2005.177782

- Nascimento ER, Pereira VLA, Nascimento MGF, Barreto ML (2005) Avian mycoplasmosis update. Revista Brasileira de Ciencia Avicola. 7 (1): 1-9. https://doi.org/10.1590/S1516-635X2005000100001
- Nicholas RAJ, Ayling RD, McAuliffe L (2009). Vaccines for Mycoplasma diseases in animals and man. J. Comp. Pathol. 140: 85-96. https://doi.org/10.1016/j.jcpa.2008.08.004
- OIE (2008). Manual of Diagnostic Tests and Vaccine for Terrestrial Animals (Mammals, Birds and Bees), Avian Mycoplasmosis (*Mycoplasma gallisepticum*, *M. synoviae*). 6th Edition, OIE. Pages 482-496. http://www.oie.int/eng
- OIE (2012). OIE Terrestrial Manual 2012. Chapter 2 .4 .1 2. haemorrhagic septicaemia.
- OIE (2018). Terrestrial Manual, Chapter 3.3.5. Avian Mycoplasmosis (*Mycoplasma gallisepticum*, *M. synoviae*). 844-859. http://www.oie.int/eng

Pan Q, Liu A, He C (2012). Co-infection of Ornithobacterium

rhinotracheale with Streptococcus zooepidemicus in chickens. Avian Dis. 56(4):680-684. https://doi. org/10.1637/10109-030112-Reg.1.

- Sokkar IM, Soliman AM, Mousa S, El-Demerdash MZ (1986). In-vitro sensitivity of mycoplasmas and associated bacteria isolated from chickens and turkeys and ducks at the area of Upper Egypt. Assiut. Vet. Med. J. 15 (30): 243-250.
- Spilker T, Coenye T, Vandamme P, LiPuma JJ (2004). PCR-Based Assay for Differentiation of Pseudomonas aeruginosa from Other Pseudomonas Species Recovered from Cystic Fibrosis Patients. J. Clin. Microbiol. 2074-2079. https:// doi.org/ 10.1128/JCM.42.5.2074-2079.2004
- Welchman DB, Ainsworth HL, Jensen TK, Boye M, King SA, Koylass MS, Whatmore AM, Manvell RJ, Ayling RD, Dalton JR (2013). Demonstration of Ornithobacterium rhinotracheale in pheasants (Phasianus colchicus) with pneumonia and airsacculitis. Avian Pathol. 42(2): 171-178. https://doi.org/ 10.1080/03079457.2013.778387