Taxometrical and numerical characterization of an isolate of *Steinernema abbasi* (Elawad *et al.*, 1997) with larger infective juveniles comprehensive from ITS1-5.8S-ITS2 region of rRNA

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Abstract

In present investigation *Steinernema* sp. isolate CS_1 was identified with the aid of numerical taxonomy as an additional tool. Presence of two horn-like structures on head region of 3rd stage infective juveniles (IJs) placed the nematode in bicornutum group and differentiated from other compared *Steinernema* spp. The morphological characters of 3rd stage infective juveniles and 1st generation males showed close resemblance with *S. abbasi, S. thermophilum* and *S. pakistanense* however, the body length was varied (583mm vs 541mm, 555 mm and 683 mm, respectively). The morphometrical characters and life history events placed the present specimen near to the *S. abbasi.* Results obtained through the phylogenetic utility of maximum parsimony, maximum likelihood and neighbour joining methods and comparative account on nucleotides analyses based on ITSs study confirmed the present specimes as isolates of *S. abbasi.* The nucleotide composition resemblance within 5.8S gene with other *Steinernema* species evident the less polymorphism and highly conserved nature of this region as compared to ITS1 and ITS2 and delimit the relationship of steinernematid nematodes.

Keyword: *Steinernema* spp., molecular tools, polythetic divisive classificatory system, ITS region, nucleotide composition.

Entomopathogenic nematodes (EPN) belonging families Steinernematidae to the and Heterorhabditidae are the lethal parasite of insects and are in the keen interest of scientific community as biological control agent from past decade (Kaya & Gagular, 1993). This can be traced by the increment in number of research publications on this group of nematodes in recent years and a large number of laboratories have engaged world-wide in EPN research. Taxonomy of these nematodes is an indispensable as the product they involved in mass production, formulation and application in insect pest management. The application of EPN has enthused frequent inspections in an effort to find new indigenous isolates and possibly also new Heterorhabditis and Steinernema species

(Hominick et al., 1996). Members of both the families showed close intimacy with each other in terms of life cycle morphology and pathogenicity but convergent evolution are not indicative towards the close phylogenetic relationship between these two (Sudhaus, 1993; Blaxter et al., 1998). The present scenario shows the richness of diversity of steinernematids throughout the world. Presently the family Steinernematidae includes two genera i.e., Steinernema with more than 100 species globally and Neosteinernema Nguyen & Smart, 1994 with only one species N. longicurvicauda Nguyen & Smart, 1994 (Adams & Nguyen, 2002). Diversity of EPN in India is not done exhaustively so far as only 16 species

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isolated from different parts of the country. Although much of the work is concentrated towards the extensive surveys and pathogenicity trials, the lack of taxonomic expertize globally and close intimacy within the genus *Steinernema* are some critical factors behind this tricky. Traditional classificatory procedure looks for the most common character for differentiation within the specimen under consideration.

Traditional methods such as morphology and morphometric were always the baseline of the taxonomy of nematodes. But now days these become supplemented with the molecular approaches to concern the identification of putative new species (Hominick et al., 1997). The aim of present study was to ensure more precise identification to isolated Steinernema sp. isolate CS_1 to clear the obscure in EPN taxonomy. Here is the polythetic divisive method of classification (Malhotra et al., 1981) successfully used for cestode taxonomy previously and was applied to distinguish the with morphological EPN species along taxometrical and molecular analyses which give a clue to resolve the uncertainities and showed their usefulness up to some extent.

Materials and Methods

Isolation of entomopathogenic nematodes: The nematode was collected from the soil of District Saharanpur from the field of *Sorghum* popularly known as Jowar through soil baiting technique of Bedding & Akhurst (1975) using *Galleria mellonella* larvae as bait. The nematode was tagged as *Steinernema* sp. isolate CS_1 as per the author's laboratory nomenclature.

Nematode culture: Population of isolated EPNs were maintained *in vivo* on the last instar larvae of *G. mellonella* at a temperature of 27 ± 2 °C for further processing. 20 larvae of *G. mellonella* were infected with 2000 infected juveniles (IJs) in a Petri plate lined by double layer of filter paper. The cadavers were transferred on White Trap (White, 1927) and IJs were isolated from white trap, disinfected with 0.1% sodium hypochlorite and stored into tissue culture flask

at $15\pm1^{\circ}C$ temperature in BOD for further processing.

Recovery of adult stages: For recovery of adults the cadaver of *G. mellonella* were dissected 2-3 days and 4-6 days after mortality for 1^{st} and 2^{nd} generations males and females, respectively in Ringer's solution (Woodring & Kaya, 1988). Freshly emerged IJs were used for the identification purpose.

Processing of nematodes for microscopy: 1st and 2nd generation males and females along with IJs of present nematode were killed by gentle heat and fixed in TAF (Courtney *et al.*, 1955) and processed to glycerol by the method adopted from Sienhorst (1959). Slides for microscopic study were prepared on glass slide where specimens were mounted in glycerine covered with cover slip. All the stages were studied under 4x, 10x, 40x and 100x magnifications using light and phase contrast microscope. Measurements were taken using phase contrast microscope inbuilt with measurement software DL1.

Morphology and taxometry: For morphotaxometrical comparison, size of 3rd stage juveniles was taken as base and nine species of Steinernema viz., S. anatoliense (Hazir, Stock & Keskin, 2003), S. carpocapsae (Weiser, 1955), S. kushidai (Mamiya, 1988), S. scapterisci (Nguyen & Smart, 1992), S. tami (Luc, Nguyen, Reid & Spiridonov, 2000), S. websteri (Cutler & Stock, 2003), S. abbasi (Elawad, Ahmad & Reid, 1997), S. pakistanense (Shahina, Anis, Reid, Rowe & Maqbool, 2001) and S. thermophilum (Ganguly & Singh, 2000) were chosen for comparison. The data of 3rd stage juveniles and 1st generation males was extracted from the literature and used for the comparison.

Genomic DNA isolation: The genomic DNA was isolated from fresh IJs culture using the DNeasy Blood and Tissue Kit (Qiagen) as per the instruction given by the manufacturer. The isolated DNA was electrophoresed with

0.7% agarose gel in TAE buffer containing EtBr. The gel was visualized under UV light for the presence of DNA.

PCR Analysis: Internal Transcribed Spacer (ITS) regions of the genomic DNA were used as marker to distinguish the present specimen form compared species of Steinernema. The amplification of ITS regions was done using the primers as suggested by Joyce et al., (1994) with modifications. The composition of 25 µL of PCR reaction volume were; Dream Taq Green master mix 2x (Thermo Scientific) 12.5 μ L, forward primer 0.7 μ L, reverse primer μ L, template DNA 2 μ L, mili Q water 9.1 μ L. the PCR running protocol was set for 35 cycles and it was as initial temperature 94 °C for 5 minutes, denaturation at 94 °C for 30 seconds, annealing temperature 64 °C for 30 seconds, primer extension 72 °C for 30 seconds, final extension 72 °C for 8 minutes, storage temperature 4 °C for infinity. 3µL of amplified DNA along with 100 bp DNA ladder was loaded in the well of 1% agarose gel in TAE buffer containing EtBr as visualization dye.

Molecular characterization: The obtained sequence was submitted to GenBank (accession no. KP0369180).10 sequences of ITS regions of already described Steinernema species showing close similarity with the sequence of present specimen based on BLAST analysis included some of those were compared morphologically downloaded from NCBI. ITS region of Caenorhabditis elegans was used as an outgroup. The sequences were aligned using the default parameters (gap opening penalty 15, gap extension penalty 6.66) in Clustal W (Tamura et al., 2004) programme in MEGA software version 6 (Tamura et al., 2013). Phylogenetic trees were constructed by neighbour joining (Saitou & Nei, 1987), maximum likelihood (Tamura & Nei, 1993) and maximum parsimony (Nei & Kumar, 2000) methods. The pair wise distance between all the sequences was also calculated using the same software. Nucleotide compositions of the sequences were calculated through Bio Edit software.

Polythetic divisive classificatory system: Total body length (L), Greatest body width (GBW), Position of excretory pore from anterior end (EP), Nerve ring (NR), Oesophagus (ES), Tail length (Tail) Ratio a (L/GBW), b (L/ES), c (L/T), D% (EP/ES×100) and E% (EP/Tail×100) were statistically compared where the method of polythetic Divisive classificatory system (Malhotra *et al.*, 1981) was applied.

Results

Morphological characters of *Steinernema* sp. isolate CS₁ (Fig. 1-2; Table 1-3.)

First generation males: Body ventrally curved when hot killed. Cuticle smooth, lateral fields and phasmid not observed under light and phase contrast microscope. Stoma reduced, cheilorhabdions prominent, posterior part of the stoma funnel shaped, moderately cuticularized or not cuticularized. Pharynx muscular, divided into cylindrical procorpus with slightly swollen metacorpus and distinct isthmus surrounded by nerve ring, basal bulb prominent, length of basal bulb is slightly more than width. Excretory pore located on isthmus, anterior to nerve ring above the basal bulb. Distance from the anterior to excretory pore always more than body width at excretory pore but less than greatest body width. Width at excretory pore is about 59% of the oesophagus length. Oesophagus muscular. Cardia present and prominent. Genital system monorchic and clearly visible with reflexed testis makes 15-20% of total body. Spicules golden yellow in view, paired, heavy, curved with internal ribs.

Capitulum rounded. Spicule tip somewhat conoid, manubrium short with $9-15\mu m$ almost similar length and width, velum prominent extending for more than 2-3rd of the blade. The spicule at its posterior 3^{rd} straight and a depressed on the ventral side before the spicule tip. Calomus (shaft) present and the blade or lamina moderately curved; rostrum usually present. Gubernaculum 70% of the spicule length, boat-shaped, tapering anteriorly, slightly swollen in the middle to a somewhat ventrally curved end. Corpus separated posteriorly while cuneus rod-like. Tail is 70% of its width, conoid, without bursa. Mucron not visible.

Second generation males: Morphologically similar to 1st generation but approximately 69% of its size. Size of excretory pore, oesophagus, nerve ring and tail are 82-86% of the 1st generation. Spicules and gubernaculum smaller and thinner in size possessing 86% and 66% size of 1st generation. Shape of the spicule and gubernaculum not different from 1st generation males, but slightly variable between individuals of same generation. Mucron like process observed rarely on tail.

First generation females: Similar to other species of Steinernema. Body spiral when heat killed. Giant females 40-50% larger to normal 1st generation females, however, similar in morphology. Cuticle smooth, lateral fields and phasmids inconspicuous. Labial region rounded with six labial palp in light and phase contrast microscopy. Excretory pore anterior to nerve ring, length slightly less than the width and 31% of oesophagus length in average. Pharynx muscular; procorpus, metacorpus and isthmus prominent. Basal bulb enlarged. Nerve ring usually surrounding anterior portion of basal bulb or posterior part of isthmus. Cardia present. Genital system amphidelphic usually reflexed and occupied with eggs, endotokiametricida common. Vulva in the form of transverse slit protruding from the body surface, doubleflapped epiptygmata present. Vagina short and muscular. Vulval aperture located posterior to mid-body. Postanal swelling usually present. Tail conoid to dome shaped, shorter than anal body, mucronate process absent.

Second generation females: Body C shaped when heat killed, similar to 1st generation females in common features but smaller (about 29%) in size and diameter (about 39% at vulva). Tail with prominent swelling, longer than anal body diameter and 1st generation female's tail, tapering or pointed with prominent mucron.

Infective juveniles: Body shape ventrally curved or J shaped or straight when heat killed, slender, elongate, tapering at both ends, labial region continuous with body. Cuticle with fine striae. Labial region smooth, usually continuous. Head bearing two horn like structure, Oral apertures and anus closed. Excretory pore poorly developed located posterior to nerve ring, 8% of the total body and 36% longer to its width. Nerve ring distinct located above the middle of excretory pore and basal bulb. Pharynx long and narrow, distinctly narrower at the level of nerve ring, terminating in a dorsally displaced, valvate bulb. Nerve ring distinct, situated anterior to basal bulb. Cardia present. Tail 26% greater of its anal body width, conical in shape and terminally pointed.

Molecular characterization and phylogenetic relationship: The amplified region of the studied specimen (KP036918) yielded a 800 nucleotide long sequence containing 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, complete sequence and internal transcribed spacer 2 partial sequence. For ITS regions, maximum parsimony analysis showed that the alignment resulted in 1017 characters of which 185 are constant, 449 are parsimony uninformative and 383 are parsimonyinformative.



Fig. 1 (A-L). Microphotographs of *Steinernema* sp. isolate CS₁. A-C: First generation female, D-E: First generation male, F-H: Second generation female, I-J: Second generation male, K-L: Third stage juvenile. A, D, F, I, K: Anterior region, B, G: Middle region, C, E, H, J, L: Posterior region.



Fig. 2. Camera Lucida sketch of *Steinernema* sp. isolate CS₁. A-F: First generation female, G-H: First generation male, I-K: Second generation female, L-M: Second generation male, N-O: Third stage juvenile, A, G, I, L, N: Anterior region, B, J: Middle region, C-F, H, K, M, O: Posterior region; C-F: Variation in tail region of 1st generation female.

Characters	(Male)			(Female)		
	1 st Generation	2 nd Generation	1 st Generation	Giant Female	2 nd Generation	Juveniles
n	15	15	15	11	15	20
T	1432.1±260.8	839.2±83	4985±710.9	8664.7±1671.1	1432.1±237.4	583.1±39.2
L	(978.4-1964.9)	(653.3-990.2)	(3200-5930.7)	(6884.1-11419.4)	(1021.2-1732.8)	(512.9-675.1)
	14.4±2.2	16.5±1.5	21.5±3.8	33.6±5.4	18.2±1.5	23.8±0.9
а	(10.2-18.6)	(13.2-19.1)	(12.8-26.5)	(24.7-43)	(15.2-20.3)	(22.6-25.6)
	10.1±1.5	7±0.7	29±4.4	46.5±7.3	22.3±3.2	5.9±0.3
b	(7.6-12.1)	(5.7-8.6)	(18.3-33.9)	(39.3-58.7)	(17-28.1)	(5.3-6.60)
	57.4±8.9	41±3.9	150.2±31.9	184.2±58.7	26.4±3.2	10.4±0.7
с	(40.8-74)	(36.4-49.1)	(105.2-242)	(113-282.2)	(21.9-33.2)	(9.1-11.8)
	0.3	0.4	0.1	0.2±0.1	0.7±0.1	2.3±0.2
C'	(0.1-0.3)	(0.3-0.5)	(0.1-0.2)	(0.1-0.3)	(0.6-0.8)	(2-2.6)
	· · · ·	· · · ·	53.5±1.9	51.8±1.6	54.6±2	
V			(49.8-56.1)	(49.5-54.4)	(52.9-60.7)	
~~~~	99.3±8.9	51.1±4.4	235.2±33.4	264.3±66.6	78.9±11.9	24.5±1.5
GBW	(89.2-121.2)	(45-59.6)	(153.5-294.3)	(185-365.4)	(65.9-102)	(20.2-26.5)
55	81.1±7.1	66.4±4.3	53.7±8.9	88.4±13	64.2±4.8	48.1±3.2
EP	(63.5-89.5)	(54.9-75.1)	(40.7-77.9)	(75.5-121.6)	(57.8-77.6)	(39.8-52)
ND	108.5+10.7	93.4+4.5	127.8+5.5	140.7±9.5	97.4±5.8	77.5±4.7
NR	(94.4-133.4)	(86.7-101.7)	(119.7-138)	(129.3-159.4)	(88.2-107.7)	(64.1-84.8)
50	138.5+10.3	119.3+3.8	172.5+9.7	185.6±13	134.8±4.1	99.3±3.5
ES	(124.3-161.5)	(114-127.7)	(157.5-196.2)	(170.5-215)	(129-142.9)	(92.3-105.1)
тр	255.5±55.1	175.8±31.2	· · · · · ·	. ,	. , ,	
IK	(196.4-383.6)	(118.3-225.3)				
Tail	24.9±1.8	20.5±2	33.8±5.2	50.8±17.2	54.1±4.2	56.3±3.7
Tan	(21.6-28.1)	(17.9-25.1)	(21.8-42.8)	(30.9-86.1)	(46.1-60.6)	(48.3-62.3)
ARW	35.6±3.5	29.3±2.3	47.5±10.2	76.9±27.9	33.8±4.8	14.7±1.3
AD W	(31.8-40.7)	(24.9-33.2)	(31.8-66.7)	(53.8-138.6)	(24.2-41.4)	(11.7-16.6)
SL	65.7±4.5	55±4.6				
51	(58.9-73.4)	(46.6-61.9)				
SW	$11.1\pm1.7$	$6.6\pm0.9$				
	(8.9-13.2)	(4.7-7.9) 20 4+3 7				
GL	(35.4-52.3)	(23.7-37.2)				
~~~	7.2±0.9	6.3±0.8				
GW	(5.8-9.3)	(4.9-7.6)				
DØ	58.6±4.6	55.7±2.9	31.3±5.7	47.5±4.9	47.6±2.8	48.5±2.4
D%	(50.9-69.6)	(47.5-58.8)	(20.8-44.6)	(41.7-56.5)	(42.6-55.4)	(42.1-51.8)
	325.8±24.72	327.8±40.1	161±26.9	193.9±77.2	119.1±9.6	85.7±5.8
E%	(64.6-366.5)	(226.5-383.6)	(114-198.7)	(99.6-338.8)	(102.1-134.4)	(71.1-92)
Ea	399.4±36.8	251.5±31.9	724.1±222.9	576±254.1	145.7±16.1	43.7±3.2
F%	(342.1-500.7)	(190.7-326.9)	(416.5-1348.9)	(325.9-989.8)	(122.9-182.9)	(38.3-50.6)
SWO	186.2±23.1	189±21.5				
S W 70	(151.6-220)	(142.8-221.6)				
GS%	67.1±5.9	53.5±5.1				
5570	(55.9-80.5)	(45.6-62.8)				

Table 1. Morphometrical characters of Steinernema sp.	isolate CS ₁ . All measurements in μ m except n
(number), M±SD followed by range.	

fo	llowed by 1	ange.								
Charactore	s	s	S	S	S	s	s	S	S	Steinernema
	anatoliense	carpocapsae	kushidai	scapterisci	tami	websteri	abbasi	pakistanense	thermophihm	sp. CS
ц	20							20	20	20
L	545 ± 21	558	589 ± 39.8	572 ± 27	530 ± 4	584 ± 13	541±24	683±21	555±34	583.1±39.2
4	(507-580)	(438-650)	(424-662)	(517-609)	(400-600)	(553-631)	(496-579)	(649-716)	(480-620)	(512.9-675.1)
	22 ± 2	21	22.5 ± 1.6	24	23 ± 2	28 ± 2.7	18±0.9	24±1.5	26±0.9	23.8±0.9
ed	(19-27)	(19-24)	(19.3-25.2)	(20-31)	(19-28)	(24-35)	(17-20)	(21-27)	(24-28)	(22.6-25.6)
	5.1 ± 0.3	4.4	5.3 ± 0.3	4.5	5 ± 0.3	5.1 ± 0.2	6±0.3	6±0.3	6.4±0.4	5.9±0.3
5	(4.6-5.7)	(4-4.8)	(4.9-5.9)	(4-4.6)	(3.7-5.1)	(4.8-5.6)	(5.5-6.6)	(2-6)	(5.8-7.1)	(5.3-6.60)
c	10.5 ± 0.5	10	11.7 ± 0.6	10.7	11 ± 0.5	12.6 ± 1.3	9.8±0.8	11±0.5	12.3±0.4	10.4 ± 0.7
o	(9.4-11.8)	(11-6)	(9.9-12.9)	(9.2-11.7)	(11-6)	(11-15.5)	(8.1-10.8)	(10-12)	(11.5-12.8)	(9.11-11.8)
~									3.4±0.3	2.3±0.2
2	•			•	4.2	•	•	•	(3-3.9)	(2-2.6)
1000	24.5 ± 2	25	26 ± 2.5	24 ± 4	23 ± 2	21 ± 17	29±1	27±1.2	21±0.7	24.5±1.5
-	(21-28)	(20-30)	(22-31)	(18-30)	(19-29)	(17-25)	(27-30)	(24-29)	(21-23)	(20.2-26.5)
дд Д	37 ± 1	38	46 ± 2	39 ± 4	36 ± 2	36 ± 3.1	48±1.5	54±2.2	40±2	48.1±3.2
1	(36-39)	(30-56)	(42-50)	(36-48)	(34-41)	(29-40)	(46-51)	(49-58)	(37-46)	(39.8-52)
8	76 ± 3	85	76 ± 3.5	97 ± 1.1		88 ± 3.6	68±2.4	80±2.1	71±4	77.5±4.7
	(71-82)	(16-99)	(70-84)	(83-106)	•	(83-95)	(64-72)	(76-83)	(62-79)	(64.1-84.8
5 H	107 ± 6	120	111 ± 4.1	127 ± 6	117 ± 4	115 ± 4.4	89±1.8	113±4.2	87±6	99.3±3.5
3	(97-124)	(103 - 190)	(106-120)	(113-134)	(110-123)	(107-122)	(85.92)	(108-122)	(80-100)	(92.3-105.1)
Tail	53 ± 3	53	50 ± 3.4	54 ± 3	50 ± 4	47 ± 4.5	56±3.2	58±2.1	45±3	56.3±3.7
TIR T	(46-58)	(46-61)	(44-59)	(48-60)	(42-57)	(37-56)	(52-61)	(53-62)	(40-52)	(48.3-62.3)
Hvalina (H)	22.5 ± 1					11 ± 0.5				
	(19-26)	•	,	,	•	(10-14)	,	•	•	•
A RW	12 ± 0.7			31 ± 3	12 ± 1	12 ± 1.2			13±1	14.7±1.3
	(11-14)	•	,	(27-40)	(11-13)	(10-14)	•	•	(12-15)	(11.7-16.6)
7% 2	35 ± 2.5	26	41 ± 1.4	73 ± 6	31 ± 0	31 ± 2.7	53±2	47±2.7	46±3.5	48.5±2.4
2	(31.5-39)	(23-28)	(38-44)	(60-80)	(28-34)	(24-34)	(51-58)	(42-53)	(42-53)	(42.1-51.8)
70%	72 ± 3.9	60			73 ± 0	77 ± 11	86±5	91±5	90∓6	85.7±5.8
۰. ۲	(64-81.5)	(54-66)	92	•	(67-86)	(62-102)	(79-94	(87-102)	(81-102)	(71.1-92)
‰4										43.7±3.2
	•	•	•	•	•	•	•	•	•	(38.3-50.6)
%H	43 ± 0.8				40 ± 0.1	33 ± 0.6				
	(41-45)				(30-50)	(30-34)				

Table 2. Comparison table of 3rd stage juveniles of *Steinernema* species. All measurements in µm except n (number), M±SD

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Charactare	s	5	5	S	8	8	S	5	5	
	anatoliense	CAIPOCAPSAC	kushidai	scapterisa	to mi	rieb steri	abbasi	pakista ense	thermoghiham	Ś
Ħ	19	25	20	10	20	19		20	20	15
Г	1532±195	1450	1400±16	1728 ± 358	1600±20	1712±92	1252±189	1357±89	1197±236	1432±261
	(1236-1841)	(01/1-0601)	(1200-1900)	(1319)	(1200-1900)	((1223-1865)	(999-1334) ((1163-1505)	(08/1-066)	(378-1965)
đ		•		•			•	011-16)	(12.8-17.8)	(10.2-18.6)
۹.	,	,		,	,	,	,	10.4±0.6	9.7±1.4	10.1±1.5
1								(8.7-11)	(8-12.5)	(7.6-12.1)
0	•	•	•	•	•		•	55±3.3	53±4	57.±8.9
								(52-62)	(48-63)	(40.8-74)
°,	•	•	•	•	•		•	•	0.6±0.1	0.3
mat	10015	100	07410	156440	110117	LITLVI	LYTLO	014001	(//n-c/n)	(5.U-1.U)
-	(87-150)	(77-131)	(75-156)	(97-231)	(89-161)	(119-175)	(82-98)	(80-128)	(00-100)	(89.2-121.2)
EP	67±0.5	, el	84±7.9	11±11	68±12	62±6	80±7.8	81±4.8	75±8.5	81±7.1
!	(61-88)	(47-74)	(71-105)	(63-98)	(43-92)	(54-73)	(68-89)	(72-92)	(64-92)	(64-90)
NR	107±15	110	129±5.3	136±11	•	01#611	103±6.5	99±6.3	93±8.2	108.5±10.7
ŝ	(8/-131)	(93-124)	(120-137)	(241-021)		(651-001)	(521-66)	(/.01-88)	(011-08)	(94-133)
S	144±8 170 1601	CCI (TAL APL)	16/±/.9	18/±21	113±80	163±12 /135 180/	133±6 /121_144)	132±5.8	125±20	139±10.3
TR	-	(101-001)	-	374+52	-	(001-CCT)	274+33		-	256455
4		(400-808)		(306-447)			(234-319)		-	196.4-383.6)
Tail	30±3	30	33±3	25±3	23±5	29±2	26±3	25±0.8	22.4±4.7	25±1.8
	(26-64)	(23-39)	(30-40)	(21-30)	(10-32)	(25-33)	(20-31)	(24-27)	(19-34)	(22-28)
ABW	52.5±3.5	43	42±5	33±5	37±6	37.5±2	43±5	36±2.3	37±7	36±3.5
SDI	(47-60)	(cc-55)	(36-54) 63±5 5	(C4-15) 83±5	(07-00)	(34-41) 68±0	(cc-15)	(32-40) 68±3 6	(28-49) 61±7	(32-41) 664-45
110	(68-84)	(59-72)	(48-72)	(12-92)		(64-72)	(57-74)	(62-73)	(44-72)	(59-73)
SPW				13±4	77±4		12+1.3			11+1.7
		(9-13)		(13-14)	(71-84)		(10-14)			(9-15)
ថ	47±3.5	47	44±4.9	65±5	48±5	49±3.2	45±4.3	41±3.2	36±4	44±5
	(42-59)	(39-56)	(39-60)	(59-75)	(38-55)	(42-56)	(35-50)	(36-45)	(30-42)	(35-52)
GW		5 (1-1)		8±0.5	•		7±0.1		•	7.2±0.9
D%	48.5±3.5	39	51±4.8	36±2	4410	40±10	60±5	60±3	63±10	59±4.6
	(46.5-55)		(42-59)	(32-39)	(30-60)	(30-50)	(51-58)	(20-60)	(50-87)	(51-70)
Е%		•		•		210 ± 20	•	310±40	343±43	326±25
i						(180-250)		(210 - 370)	(269-400)	(65-367)
r%				•			•		•	342-501)
SW	175±10	151	150	252	20040	180 ± 10	156±22	189	1.7 ± 0.4	1.9±0.2
	(160-190)			(204-280)	(140-300)	(160-210)	(107-187)		(1.2-2.8)	(1.5-2.2)
8	55 <u>4</u>	72	70	78	62	70±10	70±7	60.3	0.6-0.1	0.7±0.1
	(c9-04)			(69-84)		(00-80)	(c8-8c)		(1.0-0.0)	(0.6 - 0.8)
MUC	+	+	•	+	+	+	•	+	•	•

The maximum parsimony generated 3 parsimoneous tree with a length of 927 positions. The consistency index, retention index and the composite index were 0.652, 0.654 and 0.426, respectively for all sites and parsimony-informative sites. Phylogenetic analysis through neighbour joining (Fig. 4), maximum likelihood

(Fig. 5) and maximum parsimony (Fig. 6) yielded a total of 575 positions in the final dataset. Pair wise distance analysis (Fig. 3) showed no difference between *Steinernema* sp. isolate CS_1 , *S. abbasi* and *S. thermophilum*. Analysis of nucleotide composition were also indicative for the same results (Table 4).

	CS ₁	Sab	Sth	Spa	Sbi	Sce	Syi	Sca	Sku	Ssc	Sta	Cle
Steinernema sp. CS ₁												
S. abbasi	0.000											
S. thermophilum	0.000	0.000										
S. pakistanense	0.348	0.348	0.348									
S. bicornutum	0.281	0.281	0.281	0.345								
S. ceratophorum	0.277	0.277	0.277	0.350	0.111							
S. yirgalemense	0.239	0.239	0.239	0.346	0.324	0.321						
S. carpocapsae	0.392	0.392	0.392	0.530	0.380	0.399	0.443					
S. kushidai	0.453	0.453	0.453	0.550	0.431	0.422	0.516	0.336				
S. scapterisci	0.395	0.395	0.395	0.532	0.374	0.391	0.452	0.108	0.343			
S. tami	0.396	0.396	0.396	0.523	0.396	0.422	0.443	0.083	0.388	0.137		
C. elegans	0.978	0.978	0.978	1.084	0.955	0.936	1.040	1.015	1.132	1.063	1.019	

Fig. 3. Distance matrix of compared Steinernema species.



0.1

Fig. 4. Phylogenetic analysis of internal transcribed spacer region (ITS) of ribosomal DNA based on neighbour joining analysis.



0.1

Fig. 5. Phylogenetic analysis of internal transcribed spacer region (ITS) of ribosomal DNA. based on maximum likelyhood analysis.



Fig. 6. Phylogenetic analysis of internal transcribed spacer region (ITS) of ribosomal DNA.based on maximum parsimony analysis.

S. No	Species	Accession number	Length (bp)	ITS1	5.8 S	ITS2	G+C%	A+T%	A	С	G	Т
1	Isolate CS ₁	KP036918	739	268	157	314	36.94	63.06	175	111	162	291
2	S. tami	AY171280	728	269	155	304	38.87	61.13	163	120	163	282
3	S. carpocapsae	AY171282	731	279	157	295	37.89	62.11	170	117	160	284
4	S. scapterisci	AF122020	710	247	157	306	36.76	63.24	169	108	153	280
5	S. pakistanense	AY748449	748	291	157	300	36.9	63.1	216	118	158	256
6	S. bicornutum	AY171279	768	281	157	330	37.5	62.5	201	123	165	279
7	S. ceratophorum	AY230165	741	243	157	341	36.17	63.83	192	117	151	281
8	S. yirgalemense	AY748450	711	270	157	284	35.72	64.28	191	93	161	266
9	S. abbasi	AY248749	739	268	157	314	36.99	62.74	175	111	162	291
10	S. thermophilum	EF431958	739	268	157	314	36.94	63.06	175	111	162	291
11	S. kushidai	AB243440	741	279	157	305	42.24	57.76	172	135	178	256
12	C. elegans	X03680	1001	464	153	384	47.55	52.45	229	220	256	296

Table 4. A comparative account of nucleotide compositions.

Comparative and polythetic divisive analysis: The Steinernema sp. isolate CS_1 was placed in bicornutum clade that includes species bearing two horn like structure on head region of 3^{rd} stage infective juveniles. Available data of 9 closely related species viz., S. anatoliense, S. carpocapsae, S. kushidai, S. scapterisci, S. tami, S. websteri, pakistanense S. abbasi, S. and S. thermophilum were used for the comparison. Taxometrical characters of 3rd stage juveniles are given in Table 2 and 3. MCD, CD, C-Dis and CS values were calculated for total body length, greatest body width, position of excretory pore and nerve ring from anterior end, oesophagus length; a,b,c ratio and D% and E% based on the data available, as shown in Table 5.

Discussion: Adams (2001) and Sites & Marshall (2004) explored the difficulties of species differentiations in their studies. Morphological and reproductive diversity of nematodes are especially heavy tasks for the accurate description of a new nematode species (Nadler, The specimen 2002). present varied with S. morphologically anatoliense, S. carpocapsae, S. kushidai, S. scapterisci, S. tami and S. websteri based on the presence of 2 horn like structure on the head region of

unsheathed 3rd stage infective juveniles hence was placed in bicornutum group together with S. abbasi, S. pakistanense and S. thermophilum with whom it was compared in detail. The length of juveniles is an indicative character of classification of different groups of Steinernema species. Total body length of Steinernema sp. isolate CS_1 placed the specimen in between the S. abbasi, S. thermophilum and S. pakistanense. Other characters were also showing the divergence from all the compared species along with the species of bicornutum group. However, morphological observations were almost similar and presence of mucronate process in tail region only in 2nd generation female divert the specimen from S. pakistanense and placed near to S. abbasi and S. thermophilum.

Maximum parsimonious tree of ITS regions with 575 positions in final dataset showed closest resemblance of present specimen with *S. abbasi* and *S. thermophilum*. Similar results were obtained from neighbour joining and maximum likelihood method. The distance matrix revealed no difference among the present specimen and *S. abbasi* and *S. thermophilum*. Composition of nucleotides (the GC%, AT% and number of A, C, G, T) were also similar in ITS1-5.8.ITS2 region of the amplified region.

				MO	CD Valı	ies			
Characters	S.	S.	S. kushidai	S.	S.	S.	S.	S.	S.
T	anaioiiense	carpocapsae	<i>kusniaai</i>	scapierisci		websieri	abbasi	pakisianense	inermophium
L	0.18	0.28	0.30	0.20	0.29	0.19	0.19	0.17	0.23
a	0.21	0.16	0.18	0.24	0.22	0.26	0.13	0.17	0.13
b	0.19	0.18	0.18	0.16	0.23	0.18	0.18	0.18	0.19
С	0.22	0.21	0.23	0.22	0.21	0.30	0.24	0.20	0.17
GBW	0.24	0.28	0.26	0.32	0.29	0.26	0.17	0.20	0.16
EP	0.16	0.35	0.20	0.25	0.21	0.25	0.17	0.20	0.22
NR	0.19	0.24	0.21	0.23	-	0.19	0.18	0.16	0.21
ES	0.17	0.29	0.12	0.14	0.11	0.12	0.10	0.12	0.16
Tail	0.22	0.23	0.24	0.21	0.24	0.28	0.19	0.18	0.23
D%	0.19	0.18	0.16	0.22	0.18	0.24	0.15	0.20	0.20
Е%	0.22	0.20	-	-	0.22	0.35	0.19	0.19	0.22
	0.07	0.04	0.01	CD	Values	0.00	0.07	0.44	0.05
L	0.07	0.04	0.01	0.02	0.10	0.00	0.07	0.16	0.05
a	0.08	0.12	0.06	0.01	0.03	0.16	0.28	0.01	0.09
b	0.14	0.29	0.10	0.27	0.16	0.14	0.02	0.02	0.09
с	0.01	0.04	0.12	0.03	0.06	0.19	0.06	0.06	0.17
GBW	0.65	0.63	0.60	0.67	0.71	0.78	0.50	0.56	0.78
EP	0.41	0.43	0.61	0.46	0.38	0.38	0.65	0.75	0.48
NR	0.02	0.09	0.02	0.22	-	0.13	0.13	0.03	0.09
ES	0.07	0.19	0.11	0.25	0.16	0.15	0.11	0.13	0.13
Tail	0.06	0.06	0.12	0.04	0.12	0.18	0.00	0.03	0.22
D%	0.32	0.60	0.17	0.40	0.44	0.44	0.09	0.03	0.05
E%	0.17	0.35	0.07	0.00	0.16	0.11	0.00	0.06	0.05
				C-D	is Valu	es			
L	0.20	0.33	0.36	0.22	0.34	0.20	0.21	0.19	0.26
а	0.24	0.18	0.20	0.28	0.25	0.25	0.14	0.19	0.14
b	0.21	0.20	0.20	0.18	0.27	0.18	0.20	0.20	0.21
с	0.25	0.23	0.26	0.25	0.23	0.30	0.27	0.22	0.19
GBW	0.28	0.33	0.30	0.38	0.34	0.32	0.19	0.23	0.18
EP	0.18	0.44	0.22	0.28	0.23	0.30	0.19	0.22	0.25
NR	0.21	0.27	0.23	0.26	-	0.21	0.20	0.18	0.24
ES	0.19	0.36	0.13	0.15	0.12	0.13	0.10	0.13	0.18
Tail	0.24	0.27	0.27	0.24	0.28	0.33	0.21	0.20	0.26
D%	0.21	0.20	0.18	0.25	0.20	0.28	0.17	0.22	0.22
E%	0.25	0.23	-	-	0.25	0.37	0.21	0.21	0.24
_				CS	Values				
L	0.80	0.67	0.64	0.78	0.66	0.80	0.79	0.81	0.74
а	0.76	0.82	0.80	0.72	0.75	0.75	0.86	0.81	0.86
b	0.79	0.80	0.80	0.82	0.73	0.82	0.80	0.80	0.79
c	0.75	0.77	0.74	0.75	0.77	0.70	0.73	0.78	0.81
GBW	0.72	0.67	0.70	0.62	0.66	0.68	0.81	0.77	0.82
EP	0.82	0.56	0.78	0.72	0.77	0.70	0.81	0.78	0.75
NR	0.79	0.73	0.77	0.74	-	0.79	0.80	0.82	0.76
ES	0.81	0.64	0.87	0.85	0.88	0.87	0.90	0.87	0.82
Tail	0.76	0.73	0.73	0.76	0.72	0.67	0.79	0.80	0.74
D%	0.79	0.80	0.82	0.75	0.80	0.72	0.83	0.78	0.78
E%	0.75	0.77	-	-	0.75	0.63	0.79	0.79	0.76

Table 5. A comparative account of MCD, CD, C-Dis and CS values of infective juveniles of compared species.

*Blank columns indicate the absence of non-availability of data.

The nucleotide composition resemblance within 5.8S gene with other Steinernema species evident the less polymorphism and highly conserved nature of this region as compared to ITS1 and ITS2 delimit the relationship of steinernematid nematodes, results correlated to the study of other researchers (Nguyen et al., 2001; Szalanski et al., 2000). 100% similarity obtained through molecular analysis showed no difference in S. abbasi, S. thermophilum and Steinernema sp. isolate CS_1 and support the synonymizing of S. thermophilum as a junior synonym of S. abbasi (Hunt, 2007). Previously, the study on the same bacterial symbiont was an indicative of the similarity of the S. thermophilum with S. abbasi (Tailliez et al., 2006).

Close resemblance based on C-Dis, CS, CD and MCD values could also found some important part of taxonomical identification where total body length, position of excretory pore and nerve ring from anterior end, oesophagus length; a, b ratio and D% and E% in 3^{rd} stage juveniles also support the resemblance of *Steinernema* sp. isolate CS₁ with *S. abbasi*.

Based on the results obtained through morphology, life history events, taxometry, molecular characterization and numerical taxonomy, the present specimen was concluded as an another isolate of *S. abbasi* with larger infective juveniles form Saharanpur District of Western part of Uttar Pradesh, India.

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References

- Adams, B. J. (2001). The species delimitation uncertainty principle. *Journal of Nematology*, 33, 153-160.
- Adams, B. J. & Nguyen, K. B. (2002). Taxonomy and Systematics. In: Gaugler, R. (Ed.).

Entomopathogenic Nematology, CABI Publishing, Wallington, UK. 1-33 pp.

- Bedding, R. A. & Akhurst, R. J. (1975). A simple technique for the detection of insect parasitic rabditid nematodes in soil. *Nematologica*, 21, 109-110. http://dx.doi.org/10.1163/187529275x00419
- Blaxter, M. L., De Ley, P., Garey, J. R., Liu, L.
 X., Schheldeman, P., Vierstraete, A.,
 Vanfleteren, J. R., Mackey, L. Y., Dorris,
 M., Frisse, L. M., Vida, J. T. & Thomas, W.
 K. (1998). A molecular evolutionary
 framework for the phylum Nematoda. *Nature*, 392, 71-75.
- Courtney, W. D., Polley, D. & Miller, V. I. (1955). TAF an improved fixative in nematode technique. *Plant Disease Reporter*, 39, 570-571.
- Cutler, G. C. & Stock, S. P. (2003). *Steinernema websteri* sp. n. (Rhabditida: Steinernematidae), a new entomopathogenic nematode from China. *Nematologica Mediteranea*, 3, 215-224.
- Elawad, S. A., Ahmad, W. & Reid, A. (1997). *Steinernema abbasi* sp. n. (Nematoda: Steinernematidae) from the Sultanate of Oman. *Fundamental and Applied Nematology*, 20, 433-442.
- Ganguly, S. & Singh, L. K. (2000). *Steinernema thermophilum* sp. n. (Rhabditida: Steinernematidae) from India. *International Journal of Nematology*, 10, 183-191.
- Hazir, S., Stock, S. P. & Keskin, N. (2003). A new entomopathogenic nematode, *Steinernema anatoliense* n. sp. (Rhabditida: Steinernematidae), from Turkey. *Systematic Parasitology*, 55, 211–220. http://dx.doi.org/10.1023/A:102460700376.
- Hominick, W. M., Reid, A. P., Bohan, D. A. & Briscoe, B. R. (1996). Entomopathogenic nematodes: biodiversity, geographical distribution and convention on biological diversity. *Biocontrol Science and Technology*, 6, 317-332. DOI: 10.1080/09583159631307
- Hominick, W. M., Briscoe, B. R., Del Pino, F. G., Heng, J., Hunt, D. J., Kozodoy, E., Mráïcek, Z., Nguyen, K. B., Reid, A. P., Spiridonov, S., Stock, P., Sturhan, D., Waturu, C. & Yoshida, M. (1997). Biosystematics of entomopathogenic nematodes: current status, protocols and definitions. *Journal of*

Helminthology, 71, 271-298. http://dx.doi.org/10.1017/s0022149x00016096

- Hunt, D. J. (2007). Overview of taxonomy and systematics. In: Nguyen, K. B. & Hunt, D. J. (Eds.) Entomopathogenic nematodes: systematics, phylogeny and bacterial symbionts. Nematology Monographs and Perspectives, Volume 5. Leiden, The Netherlands. Brill Publishing, 27-57 pp.
- Joyce, S. A., Reid, A. & Curran, J. (1994).
 Application of the polymerase chain reaction (PCR) methods to the identification of entomopathogenic nematodes. In: Burnell, A. M., Ehlers, R.-U. & Masson, J. P. (Eds.). Genetics of Entomopathogenic nematode-bacterium complexes. Luxembourg, European Commission Publication EUR 15681 EN, 178-187 pp.
- Kaya, H. K. & Gaugler, R. (1993). Entomopathogenic nematodes. *Annual Review of Entomology*, 38, 181-206.
- Luc, P. V., Nguyen, K. B., Reid, A. P. & Spiridonov, S. E. (2000). Steinernema tami sp. n. (Rhabditida: Steinernematidae) from Cat Tien Forest, Vietnam. Russian Journal of Nematology, 8, 33-43.
- Malhotra, S. K., Dixit, S. & Capoor, V. N. (1981). A contribution to the study of taxa differentiation in cestode taxonomy. *Proceedings of Indian Academy of Science* (Animal Sci), 90, 343-349.
- Mamiya, Y. (1988). *Steinernema kushidai* n. sp. (Nematoda: Steinernematidae) associated with scarabaeid beetle larvae from Shizuoka, Japan. *Applied Entomology and Zoology*, 23, 313-320.
- Nadler, S. A. (2002). Species delimitation and nematode biodiversity: phylogenies rule. *Nematology*, 4, 615-625.
- Nei, M. & Kumar, S. (2000). Molecular Evolution and Phylogenetics. Oxford University Press, New York. DOI:10.1093/molbev/mst197.
- Nguyen, K. B., Maruniak, J. & Adams, B. J. (2001). Diagnostic and phylogenetic utility of the rDNA internal transcribed spacer sequences of *Steinernema*. *Journal of Nematology*, 33, 73-82.

- Nguyen, K. B. & Smart, Jr. G. C. (1992). Life cycle of *Steinernema scapterisci* Nguyen & Smart, 1990. *Journal of Nematology*, 24, 160-169.
- Saitou, N. & Nei, M. (1987). The neighborjoining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4, 406-425.
- Seinhorst, J. W. (1959). A rapid method for the transfer of nematodes from fixative to anhydrous glycerin. *Nematologica*, 4, 67-69. http://dx.doi.org/10.1163/187529259x00381.
- Shahina, F., Anis, M., Reid, A. P., Rowe, J. & Maqbool, M. A. (2001). Steinernema pakistanense sp. n. (Rhabditida: Steinernematidae) from Pakistan. International Journal of Nematology, 11, 124-133.
- Sites, Jr. J. W. & Marshall, J. C. (2004). Operational criteria for delimiting species. *Annual Review of Economic and Evolutionary Systematics*, 35, 199-227.

DOI: 10.1146/annurev.ecolsys.35.112202.130128

Sudhaus, W. (1993). Die mittels symbiontischer Bakterien entomopathogenen Nematoden Gattungen Heterorhabditis and Steinernema sindkeine Schwester taxa. Verhandlungen der Deutschen Zoologischen Gesellschaft, 86, 146.

- Szalanski, A. L., Sui, D. D., Harris, T. S. & Powers, T. O. (1997). Identification of cyst nematodes of agronomic and regulatory concern with PCR-RFLP of ITS1. *Journal of Nematology*, 29, 255-267.
- Tailliez, P., Pagès, S., Ginibre, N. & Boemare, N. (2006). New insight into diversity in the genus *Xenorhabdus*, including the description of ten new species. *International Journal of Systematics and Evolutionary Microbiology*, 56, 2805-2818.
- Tamura, K., Nei, M. & Kumar, S. (2004). Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proceedings of the National Academy of Sciences (USA)*, 101, 11030-11035.
- Tamura, K. & Nei, M. (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in

humans and chimpanzees. *Molecular Biology and Evolution*, 10, 512-526.

- Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. (2013). MEGA6: Molecular Evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, 30, 2725-2729. DOI: 10.1093/molbev/mst197.
- Weiser, J. (1955). Neoaplectana carpocapsae n. sp. (Anguillulata; Steinernematidae) novycizopasnichousenikobalecejablecneho, Carpocapsa pomonella L. Vestnik Cesk. Zoologic késpolečnosti, 19, 44-52.
- White, G. F. (1927). A method for obtaining infective nematode larvae from culture. *Science*, 66, 302-303. http://dx.doi.org/10.1126/science.66.1709.3 02-a.
- Woodring, J. L. & Kaya, H. K. (1988). Steinernematid and heterorhabditid nematodes: A handbook of biology and techniques. Southern Cooperative Series Bulletin 331, Arkansas Agricultural Experiment Station, Fayetteville, AR, 30 pp.

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