Effect of papaya and neem seeds on Ascaridia galli infection in broiler chicken

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Abstract

Experiment was carried out to assess the efficacy of ethanolic extract of papaya (*Carica papaya*) and neem (*Azadirachta indica*) plant seeds on *Ascaridia galli* infectivity in broiler chicken. A total of eighteen broiler birds were randomly selected that were divided into three groups (A, B and C) with 6 birds in each group. The birds were then artificially infected with *Ascaridia galli* @ 2000 eggs/bird. Ethanolic extracts of papaya and neem were applied to Group B and C, respectively while Group A was left untreated that served as control. The fecal egg count (FEC) was conducted on weekly basis. The pre-treatment values of FEC in all three groups found negative from day 0 to 14 after artificially inducing infection. On day 21, the FEC pretreatment values in group A, B and C were recorded as 1424.5, 1346.3 and 1806.4, respectively. The FEC post treatment values of groups B and C were significantly (P>0.05) decreased as compared to the control (group A). However, no significant difference was observed between group B and C. It was concluded that the ethanolic extract of both the papaya and neem was effective in controlling the *Ascaridia galli* infection in chicken. However, papaya extract was found more effective than neem.

Keywords: Anthelmintics, Ascaridia galli, neem, papaya, broiler chicken.

The intestinal parasitic nematodes cause severe diseases of poultry birds in developing countries including Pakistan (Basit et al., 2014). Ascaridia galli is the most common and important nematode that causes sustains economic losses to the poultry birds. Birds infected with A. galli usually suffer from severe diarrhea, anemia leading to the loss of body weight resulting onto heavy economic losses in poultry farming (Permin & Raving, 2001) therefore their timely control may help bear better economic benefits. Mostly the synthetic compounds are used for controlling the internal parasites. The use of these compounds affect the drawbacks of negative impact on health of bird itself as well as lead to public health issues with residues of drugs in poultry meat leading to carcinogenesis (Butaye et al., 2003). These compounds also develop resistance against the causative agents.

However, high cost of antihelmintics has diverted the interest towards alternative control practices, including the usage of medicinal plants in traditional remedies. World Health Organization reported that 80% of the rural population in the world has confidence on the herbal remedies for their basic health care. Since the days of human civilization, numerous plants are being used in conventional medicine. The data on usefulness of majority of plants is unavailable. However, the research is going on to confirm the efficacy of various plants (Nayak et al., 2011; Sofowora et al., 2013). The herbal remedies are cost effective, having minimum toxicity with reduced health hazards and easily available in market as compared to synthetic medicines (Khandaker et al., 2016). In the current research study, the efficiency of seed of traditional remedial plants viz., neem (Azadirachta indica) and papaya (Carica papaya) was evaluated for controlling the A. galli parasite in broiler chicken. Papaya fruit has the tremendous health benefits and the leaf and seeds are used against the various health disorders including liver cirrhosis, parasitic infections and digestive disorders. Its anticancer properties have also been reported (Aruoma et al., 2014; Gajowik & Dobrzyńska, 2014). Previously, the effects of leaf on growth performance of chicken have well been documented (Adeyemo & Akanmu, 2012; Kamal et al., 2015; Sorwar et al., 2016).

The medicinal properties of neem tree have been recognized since thousands of years in subcontinent where people are using its bark, root, leaf, seed, oil and kernel for the treatment and control of different parasites of human and animals (El-Tahir et al., 1998; Biswas et al., 2002; Parida et al., 2002; Udeinya et al., 2004; Yanes et al., 2004; Udeinya et al., 2008). The leaf extract as growth promoter has also been reported by many researchers (Adeyemo & Akanmu, 2012; Kharde & Soujanya, 2014; Sarker et al., 2014; Kamal et al., 2015). The antifungal properties of neem seed have been reported by Moslem & El-Kholie (2009), Ospina-Salazár et al., (2015). However, the effectiveness of neem seeds against red mites in poultry has also been observed (Abdel-Ghaffar et al., 2008). In present investigation controlling potential of papaya and neem seed's ethanolic extracts have been evaluated against the A. galli in broiler chicken.

Materials and Methods

The study was carried out at Department of Veterinary Parasitology, Sindh Agriculture University, Tandojam during 2015.

Sample size and experimental birds: Three weeks old, 18 commercial broiler chicks were bought from market and placed them into 3 groups, viz., A, B and C, each comprising of 6 birds. The birds were reared in experimental sheds of Parasitology Department with equal

management environment. The birds were vaccinated against Coccidiosis, Gumboro, New Castle Disease, *E. coli*, Salmonella and other bacterial infections.

Collection of the A. galli eggs: The eggs were obtained from infected birds, processed for direct fecal examination, identified with published keys. Briefly, a small drop of equal amount of water and feces was thoroughly mixed to obtain a fairly homogenous and adequately clear preparation and placed on a microscope slide. A cover slip was placed on fluid, allowed to settle for few minutes and was then observed magnification under lower (10x)of compound microscope. A. galli eggs were identified by size, shape and colour of the shell using the key provided by Soulsby (1983).

Ethanolic extraction of the plant seeds: The seeds of papaya and neem were collected from field. The collected seeds were dried under shade at existing air temperature of the surrounding area, minced to powder by electrical blender and stored in air tight glass bottles.

Extraction from seeds powder was carried out by following the procedure given before (AOAC, 2000; Wang & Waller, 2006). Briefly, 4g of a moisture free sample was measured in extraction thimble, plugged with absorbent cotton wool and then placed in an extractor that was fixed under the condenser of the extraction apparatus. Receiving flask contained 150 ml of the solvent was connected to the apparatus and placed on water bath at 40 °C with the rate of condensation at 3-4 drops per second. After 10 hours, thimbles from extractors were removed and just before drying the solvent, extract was transferred into a clean tarred evaporating basin with ether washing. Basin was placed in oven at 105°C for 2 hrs and then cooled in desiccators for 30 min. Extraction was measured as follows:

% Ether extract	Weight of ether extract	100
(DM) =	Sample weight	- x 100

Inoculation of experimental birds: All the experimental birds were artificially infected orally with 2000 A. galli eggs. The fecal samples of all birds were collected and checked on a regular basis for confirmation of A. galli eggs. On day 21 (after confirmation of infection) the groups B and C were treated with extraction of @ 20 ml/kg, papaya and neem seeds respectively. While group A received no treatment and served as control. The fecal examination was continued up to 42 days and the Fecal Eggs Counts - FEC (quantitative examination) was carried out by McMaster Technique according to the method given by Urquhart et al., (1996).

Statistical analyses: Data analysis was done through statistical program Graphed Prism (Graphed Software Incorporated 2000) by oneway analysis of variance (ANOVA) and Chisquare analysis (x-analysis) t-test.

Results

Effect of medicinal plants on Ascarida galli infection: In present study the seeds of two plants viz., neem and papaya were used against A. galli infection in chicken. The aim of the present study was to explore the potential of these plants to control the A. galli infection in chicken.

In vivo study: Table 1 represents the data of all experimental birds shading *A. galli* eggs from day 1 to 42 recorded on weekly basis. The groups A (control), B (infected-papaya treated) and C (infected-neem treated) showed no egg shedding from day 1 to 14 after inoculation. However, the egg shedding was started from day 21^{st} and the pretreatment fecal egg count (FEC) of groups A, B and C was recorded as 1424.5, 1346.3 and 1806.4, respectively. It was further revealed that the egg shedding was significantly (P>0.05) reduced after the treatment of seed

extracts and lower FEC was recorded on days 28, 35 and 42 in group B and C while group A showed continued increase in FEC up to the end of trial. The egg per gram was recorded on day 28 (A, 1705.2; B, 1013; C, 1416.3); 35 (A, 2166.2; B, 233; C, 399.75) and 42 (A, 2606.4; B, 6.6; C, 41.5). The FEC of group B (99.5%) and C (97.7%) was significantly reduced as compared to the group A. The comparisons of groups A *vs* B and A *vs* C was very significant (P<0.0001) but group B *vs* C didn't show any significant difference (P>0.05).

Discussion

The use of medicinal plants against various diseases of humans and animals is widely documented. Various parts of medicinal plants have shown anti parasitic characteristics and have usually been applied as a remedy against internal and external parasites (Bauri et al., 2015). Many workers demonstrated the medicinal effect of numerous plants, including papaya and neem, against A. galli infection in poultry and some other animals (Akhter & Riffat, 1984; Muhamd & Riffat, 1985; Hafeez & Venkataratnam, 1989; Hammond et al., 1997; Purwati & He, 1991; Taylor et al., 2001; Lateef, 2002; Rahman, 2002; Adu et al., 2002; Hordegen et al., 2003; Beloin et al., 2005; Iqbal et al., 2005; Hordegen et al., 2006; Ali, 2006; Das et al., 2006; Fakim, 2006; Siamba et al., 2007; Thomas et al., 2007; Shahadat et al., 2008; Islam et al., 2008; Satish et al., 2009; Adu et al., 2009; Ayub et al., 2011; Adeyemo & Akanmu, 2012; Kamal et al., 2015). The results of present study, revealed that on day 21 of artificial inoculation of A. galli, the infection established and EPG of groups A, B & C was 1424.5, 1346.3 and 1806.4, respectively. On the same day, the treatment of papaya and neem seed extract was given to group B and C, respectively and significant (P>0.05; P>0.01) decrease in parasitic burden was recorded in the following days while in group A (controlwithout treatment), the significant increase in infection was observed. On day 42, the EPG in group B (papaya) and C (neem) was found to

Day	Group A (EPG) Control	Group B (EPG) Papaya treated	Group C (EPG) Neem treated
1	0	0	0
7	0	0	0
14	0	0	0
21	1424.5 <u>+</u> 367.46	1346.3 <u>+</u> 45.88	1806.4 <u>+</u> 249.96
28	1705.2 <u>+</u> 357.50**	1013.0 <u>+</u> 215.50**	1416.3 <u>+</u> 38.13
35	2166.2 <u>+</u> 228.39***	233.00 <u>+</u> 52.81***	399.75 <u>+</u> 118.38***
42	2606.4 <u>+</u> 244.39***	6.600 <u>+</u> 14.75*** (99.5% decreased)	41.500 <u>+</u> 49.89*** (97.7% decreased)

Table 1. Screening of A. galli in experimental chickens.

decrease 99.5 and 97.7%, respectively (Table 1). It was shown that the ethanolic extraction of papava and neem seeds were effective against A. galli in chicken although papaya was more effective than neem. Adu et al., (2009) used extraction from latex of papaya and reported 77.7% decrease in EPG of A. galli in the feces of chicken. Present observations revealed strong effectiveness of papaya seeds against A. galli infection. Our observation is in-agreement with the study of Ameen et al., (2012) who demonstrated successful control of helminths including A. galli through extraction of papaya seeds. A successive control of A. galli was also reported (in-vitro) by papaya seed extract (Alam et al., 2014).

Previous studies have shown the effectiveness of neem leaves extract to control the *A. galli* infection in poultry (Khokan *et al.*, 2014) and also act as growth promoter (Adeyemo & Akanmu, 2012; Kamal *et al.*, 2015), antifungal (Moslem & El-Kholie, 2009; Ospina-Salazár *et al.*, 2015). Subapriya & Nagini (2005) reviewed the activities of neem leaf extract as antiviral, antibacterial, anti-hypertensive, antifungal,

anti-malarial, anti-fertility, antipyretic, anti-inflammatory and antianalgesic. ulcerogenic. Neem leaves extract proved 22-30% more effective than commercial anthelmintic for the control of gastrointestinal parasites in sheep and goat also (Thomas et al., 2007). In present study, papaya was proved to be more effective as compared to neem for controlling the A. galli infection. These findings are thus in conformity with Alam et al., (2014) who testified the various medicinal plants against A. galli and demonstrated the best control with papaya seed extract followed by neem. Islam et al., (2008) also reported that the papaya leaf extract is more effective than neem against A. galli eggs.

Conclusion

It is concluded that ethanolic seed extracts of *Azadirachta indica* (neem) and *Carica papaya* (papaya) are effective against the *A. galli* infection in chicken. However, the papaya extract was found to be more effective. However, more research towards particular active compound against the gastrointestinal parasites in poultry is needed.

References

- Abdel-Ghaffar, F., Sobhy, H. M., Al-Quraishy, S. & Semmler, M. (2008). Field study on the efficacy of an extract of neem seed (Mite-Stop) against the red mite *Dermanyssus gallinae* naturally infecting poultry in Egypt. *Parasitology Research*, 103, 481-485.doi: 10.1007/s00436-008-0965-9
- Adeyemo, G. O. & Akanmu, A. M. (2012). Effects of neem (Azadirachta indica) and pawpaw (Carica papaya) leaves supplementation on performance and carcass characteristics of broilers. International Journal ofCurrent Research, 4, 268-271.
- Adu, O. A. & Akingboye, A. K. (2002). Anthelmintic efficacy of papaya (*Carica papaya*) latex in Pacoc. 7th Annual Conference of Animal Science Association of Nigeria, 16-19.
- Adu, O. A., Akingboye, K. A. & Akinfemi, A. (2009). Potency of pawpaw (*Carica papaya*) latex as an anthelmintic in poultry production. *Botany Research International*, 2, 139-142.
- Akhter, S. M. & Riffat, S. (1984). Efficacy of Melia azedarach, Linn. (Bakain) and Morantel against naturally acquired gastrointestinal nematodes in goats. Pakistan Veterinary Journal, 4, 176-179.
- Ali, M. A. (2006). In vitro anthelminic effects of some indigenous plants against Ascaridia galli of indigenous chickens.
 MS thesis. Department of Parasitology, Bangladesh Agricultural University, Mymensingh.
- Ameen, S. A., Adedeji, O. S., Ojedapo, L. O., Salihu, T. & Fakorode, O. L. (2012).
 Anthelmintic efficacy of pawpaw (*Carica papaya*) seeds in commercial layers. *African Journal of Biotechnology*, 11, 126-13.
- AOAC, (2000). *Methods of analysis*, 15th Edition. Association of Official Analytical Chemists, Arlington.

- Aruoma, O. I., Somanah, J., Bourdon, E., Rondeau, T. & Bahorun, T. (2014). Diabetes as a risk factor to cancer: Functional role of fermented papaya preparation as phytonutraceutical adjunct in the treatment of diabetes and cancer. *Mutation Research*, 768, 60-68.
- Alam, M., Alam, K., Begum, N. & Amin, M. (2014). Comparative efficacy of different herbal and modern anthelmintics against gastrointestinal nematodiasis in fowl. *International Journal of Biological Research*, 2, 145-148.
- Ayub, A. M., Inaotombi Devi, L., Lyngdoh, W. M., Das, G., Prasad, H., Chanu, K. H. V., Prava, M., Tolenkhomba, T. C., Singh, Y. D. & Lallinchhunga, M. C. (2011). Comparative biochemical profile of *Ascaridia galli* infected broiler chickens on administration of pineapple and neem leaves and piperazine. *International Journal of Poultry* Science, 10, 542-546.
- Basit, A., Ali, A. A., Malik, M. S., Malik, A. N., Iftikhar, M., Anwar ul Haq, H. M. & Nadeem, S. M. (2014). A study of gastro–intestinal helminths in native peafowl and comparative efficacy of Albendazol and a Pyrantelpamoate against the helminth parasites. *Journal of Infection and Molecular Biology*, 2, 22-25.
- Bauri, R. K., Tigga, M. N. & Kullu, S. S. (2015). A review on use of medicinal plants to control parasites. *Indian Journal* of Natural Products Resources, 6, 268-277.
- Beloin, N., Gbeassor, M., Akpagana, K., Hudson, J., Soussa, K., Koumaglo, K. & Arnason, J. T. (2005). Ethnomedicinal uses of *Momordica charantia* (Cucurbitaceae) in Togo and relation to its phytochemistry and biological activity. *Journal of Ethnopharmacology*, 496, 49-55.
- Biswas, K., Chattopadhyay, I., Banerjee, R.
 K. & Bandyopadhyay, U. (2002).
 Biological activities and medicinal properties of neem (*Azadirachta indica*). *Current Science*, 82, 1336-1345.

- Butaye, P., Devriese, L. A. & Haesebrouck, F. (2003). Antimicrobial growth promoters used in animal feed: Effects of less well known antibiotics on gram positive bacteria. *Clinical Microbiology Reviews*, 16, 175-188.
- Das, P., Sinhababu, S. P. & Dam, T. (2006). Screening of anthelminthic effects of Indian plant extracts, a preliminary report. *Journal of Alternative and Complimentary Medicine*, 12, 299-301. *doi.*org/10.1645/GE-1446.1
- El-Tahir, A., Ibrahim, A. M., Satti, G. M. H., Theander, T. G., Kharazmi, A. & Khalid,
 A. S. (1998). The potential antileishmanial activity of some Sudanese medicinal plants. *Phytotherapy Research*, 12, 576-579. DOI: 10.1002/(SICI)10991573(199812)12:8<5 76
- Fakim, A. G. (2006). Medicinal plants: Traditional of yesterday and drugs of tomorrow. *Molecular Aspects of Medicine*, 27, 1-93.
- Gajowik, A. & Dobrzynska, M. M. (2014). Lycopene-antioxidant with radioprotective and anticancer properties. *Roczniki Państwowego Zakładu Higieny*, 65, 263-71.
- Hafeez, M. & Venkataratnam, A. (1989).
 Wopell-an effective anthelmintic against Ascaridiasis in poultry. Indian Journal of Indigenous Medicine, 6, 9-12.
- Hammond, J. A., Fielding, D. & Bishop, S. C. Prospects (1997).for plant anthelmintics tropical veterinary in medicine. Veterinary Research 213-228.doi: Communications, 21. 10.1023/A:1005884429253
- Hordegen, P., Hertzberg, H., Heilmann, J., Langhans, W. & Maurer, V. (2003). The anthelmintic efficacy of five plant products against gastrointestinal *Trichostrongyloids* in artificially infected lambs. *Veterinary Parasitology*, 117, 51-60.
- Hordegen, P., Cabaret, J., Hertzberg, H., Langhans, W. & Maurer, V. (2006). In

vitro screening of six anthelmintic plant products against larval *Haemonchus* contortus with a modified methylthiazolyl-tetrazolium reduction assay. Journal of Ethnopharmacology, 108, 85-89.

- Iqbal, Z., Jabbar, A., Akhter, M. S., Muhammad, G. & Lateef, M. (2005).
 Possible role of ethnoveterinary medicine in poverty reduction in Pakistan: Use of botanical, anthelmintic as an example. *Journal of Agriculture & Social Sciences*, 1, 188-195.
- Islam, K. R., Farjana, T., Begum, N. & Mondal, M. M. H. (2008). *In vitro* efficacy of indigenous plants on the inhibition of development of eggs of *Asciridia A. galli* (Digenia; Nematode). *Bangladesh Journal of Veterinary Medicine*, 6, 159-167.
- Kamal, M. M., Boby, F., Shahjada, Z., Lutfuzzaman, A. K. M., Abdullah-Al-Motin, Alam, K. J. & Mostofa, M. (2015). Effects of neem, nishyinda and papaya leaves as growth promoters in broiler chicks. *International Journal of Natural and Social Sciences*, 2, 49-55.
- Khandaker, S., Das, S., Opo, F. A. D. M., Akhter, R. & Shahriar, M. (2016). *In vivo* pharmacological investigations of the crude extracts of *Calamus viminalis* (L.). *Journal of Pharmacognosy and Phytochemistry*, 5, 263-269.
- Kharde, K. R. & Soujanya, S. (2014). Effect of garlic and neem leaf powder supplementation on growth performance and carcass traits in broilers. *Veterinary World*, 7, 799-802.
- Khokon, J. U., Sharifuzzaman, Sarker, E. H., Rahman, M. A., Kisku, J. J. & Mustofa, M. (2014). Efficacy of neem leaf extract against ascariasis in indigenous chicken. *International Journal of Natural & Social Sciences*, 1, 25-30.
- Lateef, M. (2002). Trichostrongyloid nematodes of sheep: Epidemiological aspects and evaluation of anthelmintic

activity of indigenous plants. Ph. D Thesis, Univ. Agri., Faisalabad, Pakistan.

- Moslem, M. A. & El-Kholie, E. M. (2009). Effect of neem (Azardirachta indica A. Juss) seeds and leaves on some plant pathogenic fungi. Pakistan Journal of Biological Sciences, 12, 1045-1048.
- Muhammad, S. A. & Riffat, S. (1985). Evaluation of *Melia azedarach* Linn, fruit (Bakain) against *Ascaridia galli* infection in chicken. *Pakistan Veterinary Journal*, 5, 23-45.
- Nayak, A., Nayak, R. N., Somya, G. B., Kishore, B. & Mithun, K. (2011). Evaluation of antibacterial and anticandidial efficacy of aqueous and alcoholic extracts of neem (*Azadirachta indica*) an *in-vitro* study. *International Journal of Research in Ayurveda and Pharmacy*, 2, 230-235.
- Ospina-Salazár, D. I., Hoyos-Sánchez, R. A., Orozco-Sánchez, F., Arango-Arteaga, M. & Gómez-Londoño, L. (2015). Antifungal activity of neem (*Azadirachta indica*: Meliaceae) extracts against dermatophytes. *Acta Biologica Colombiana*, 20, 201-207. doi.org/10.15446/abc.v20n3.45225
- Parida, M. M., Upadhyay, C., Pandya, G. & Jana, A. M. (2002). Inhibitory potential of neem (*Azadirachta indica* Juss) leaves on dengue virus type-2 replication. *Journal of Ethnopharmacology*, 79, 273-8.
- Purwati, E. & He, S. (1991). A potential role of papaya latex as an anthelmintic against *Ascaridia galli* infection in chicken. *Hemara Zoa*, 74, 1-5.
- Rahman, M. (2002). In-vitro and in-vivo anthelmintic effects of some plants against gastro intestinal nematodes of goats. MS thesis, Department of Parasitology, Bangladesh Agriculture University, Mymensingh. pp. 108.
- Permin, A. & Raving, H. (2001). Genetic resistance to Ascaridia galli infection in chickens. Veterinary Parasitology, 102, 101-111.

- Sarker, S. K., Mostofa, M., Akter, F., Rahman, M. M. & Sultana, M. R. (2014). Effects of aqueous extract of neem (*Azadirachta indica*) leaves as growth promoter and anti-colibacillosis in broilers. Bangladesh Journal of Animal Sciences, 43, 138-14.
- Shahadat, H. M., Mostofa, M., Mamun, M. A.
 A., Hoque, M. E. & Awal, M. A. (2008).
 Comparative efficacy of corolla (*Momordica charantia*) extract and ivermec pour on with their effects on certain blood parameters and body weight gain in indigenous chicken infected with Ascaridia galli. Bangladesh Journal of Veterinary Medicine, 6, 153-158.
- Siamba, D. N., Okitoi, L. O., Watai, M. K., Wachira, A. M., Lukibisi, F. B. & Mukisira, E. A. (2007). Efficacy of **Tephrosia** vogelli and Vernonia amvgdalina as anthelmintics against Ascaridia galli in indigenous chicken. Livestock Research Rural for Development, 19, 12.
- Sofowora, A., Ogunbodede, E. & Onayad, A. (2013). The role and place of medicinal plants in the strategies for disease prevention. *African Journal of Traditional, Complementary and Alternative medicines*, 10, 210-229. doi.org/10.4314/ajtcam.v10i5.2.
- Sorwar, M. G., Mostofa, M., Hasan, M. N., Billah, M. & Rahman, M. T. (2016). Effect of kalo jeera seeds and papaya leaf supplementation on the performance of broiler. *Bangladesh Journal of Veterinary Medicine*, 14, 37-42.
- Soulsby, E. J. L. (1983). *Helminths, Athropods and Protozoa of Backyard Animals*.7th Edition. Lea & Febiger, Philadelphia, pp. 156-239.
- Subapriya, R. & Nagini, S. (2005). Medicinal properties of neem leaves: a review. *Current Medicinal Chemistry-Anti-Cancer Agents*, 5, 149-156.
- Taylor, J. L. S., Rabe, T., McGaw, L. J., Jager,A. K. & Van Staden, J. (2001). Towardsthe scientific validation of traditionalmedicinal plants. *Plant Growth*

Regulation, 34, 23-7.doi:10.1023/A:1013310809275.

- Thomas, N., Teshale, S. & Kumsa, B. (2007). Abomasal nematodes of sheep and goats slaught. *Helminthologia*, 44, 70-75.
- Udeinya, I. J., Mbah, A. U., Chijioke, C. P. & Shu, E. N. (2004). An antimalarial extract from neem leaves is antiretroviral. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 98, 435-437.
- Udeinya, J. I., Shu, E. N., Quakyi, I. & Ajayi, F. O. (2008). An antimalarial neem leaf extract has both schizonticidal and gametocytocidal activities. *American Journal of Therapeutics*, 15, 108-110.

- Urquhart, G. M., Armour, J., Dunncan, J. L. & Jennings, F. W. (1996). *Veterinary Parasitology*, 1st Ed. ELBS, Longman, London, UK, pp 307.
- Wang, L. & Weller, L. C. (2006). Recent advances in extraction of naturaceuticals from plants. *Trends in Food Sciences & Technology*, 17, 300-312.
- Yanes, A., Finol, H. J. & Hasegawa, M. (2004). Effects of Azadirachta indica and Melia azedarach (Meliaceae) extracts from leaves on Trypanosoma cruzi growth and ultrastructure. Journal of Submicroscopic Cytology and Pathology, 36, 149-154.

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