



# Impact of Locally Characterized Protease from *Geobacillus* SBS 4S on the Growth of Poultry Bird

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## ABSTRACT

Present study was conducted to determine the efficacy of novel thermostable protease in poultry production. The protease was produced from *Geobacillus* sp. SBS-4S using Luria Bertani medium and it was used as supplement in poultry feed trial. The data were statistically analyzed by Multivariate Analysis of Variance. For feeding, 150 day-old broiler chicks were divided randomly into 5 groups having 30 chicks each. Group A served as negative control, group B, C and D as experimental groups which were supplemented in the basal diet with 2500, 5000 and 7500 IU/kg of locally characterized thermostable protease while group E was supplemented with the commercially available neutral protease (5000 IU/kg) that served as a positive control. The trial lasted for five weeks and during the trial period birds were offered *ad libitum* access to water and feed. The maximal weight gain was recorded when the feed was supplemented with 7500 IU/Kg of locally characterized protease. The evaluation of data of various groups obtained after 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> weeks of trial demonstrated significant change ( $P \leq 0.05$ ) in weight gain, feed up take and feed conversion ratio (FCR) as compared to negative control. Locally produced protease showed promising results as compared to positive control which is being used currently in the poultry industry. The weight gain, feed intake and feed conversion ratio of 1891.54g, 3750g and 1.98 of experimental group D were comparable to 1874.49g, 3740 g and 1.995 in case of positive control. The ability of protease to enhance weight gain, feed consumption and FCR in poultry birds makes it a strong candidate for the replacement of imported counterpart being utilized currently in poultry feed industry.

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## Authors' Contribution

WA performed experimental work. M.T. planned and supervised the study and guided for manuscript write-up and editing. MNA, ASH, MDA, SF, SS, BM and MT helped in facilitated the student for the conduction of experiments. MW and ARA helped in manuscript write up. MA helped in data analysis.

## Key words

Protease, *Geobacillus* sp. SBS-4S, broiler, Weight gain, Feed supplementation, Evaluation

## INTRODUCTION

Poultry sector is one of the fast growing industries of Pakistan that is playing a significant role in bridging gap

in meat demands at economical prices (Hussain *et al.*, 2015). However, the increased demand of feed ingredients has resulted in high feed cost and narrowing down the profit margin (Thirumalaisamy *et al.*, 2016). The plant source proteins derived from grains and soyabean contain various complex molecules which are not being hydrolyzed by the mono-gastric animals due to low or unavailability of enzymes. This problem can be overcome by supplementing the poultry feed with hydrolytic enzymes, responsible for the breakdown of complex to simple absorbable monomeric components.

The research on utilization of exogenous enzymes in poultry feed to decrease the total nutrient excretion and to enhance nutrient utilization by improving the digestive

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process has been ongoing for the past several years (Leeson and Summers, 2005; Cambell and Bedford, 1992; Rada *et al.*, 2016; Ganapathi *et al.*, 2014; Seskeviciene *et al.*, 1999; Toghiani *et al.*, 2017). During the first week, the broiler chicken's gastro intestinal tract is not fully developed. Therefore, the optimal level of enzymatic secretions is not gained (Mahagna *et al.*, 1995). This deficiency of endogenous enzymes is further augmented due to high demand of amino acids and fast passage rate by the body of broiler chicks (Doskovic *et al.*, 2013).

There are reports on digestibility of amino acids and crude proteins indicating that considerable amounts of proteins (18-20%) travel through the gastrointestinal tract without being digested completely (Lemme *et al.*, 2004; Wang and Parsons, 1998; Angel *et al.*, 2011; Applegate and Angel, 2014). This aggravated amino acid demand can be fulfilled either by utilizing higher protein content or by supplementing the feed with protease for aiding the endogenous proteolytic system for the enhanced digestion of dietary protein. Therefore, poultry feed has been supplemented with exogenous proteases to improve nutritive value of poultry diet (Cowieson and Roos, 2016). The use of synthetic enzymes significantly decrease the amounts of excreted nutrients such as phosphorous and nitrogen and stimulate the digestibility of nutrients resulting in reduction of anti-nutritional impact of complexation of phosphorous and amino acids (Sabir *et al.*, 2018; Khalid *et al.*, 2019; Costa *et al.*, 2006; Ghazi *et al.*, 2002).

The feed supplemented with proteases significantly improves the body weight, feed conversion ratio and protein digestibility of feed stuffs in poultry chicks (Oxenboll *et al.*, 2011; Fru-Nji *et al.*, 2011; Puente and Lopez, 2004; Angel *et al.*, 2011; Freitas *et al.*, 2011; Bolan *et al.*, 2010; Ndazigaruye *et al.*, 2019; Mahmood *et al.*, 2017). The increased digestibility of poultry birds increases the possibility of reduced nitrogen excretion by the bird's body, reduced ammonia production from the excreta of animals, decrease the content of protein wasted and cost of production (Oxenboll *et al.*, 2011). The addition of protease to the poultry feed shows more prominent and positive effects on feed conversion ratio (FCR), which leads to the significant environmental benefits (Angel *et al.*, 2011; Fru-Nji *et al.*, 2011). Many scientists have worked on the characterization of proteases for the poultry feed industry but unfortunately none of the protease is available in the market for the fulfilment of local industrial demand (Ali *et al.*, 2016; Aftab *et al.*, 2006; Mukhtar and Haq, 2012; Ahmad *et al.*, 2020). Moreover, Pakistan is spending a huge foreign exchange for the import of the enzymes per annum. The present study was planned to utilize the locally characterized protease from *Geobacillus* sp. SBS-4S for the evaluation of its efficacy on poultry bird's growth. This

protease has been characterized which showed its optimal activity at 65°C and pH 9 in the presence of  $Mn^{2+}$  (Ahmad *et al.*, 2020).

## MATERIALS AND METHODS

### *Protease production*

The protease was produced under pre-optimized conditions from 34 liters of LB medium supplemented with 2% yeast extract and 5% wheat bran, inoculated with the overnight grown *Geobacillus* sp. SBS-4S cells followed by incubation at 60°C for 18h under shaking conditions at 120rpm in 5 batches. The microbial growth was centrifuged and supernatant was assessed for protease activity (Pant *et al.*, 2015; Sharma *et al.*, 2017; Catara *et al.*, 2003). Protein contents were calculated by Bradford method using bovine serum albumin as a standard (Bradford, 1976).

### *Feed formulation*

Broiler feed was formulated as per recommendations of American National Research Council (NRC, 2014) and is being used by the poultry industry (corn, 58.0%; soya bean meal 46, 29.5%; rice polish 14, 7.50%; calcium carbonate ( $CaCO_3$ ), 0.95%; oil, 2.00%; bone ash, 0.75%; DCP (dicalcium phosphate), 0.50%; vitamin premix, 0.30%; L-HCl (lysine HCl), 0.25%; sodium bicarbonate ( $NaHCO_3$ ), 0.17%; DLM (DL-methionine), 0.15%). The poultry feed was prepared in an automated unit at Crescent Feeds and Allied Products, Sundar Sharif, Lahore, Punjab, Pakistan. Five diets were prepared. Diet I was not supplemented with the protease (negative control) while diets II, III and IV were supplemented with protease 2500, 5000 and 7500 IU/kg of feed, respectively. Diet V was supplemented with 5000 IU/kg of commercially available protease as positive control.

### *Feeding trials on broiler chicks*

Feeding trial was conducted under controlled environment at Dua Poultry Farms, Niaz Kot, Kala Shah Kaku, Tehsil Ferozewala, District Sheikhpura, Punjab, Pakistan in collaboration with Crescent Feeds and Allied Products, Sundar Sharif, Lahore, Punjab, Pakistan. For feeding trial, 150 one-day-old broiler chicks with an average weight of 39g per chick were purchased from a commercial hatchery and randomly divided into 5 groups A, B, C, D and E. Each group consisted of 30 birds, having 3 replicates of 10 birds each. Group A was declared as negative control and was fed on diet without protease. Group B, C and D were fed on diets II, III and IV respectively, which were supplemented with 2500, 5000 and 7500 IU/kg of locally produced protease. Group E acted as positive control and was fed on diet V supplemented with commercially available protease (5000

IU/kg of feed). Water and feed were given *ad libitum* during the feeding trial for a period of 5 weeks. The body weight, feed consumption and feed conversion ratio were recorded on week basis (Sabir *et al.*, 2018; Rahman *et al.*, 2014).

#### Statistical analysis

The obtained data was analyzed statistically by Multivariate Analysis of Variance using SPSS software (Okafor and Anosike, 2012). The results obtained were represented as significance of differences between means calculated by Fisher's Least Significant Difference test. The differences were significant at  $P \leq 0.05$  (Steel *et al.*, 1997).

## RESULTS

Feeding trials on poultry birds demonstrated that supplementation of broiler feed with locally characterized protease significantly enhanced the weight gain and feed consumption of poultry birds with improved feed conversion ratio. The weight gain data regarding 1<sup>st</sup> and 2<sup>nd</sup> week of trials showed significant increase in weight gain of various groups fed on diet supplemented with locally characterized protease as compared to negative control (Group A). Significant impact on weight gain of birds was recorded by the end of 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> weeks of experiment (Table I). Comparison of group A (negative control) with group C in the 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> weeks of trials demonstrated the weight gain from 831.66±0.76 to 857.41±0.56g, 1256.35±0.85 to 1291.45±0.74g and 1744.52±0.94 to

1847.55±0.95g, respectively when the poultry diet was supplemented with protease 5000 IU/kg of feed. The comparison of weight gain data of group D with that of group A, B and C demonstrated a boosting effect of locally produced protease on weight gain in poultry birds (Table I).

The increase in protease concentration from 5000 IU/kg (group C) to 7500 IU/kg (group D) showed significant enhancement regarding weight gain and feed consumption. Supplementation of feed with protease enhanced the digestive ability and feed consumption of birds. Comparative analysis indicated the improved feed consumption with the increase in protease concentration. The results demonstrated that group B, C and D utilized 3710, 3730 and 3750 g of feed as compared to group A which utilized 3680 g of feed by the end of 5 weeks of trials. Same pattern of improvement of FCR was recorded in the groups fed on feed supplemented with locally produced protease. The FCR value was improved from 2.11 (group A) to 1.98 (group D) (Table II). A comparison of group D and E revealed the better performance of locally produced protease as compared to enzyme currently being used in the poultry feed industry. Multivariate analysis of variance demonstrated the significant results for various tests (Table III) which supports the positive impact on weight gain due to supplementation of feed with local protease. Multiple comparative analysis among the weeks with respect to various groups showed significant results (Tables III, Supplementary Table I).

**Table I. Effect of protease supplementation on weight of poultry chicks during feeding trials.**

Groups	1 <sup>st</sup> Week (0.000***)	2 <sup>nd</sup> Week (0.000***)	3 <sup>rd</sup> Week (0.000***)	4 <sup>th</sup> Week (0.000***)	5 <sup>th</sup> Week (0.000***)
A (-ve control)	164.68±0.96 <sup>c</sup>	448.49±0.66 <sup>c</sup>	831.66±0.76 <sup>c</sup>	1256.35±0.85 <sup>c</sup>	1744.52±0.94 <sup>c</sup>
B (2500 IU/kg)	171.56±0.68 <sup>d</sup>	458.49±0.68 <sup>d</sup>	847.42±0.80 <sup>d</sup>	1269.44±0.90 <sup>d</sup>	1826.60±0.66 <sup>d</sup>
C (5000 IU/kg)	175.57±0.73 <sup>c</sup>	467.43±0.74 <sup>c</sup>	857.41±0.56 <sup>c</sup>	1291.45±0.74 <sup>c</sup>	1847.55±0.95 <sup>c</sup>
D (7500 IU/kg)	184.43±0.73 <sup>a</sup>	485.49±0.81 <sup>a</sup>	887.52±0.79 <sup>a</sup>	1356.50±0.68 <sup>a</sup>	1891.54±0.89 <sup>a</sup>
E (5000 IU/kg) (+ive control)	183.31±0.64 <sup>b</sup>	473.59±0.70 <sup>b</sup>	872.42±0.78 <sup>b</sup>	1341.19±0.93 <sup>b</sup>	1874.49±0.75 <sup>b</sup>

\*P values indicate the significance among groups of each week. \*\*Means that do not share a letter i.e., a, b, c, d and e are significantly different. Group A, negative control; Group B, 2500 IU/kg; Group C, 5000 IU/kg; Group D, 7500 IU/kg; Group E, +ve control.

**Table II. Effect of protease on average feed intake, feed conversion ratio and body weight during feeding trials.**

Groups	A	B	C	D	E
Average feed intake (g)	3680	3710	3730	3750	3740
Overall body weight (g)	1744.52±0.94	1826.60 <sup>d</sup> ±0.66	1847.55 <sup>c</sup> ±0.95	1891.54 <sup>a</sup> ±0.89	1874.49 <sup>b</sup> ±0.75
Feed conversion ratio (FCR)	2.11 <sup>c</sup>	2.031 <sup>d</sup>	2.019 <sup>c</sup>	1.98 <sup>b</sup>	1.995 <sup>a</sup>

Group A, negative control; Group B, 2500 IU/kg; Group C, 5000 IU/kg; Group D, 7500 IU/kg; Group E, +ve control.

**Table III. Multivariate analysis of variance for multivariate tests.**

Effect		Value	F	Hypothesis df	Error df	Sig.
Intercept	Pillai's trace	1.000	20606492.318 <sup>b</sup>	5.000	141.000	.000
	Wilks' lambda	.000	20606492.318 <sup>b</sup>	5.000	141.000	.000
	Hotelling's trace	730726.678	20606492.318 <sup>b</sup>	5.000	141.000	.000
	Roy's largest root	730726.678	20606492.318 <sup>b</sup>	5.000	141.000	.000
	Pillai's trace	1.647	20.151	20.000	576.000	.000
	Wilks' lambda	.003	119.073	20.000	468.594	.000
	Hotelling's trace	178.024	1241.714	20.000	558.000	.000
	Roy's largest root	177.003	5097.697 <sup>c</sup>	5.000	144.000	.000

a, Design: Intercept + Groups; b, Exact statistic; c, The statistic is an upper bound on F that yields a lower bound on the significance level.

## DISCUSSION

Proteases play essential role in the maturation, working and degradation of proteins in the cell and are key component of cell (Douglas *et al.*, 2000; Ghazi *et al.*, 2002). Proteases are also responsible for the removal of structural proteins in the cell wall polysaccharides which allow faster access to other catabolic enzymes which results in better digestion (Colombatto and Benuchemin, 2009).

Poultry trial confirmed the ability of locally produced protease to improve the digestibility of feed, as supplementation of feed boosted the weight gain and feed uptake of the broiler birds. The comparative analysis of data regarding weight gain of chicks from groups B, C and D with respect to group A (Table I) clearly demonstrated an increase in weight gain with a maximum weight gain of 1891.54±0.89 g (group D) which was comparable to 1874.49±0.75 g (group E) fed on feed supplemented with commercially available protease. These results are similar to previous findings (Vieira *et al.*, 2013), who reported the role of protease for the enhancement of birds weight gain. Similar results were demonstrated when poultry birds were fed with diet supplemented with protease from *Bacillus subtilis* C-3102 (Hooge *et al.*, 2004).

Weight gain data indicated that protein digestibility, weight gain and feed utilization of poultry birds were improved with the increase in concentration of enzyme. The weight gain and FCR values of 1891.54±0.89g and 1.98 were recorded, respectively for 7500 IU/kg of locally produced protease as compared with results of 1874.49±0.75g and 1.995 for 5000 IU/kg of commercially available protease being imported to the country with significant *p* value ≤0.05. These results are in agreement with previous reports on the improvement of FCR due to supplementation of feed with protease (Angel *et al.*,

2011; Ajayi and Imouokhome, 2015; Aureli *et al.*, 2010; Castanon and Marquardt, 1989; Fru-Nji *et al.*, 2011; Kocher *et al.*, 2015; Ndazigaruye *et al.*, 2019; Park and Kim, 2018; Rada *et al.*, 2013; Sonu *et al.*, 2018).

## CONCLUSION

The locally characterized protease from *Geobacillus* sp. SBS-4S has strong potential for enhanced protein digestion and for the improved weight gain, feed consumption and FCR value in broiler chicks. The domestic production of this protease may result in low cost availability of the enzyme that can replace the imported counterpart being utilized currently in poultry feed industry.

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### Supplementary material

There is supplementary material associated with this article. Access the material online at: <https://dx.doi.org/10.17582/journal.pjz/20220324050332>

### Statement of conflict of interest

The authors have declared no conflict of interest.

## REFERENCES

- Aftab, S., Ahmad, S., Saeed, S. and Rasool, S.A., 2006. Screening, isolation and characterization of alkaline protease producing bacteria from soil. *Pak. J. Biol. Sci.*, **9**: 2122-2126. <https://doi.org/10.3923/pjbs.2006.2122.2126>

- Ahmad, W., Tayyab, M., Aftab, M.N., Hashmi, A.S., Ahmad, M.D., Firyal, S., Wasim, M. and Awan, A.R., 2020. Optimization of conditions for the higher level production of protease: Characterization of protease from *Geobacillus* SBS-4S. *Waste Biomass Valoriz.*, **11**: 6613–6623. <https://doi.org/10.1007/s12649-020-00935-4>
- Ajayi, H.I., and Imouokhome, J.I., 2015. Blood parameters and performance of broiler chickens fed diets containing feather meal at three crude protein levels, with or without protease supplementation. *Nigeria J. Agric. Fd. Environ.*, **11**: 146–149. <https://www.researchgate.net/publication/279257084>.
- Ali, N., Ullah, N., Qasim, M., Rahman, H., Khan, S.N., Sadiq, A. and Adnan, M., 2016. Molecular Characterization and growth optimization of halo-tolerant protease producing *Bacillus subtilis* Strain BLK-1,5 isolated from salt mines of Karak, Pakistan. *Extremophiles*, **20**: 395- 402. <https://doi.org/10.1007/s00792-016-0830-1>
- Angel, C.R., Saylor, W., Vieira, S.L. and Ward, N., 2011. Effects of a mono-component protease on performance and protein utilization in 7 to 22 days old broiler chickens. *Poult. Sci.*, **90**: 2281–2286. <https://doi.org/10.3382/ps.2011-01482>
- Applegate, T.J. and Angel, C.R., 2014. Nutrient requirements of poultry publication: History and need for an update. *J. appl. Poult. Res.*, **23**: 567–575. <https://doi.org/10.3382/japr.2014-00980>
- Aureli, R., Kluenter, A.M. and Fru, F.N., 2010. Efficacy of a phytase on growth performance and nutrient digestibility of male broiler chicks fed a diet based on maize and soybean meal. *World Poult. Sci. J.*, **66** (suppl.).
- XIII<sup>th</sup> European Poultry Conference Tours, France, 23-27. 2010. Proceedings World's Poultry Science Journal. Abstract Number, **193- 512**: 1-5.
- Bolan, N.S., Szogi, A.A., Chusavavathi, T., Seshadri, B., Rothrock, M. and Panneerselvam, P., 2010. Uses and management of poultry litter. *World Poult. Sci. J.*, **66**: 673–698. <https://doi.org/10.1017/S0043933910000656>
- Bradford, M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.*, **72**: 248–254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- Campbell, G.L. and Bedford, M.R., 1992. Enzyme application for monogastric feeds: A review. *Can. J. Anim. Sci.*, **72**: 449–466. <https://doi.org/10.4141/cjas92-058>
- Castanon, J.I.R., and Marquardt, R.R., 1989. Effect of enzyme addition, autoclave treatment and fermenting on the nutritive value of field beans (*Vicia faba* L.). *Anim. Feed Sci. Technol.*, **26**: 71–79. [https://doi.org/10.1016/0377-8401\(89\)90007-2](https://doi.org/10.1016/0377-8401(89)90007-2)
- Catara, G., Ruggiero, G., La-Cara, F., Digilio, F.A., Capasso, A. and Rossi, M., 2003. A novel extracellular subtilisin-like protease from the hyperthermophile *Aeropyrum pernix* K1: biochemical properties, cloning, and expression. *Extremophiles*, **7**: 391–399. <https://doi.org/10.1007/s00792-003-0337-4>
- Colomatto, D. and Benuchemin, K., 2009. A protease additive increases fermentation of alfalfa diets by mixed ruminal microorganisms *in vitro*. *J. Anim. Sci.*, **87**: 1097–1105. <https://doi.org/10.2527/jas.2008-1262>
- Costa, F.G.P., Clementino, R.H., Jacome, I.M.T.D., Nascimento G.A.J. and Pereira, W.E., 2006. Utilização de um complexo multienzimático em dietas de frangos de corte. *Ciencia. Anim. Bras.*, **5**: 63–67.
- Cowieson, A.J. and Roos, F.F., 2016. Toward optimal value creation through the application of exogenous mono-component protease in the diets of non-ruminants. *Anim. Feed. Sci. Technol.*, **221**: 331–340. <https://doi.org/10.1016/j.anifeedsci.2016.04.015>
- Doskovic, V., Boskovic, B.S., Pavlovski, Z., Milosevic, B., Skrbic, Z., Rakonjac, S. and Petricevic, V., 2013. Enzymes in broiler diets with special reference to protease. *Worlds Poult. Sci. J.*, **69**: 343–360. <https://doi.org/10.1017/S0043933913000342>
- Douglas, M., Parsons, C. and Bedford, M., 2000. Effects of various soyabean meal sources and Avizyme on chick growth performance and Ileal digestible energy. *J. appl. Poult. Res.*, **9**: 74–80. <https://doi.org/10.1093/japr/9.1.74>
- Freitas, D.M., Viera, S.L., Angel, C.R., Favero, A. and Maiork, A., 2011. Performance and nutrient utilization of broilers fed diets supplemented with a novel mono component protease. *J. appl. Poult. Res.*, **20**: 322–334. <https://doi.org/10.3382/japr.2010-00295>
- Fru-Nji, F., Kluenter, A.M., Fischer, M. and Pontoppidan, K., 2011. A feed serine protease improves broiler performance and increases protein and energy digestibility. *J. Poult. Sci.*, **48**: 239–246. <https://doi.org/10.2141/jpsa.011035>
- Ganapathi, R., Murugesan, Luis, F., Romero, M., and Persial, E., 2014. Effects of protease, phytase and a *Bacillus* sp. direct-fed microbial on nutrient and energy digestibility, ileal brush border digestive enzyme activity and cecal short-chain

- fatty acid concentration in broiler chickens. *PLoS One*, **9**: 101888. <https://doi.org/10.1371/journal.pone.0101888>
- Ghazi, S., Rooke, J.A., Galbraith H. and Bedford, M.R., 2002. The potential for the improvement of the nutritive value of soya-bean meal by different proteases in broiler chicks and broiler cockerels. *Br. Poult. Sci.*, **43**: 70–77. <https://doi.org/10.1080/00071660120109935>
- Hooge, D., Ishimaru H. and Sims M., 2004. Influence of dietary *Bacillus subtilis* C-3102 spores on live performance of broiler chickens in four controlled pen trials. *J. appl. Poult. Res.*, **13**: 222–228. <https://doi.org/10.1093/japr/13.2.222>
- Hussain, J., Rabbani, I., Aslam, S. and Ahmad, H.A., 2015. An overview of poultry industry in Pakistan. *Worlds Poult. Sci. J.*, **71**: 689–700. <https://doi.org/10.1017/S0043933915002366>
- Khalid, A., Tayyab, M., Shakoori, A.R., Hashmi, A.S., Yaqub, T., Awan, A.R., Wasim, M., Firyal, S., Hussain Z. and Ahmad, M., 2019. Cloning, expression and characterization of highly active recombinant thermostable cellulase from *Thermotoga naphthophila*. *Pakistan J. Zool.*, **51**: 925–934. <https://doi.org/10.17582/journal.pjz/2019.51.3.925.934>
- Kocher, A., Hower, J.M. and Moran, C.A., 2015. A dual-enzyme product containing protease in broiler diet: Efficacy and tolerance. *J. appl. Anim. Nutr.*, **3**: 1–14. <https://doi.org/10.1017/jan.2015.4>
- Leeson, S. and Summers, J.D., 2005. *Commercial poultry nutrition*. 3<sup>rd</sup> ed. University Books, Guelph. Published by Nottingham University Press Manor Farm, Church Lane, Thrumpton, Nottingham, NG11 0AX, England.
- Lemme, A., Ravindran, V., and Bryden, W.L., 2004. Ileal digestibility of amino acids in feed ingredients for broilers. *World Poult. Sci. J.*, **60**: 423–437. <https://doi.org/10.1079/WPS200426>
- Mahagna, M., Nir, I., Lerbier, M. and Nitsan, Z., 1995. Effect of age and exogenous amylase and protease on development of the digestive tract, pancreatic enzyme activities and digestibility of nutrients in young meat-type chicks. *Reprod. Nutr. Dev.*, **35**: 201–212.
- Mahmood, T., Mirza, M.A., Nawaz, H. and Shahid, M., 2017. Effect of different exogenous proteases on growth performance, nutrient digestibility, and carcass response in broiler chickens fed poultry by-product meal-based diets. *Livest. Sci.*, **200**: 71–75. <https://doi.org/10.1016/j.livsci.2017.04.009>
- Mukhtar, H. and Haq, I.U., 2012. Purification and characterization of alkaline protease produced by mutant strain of *Bacillus subtilis*. *Pak. J. Bot.*, **44**: 1697–1704.
- National Research Council., 2014. *Nutrient requirements of poultry*. Ninth revised edition, 1994. The National Academies Press, Washington DC.
- Ndazigaruye, G., Kim, D.H., Kang, C.W., Kang, K.R., Joo, Y.J., Leeand, S.R. and Lee, K.W., 2019. Effects of low-protein diets and exogenous protease on growth performance, carcass traits, intestinal morphology, cecal volatile fatty acids serum parameters broilers animal. *Animals*, **9**: 226. <https://doi.org/10.3390/ani9050226>
- Okafor, U.O.G. and Anosike, E.E.M., 2012. Screening and optimal protease production by *Bacillus* sp. Sw-2 using low cost substrate medium. *Res. J. Microbiol.*, **7**: 327–336. <https://doi.org/10.3923/jm.2012.327.336>
- Oxenboll, K.M., Pontoppidan K. and Fru-Nji F., 2011. Use of a protease in poultry feed offers promising environmental benefits. *Int. J. Poult. Sci.*, **10**: 842–848. <https://doi.org/10.3923/ijps.2011.842.848>
- Pant, G., Prakash, A., Pavani, J.V.P., Bera, S., Deviram, G.V.N.S., Kumar, A., Panchpuri, M. and Prasuna, R.G., 2015. Production, optimization and partial purification of protease from *Bacillus subtilis*. *J. Taibah Univ. Sci.*, **9**: 50–55. <https://doi.org/10.1016/j.jtusci.2014.04.010>
- Park, J.H. and Kim, I.H., 2018. Effects of a protease and essential oils on growth performance, blood cell profiles, nutrient retention, ileal microbiota, excreta gas emission, and breast meat quality in broiler chicks. *Poult. Sci.*, **97**: 2854–2860. <https://doi.org/10.3382/ps/pey151>
- Puente, X.S. and Lopez, O.C., 2004. A genomic analysis of rat proteases and protease inhibitors. *Genome Biol.*, **14**: 609–622. <https://doi.org/10.1101/gr.1946304>
- Rada, V., Lichovnikova, M., Foltynl, M. and Safarik, I., 2016. The effect of exogenous protease in broiler diets on the apparent ileal digestibility of amino acids and on protease activity in jejunum. *Acta. Univ. Agric. Silv. Mend. Brun.*, **64**: 1645–1652. <https://doi.org/10.11118/actaun201664051645>
- Rada, V., Foltyn, M., Lichovniková, M. and Musilova, A., 2013. Effects of protease supplementation of low protein broiler diets on growth parameters and carcass characteristic. *Mendel Net*, pp. 268–272. [https://mnet.mendelu.cz/mendelnet2013/mnet\\_2013\\_full.pdf](https://mnet.mendelu.cz/mendelnet2013/mnet_2013_full.pdf)
- Rahman A., Saima, Pasha, N.T., Younas, M., Yassar, A. and Ditta, Y.A., 2014. Effect of multi-enzymes

- supplementation on growth performance of broiler. *Pakistan J. Zool.*, **46**: 417- 422.
- Sabir, F., Tayyab, M., Awan, A.R., Muneer, B., Hashmi, A.S., Wasim, M. and Firyal, S., 2018. Biological evaluation of locally produced recombinant phytase in broiler chicks. *J. Anim. Pl. Sci.*, **28**: 946–950. <http://www.thejaps.org.pk>
- Seskeviciene, J., Jeroch, H., Danicke, S., Gruzauskas, R., Volker, L. and Broz, J., 1999. Feeding value of wheat and wheat-based diets with different content of soluble pentosans when fed to broiler chickens without or with enzyme supplementation. *Arch. Geflugelk.*, **63**: 129–132.
- Sharma, K.M., Kumar, R., Panwar, S. and Kumar, A., 2017. Microbial alkaline proteases: Optimization of production parameters and their properties. *J. Genet. Eng. Biotechnol.*, **15**: 115–126. <https://doi.org/10.1016/j.jgeb.2017.02.001>
- Sonu, S.Z.S., Ahlawat, P.K. and Dalal, R., 2018. Effect of protease enzyme on the growth performance and carcass traits of broilers fed with DDGS supplemented diet. *Int. J. Curr. Microbiol. appl. Sci.*, **7**: 2713–2719. <https://doi.org/10.20546/ijemas.2018.705.314>
- Steel, R.G.D., and Torrie, J.H. 1997. *Principles and procedures of statistics: A biometrical approach*. (3<sup>rd</sup> Ed.) McGraw Hill Book Co. Inc, New York, USA, ISBN-13, 978-0070610286, ISBN-10: 0070610282.
- Thirumalaisamy, G., Muralidharan, J., Senthilkumar, S., Sayee, R.H. and Priyadharsini, M., 2016. Cost-effective feeding of poultry. *Int. J. Sci. environ. Technol.*, **5**: 3997–4005.
- Toghyani, S., Wu, B., Maldonado, P.R.A., Iji, P.A. and Swick, R.A., 2017. Performance, nutrient utilization, and energy partitioning in broiler chickens offered high canola meal diets supplemented with multicomponent carbohydrase and mono-component protease. *Poult. Sci.*, **96**: 3960–3972. <https://doi.org/10.3382/ps/pex212>
- Vieira, S.L., Angel, C.R., Miranda, D.J.A., Favero, A., Cruzand, R.F.A. and Sorbara, J.O.B., 2013. Effects of a monocomponent protease on performance and protein utilization in 1 to 26 days of age turkey poult. *J. appl. Poult. Res.*, **22**: 680–688. <https://doi.org/10.3382/japr.2012-00558>
- Wang, X. and Parsons, C.M., 1998. Effect of raw material source, processing system, and processing temperatures on amino acid digestibility of meat and bone meals. *Poult. Sci.*, **77**: 834–841. <https://doi.org/10.1093/ps/77.6.834>