

et al., 2011). Phytochemistry showed the presence of different phytochemicals in the methanol extract of seeds of this plant e.g., alkaloids, flavonoids, saponins, glycosides and tannins. The analytical study revealed the occurrence of calcium, sodium, magnesium and zinc. Proximate analytical study has shown high concentration of fat, carbohydrates and protein (Lucky *et al.*, 2012).

No previous investigation has been carried out to assess anti-hyperlipidemic effect of the seeds of this plant in Triton X-100 induced hyperlipidaemia rat model. So, the objective of present investigation was to evaluate antihyperlipidemic effects of the seeds of this plant in hyperlipidemic albino rats.

MATERIALS AND METHODS

Collection and authentication of plant

Seeds of *Citrullus lanatus* were purchased from Gulberg, Lahore. Identification and authentication of the plant was done by Prof. Dr. Zaheer-ud-Din Khan, Botany Department, G.C University, Lahore. A voucher specimen with number (GC. Herb. Bot. 3383) was deposited in the herbarium of University.

Preparation of plant see extract

Seeds of the plant were washed, air dried, chopped and powdered with herbal grinder. The plant material was placed in well closed container and stored in the refrigerator at 4°C before use. 500 g of the powdered seeds was added to 2100 ml of methanol (95%) and the extraction was processed in hot Soxhlet apparatus at 70°C for 72 h. The extract was dried by a rotary evaporator (Heidolph, Laborata 4002, Germany) under reduced pressure at 60°C. Methanol extract was stored in sample bottles at 0 - 4 °C for further studies (Sathya and Shoba, 2014).

Animals used for the study

All the animal experimental methods were approved by the Animal Ethical Committee (AEC) of The University of Lahore (Approval number: IREC-2019-79, 28/03/2019). The animals were purchased from Post Graduate Medical Institute and were placed in the animal house, Faculty of Pharmacy, The University of Lahore. The rats were placed in polyacrylic cages and were maintained at the temperature 25±1°C under the standard laboratory conditions for 20 days. The rats were permitted to free access of water and the standard dry pellet diet *ad libitum* and acclimatized at the laboratory conditions before the start of experiment.

Acute toxicity studies

Toxicological studies of methanol extract (5000 mg/kg) of the plant were done as per OECD (Organization for

Economic Co-operation Development) guidelines No. 425 (OECD, 2008).

Induction of hyperlipidaemia

The standard pellet diet was mixed with egg yolks and butter and given to the groups (B, C and D) daily for the development of hyperlipidaemia. Group A was kept on normal standard diet. The solution of Triton X-100 was freshly prepared in the normal saline solution and was given through i.p. injection at the dose 100 mg/kg to groups B, C and D after fasting for 18 h.

Study design

The study was done using 20 male Wistar albino rats weighing 180-250 g. After 72 h of i.p. injection of Triton X-100 to all groups except group A. The rats were randomly distributed into four groups (n=5) labelled as Group A, B, C and D. Group A was kept as normal control while group B was kept without treatment, group C was treated with the standard simvastatin (10 mg/kg O.D) and group D was treated with the methanol extract of seeds of the plant (800 mg/kg) orally for 15 days. The amount of seed extract for every rat was calculated based on body weight. The suspension of the plant extract was administered by oral route to every rat by the use of a stomach tube with disposable syringe. Detail of the groups was as follow: Group A, normal control (NC) administered with dry pellet diet, normal saline and fresh water by oral route during treatment. Group B, hyperlipidaemia control (HC) in which Hyperlipidaemia was induced by high cholesterol diet with 100 mg/kg intraperitoneal injection of Triton X-100 (Sudha *et al.*, 2011). This groups was administered with the normal saline by oral route during the treatment. Group C, hyperlipidaemia standard (H-S): This group was administered with Simvastatin (10 mg/kg) O.D. after 72 h of induction of hyperlipidaemia for 7 days (Shinde *et al.*, 2013) and Group D, hyperlipidaemia *C. lanatus* (H-CL): This group was administered with methanol extract of seeds of the plant (800 mg/kg) orally for 7 days (Lucky *et al.*, 2012).

Finally, the samples of blood were taken by the cardiac puncture and were taken into the gel clot activator tubes to separate serum. The liver and heart tissues were placed into 100 mL beakers containing formalin (10%) and stored at temperature of -20°C for further histopathological studies.

Biochemical analysis

TC, TG, HDL-C, ALT, AST levels were determined using respective assay kits of Chemistry Analyzer Cobas C111 (Roche Diagnostics).

Chemistry analyser Cobas C 111 (Assay procedure)

The sample was pipetted into a prepared Cobas

sample tube (RD standard false bottom tube) and placed in Cobas C111 analyzer. The desired test was selected in the user menu and the measurement was started. The measurement was carried out automatically. Cholesterol level was measured using CHOD-PAP method. Applying Friedewald's formula LDL and VLDL levels were calculated as follows:

LDL= TC-(HDL-VLDL) and VLDL= TG/5 respectively (Friedewald *et al.*, 1972) while Non-HDL was determined using the following formula (Non HDL-C = TC-HDL) (Ridker *et al.*, 2005).

Histopathological studies

Tissue samples were collected for histopathological examination. The tissues were passed through various standard techniques like fixation, washing, dehydration, clearing, placed, sectioning and staining (Carleton *et al.*, 1980; Winsor, 1998). Fixation of tissues was done by chemical fixation procedure using aqueous solution of formaldehyde (10%) at normal pH. Dehydration and clearing process was done using xylene. The tissues were placed in paraffin. Using microtome tissue sections of 5-8 μ m thickness were cut and floated on water.

At the end, the slides with various sections were placed in the incubator at temperature 37°C for about 3 h for further adhesion. The histological sections were stained with eosin and haematoxylin (Winsor, 1998) and then examined under light microscope (Leitz, Wetzle, Germany).

Statistical analysis

The results were presented as mean \pm SEM. The statistical analysis of data was done by one way analysis of variance (ANOVA) which is followed by Turkey's Multiple Comparison as *post hoc* test using Graph Pad

Prism version 5. The value at $P < 0.05$ was considered significant statistically.

RESULTS

Toxicological studies

Intraperitoneal (i.p.) administration of methanol extract (5000 mg/kg b.w.) of the plant seeds caused no death in 2 stages of the test. Thus i.p. LD₅₀ of methanol extract was estimated to be equal to or greater than 5000 mg/kg b. w. (Omoboyowa and Ajayi, 2016).

Effects of *C. lanatus* on hyperlipidaemia

The effect of plant seed extract, administered for 15 days, on serum cholesterol, triglycerides, LDL-C, HDL-C and VLDL-C (mg/dl \pm SEM) in hyperlipidaemia induced rats is shown in Table I. The levels of total cholesterol, total glycerides, LDL-C, non HDL-C, VLDL-C and ALT in serum of group D treated with methanol extract were found to be significantly ($P < 0.05$) decreased as compared to group A. The serum levels of HDL and AST in group D not significantly ($P > 0.05$) different from those of group A.

Histopathological examination of liver

Normal configuration of liver in control rats is shown in Figure 1A. Severe congestion of blood vessels, necrotic foci along with fatty changes of hepatocytes resulted in shrunken sinusoids in Triton X-100 treated rats (Fig. 1B). Hepatocytes of zone 1 are normal but of zone 3 had undergone mild fatty changes due to the accumulation of fat droplets in rats treated with simvastatin (Fig. 1C). The seed extract administered rats depicted mild congestion, accumulation of some inflammatory cells in portal areas but no hyperlipidaemia signs are seen in Figure 1D.

Table I. Effect of *C. lanatus* seeds on serum cholesterol, triglyceride, HDL-C, Non-HDL-C, LDL-C, VLDL-C, ALT and AST levels in triton X-100 induced rats after 15 days of methanol extract administration.

Parameter	Group NC	Group HC	Group H-S	Group H-CL
TC (mg/dl)	55.6 \pm 4.22	100.6 \pm 7.83 ^{c***}	55 \pm 2.12 ^{b**}	53.6 \pm 6.89 ^{d****}
TG (mg/dl)	79.2 \pm 12.78	255.2 \pm 58.69 ^{b**}	81.0 \pm 7.19 ^{b**}	72.0 \pm 4.25 ^{b**}
HDL-C (mg/dl)	41.4 \pm 3.10	22.96 \pm 5.97 ^{a*}	44.2 \pm 3.51 ^{a*}	27.8 \pm 3.83
Non-HDL-C (mg/dl)	14.2 \pm 1.20	77.64 \pm 11.50 ^{d****}	10.8 \pm 1.93 ^{d****}	13.4 \pm 3.07 ^{d****}
LDL-C (mg/dl)	30.04 \pm 3.13	103.72 \pm 21.66 ^{c***}	27.0 \pm 3.13 ^{b**}	40.2 \pm 4.12 ^{c***}
VLDL-C (mg/dl)	15.84 \pm 2.55	51.04 \pm 11.73 ^{b**}	16.2 \pm 1.43 ^{b**}	14.4 \pm 0.85 ^{b**}
ALT (IU/dl)	43.0 \pm 3.36	70.6 \pm 6.55 ^{b**}	38.6 \pm 3.48 ^{b**}	40.4 \pm 5.33 ^{b**}
AST (IU/dl)	122.8 \pm 13.05	183.0 \pm 14.60	138.6 \pm 13.21	129.4 \pm 9.79 ^{a*}

The values are expressed as mean \pm SEM (n=3). Both significance ($P < 0.05$) and non-significance ($P > 0.05$) results were obtained when compared to normal control. NC, normal control; HC, hyperlipidaemia control; H-S, hyperlipidaemia standard; H-CL, hyperlipidaemia *C. lanatus*; TC, total cholesterol; TG, triglyceride; HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol; VLDL-C, very low density lipoprotein cholesterol; ALT, alanine aminotransferase; AST, aspartate transaminase.

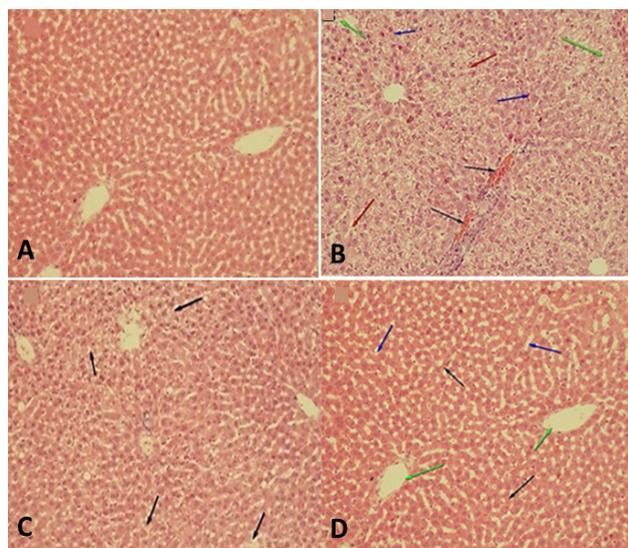


Fig. 1. Histopathological changes in liver of animals induced by Triton X-100. Haematoxylin and Eosin (*H and E*) $\times 100$. A: normal configuration of liver. B: untreated Triton X-100 induced hyperlipidaemia rats having severe congestion of hepatic blood vessels (arrows). C: Simvastatin treated rats with mild fatty changes in hepatocytes. D: *C. lanatus* seeds treated rats having mild congestion of hepatic blood vessels.

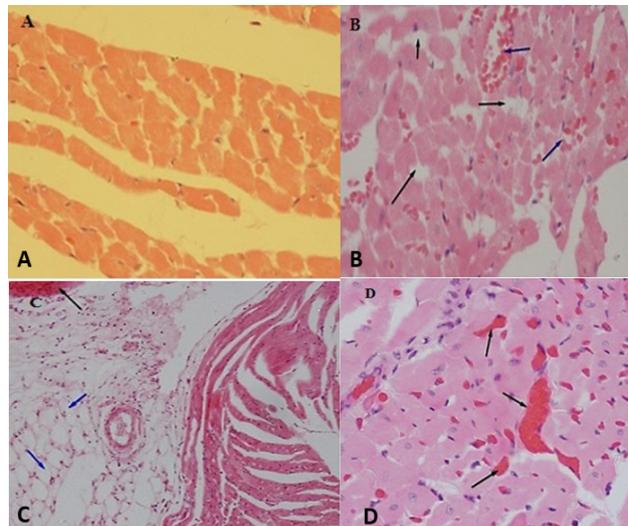


Fig. 2. Effect of methanol extract of the plant seeds on histopathological changes in transverse section of hearts of rats induced by Triton X-100. A: normal configuration of heart muscle and coronary vessels. B: accumulation of lipid particles (black arrow) and severe congestion of blood coronary vessels (blue arrow). C: cardiac muscle fibres had vacuolation of lipids droplets (blue arrow) and mild congestion of coronary blood vessels (black arrow). D: mild congestion of blood coronary vessels with no hyperlipidaemia condition.

Histopathological examination of the heart

Severe hyperlipidaemia condition e.g., accumulation of lipid particles and severe congestion of blood vessels are seen in the hearts of Group B rats (Fig. 2B) as compared to group A control rats (Fig. 2A). Group C treated with simvastatin shows severe congestion of blood vessels with mild hyperlipidaemia condition. Histological structure of myocardium shows lipids droplets in cardiac muscle fibres (Fig. 2C). The group D treated with the plant had mild congestion of blood vessels and no hyperlipidaemia condition. No evidence of the accumulation of lipid droplet is found (Fig. 2D).

DISCUSSION

In this study, seeds extract of *C. lanatus* caused a significant improvement in lipid profile, ALT and AST levels as well as histological features of liver and heart in hyperlipidaemia induced rats. Hyperlipidaemia is the most vital risk factor of atherosclerosis, manifested by elevated plasma levels of lipid profile (TC, TG, non-HDL, LDL and VLDL) accompanied with a decline in levels of HDL (Koba and Hirano, 2011). Lipid profile was kept to be high in hyperlipidaemia untreated group while improvements were seen in the simvastatin and the seed extract treated groups. The abnormal lipid profile is the indicator of hyperlipidaemia where liver and small intestine could be the hypothesized as sites of the effect of plant extract on lipid profile. The improvement of lipids would be compensated by upsurge excretion of total cholesterol. The intestinal antihyperlipidemic effect might be facilitated by obstructing absorption of cholesterol in intestine and eliminating reverse movement of cholesterol, as explained with the standard therapeutic agents like Ezetimibe (Sudhop *et al.*, 2002; Catapano *et al.*, 2014) and bile acid sequestrant, cholestyramine (Davidson *et al.*, 2013; Terunuma *et al.*, 2013).

The administration of methanol extract of the plant led to low concentration of TC which could be due to lower levels of LDL. It might be due to the anti-oxidant effects of the plant (Singh *et al.*, 2011). Our findings are related with the previous studies showing that the fruit juice of this plant decreased LDL and TC levels with significant ($P < 0.05$) results in Sprague-Dawley rats linked with anti-oxidant effect by the changes in gene expression by lipid metabolism (Hong *et al.*, 2015). The plant extract increased HDL-C levels which indicates its anti-atherogenic activity in hyperlipidaemia condition. Our results are concordant with the earlier research in which the anti-atherosclerotic activity of extract prepared from the plant on LDL-deficient mice model was evaluated in which decreased levels of

cholesterol by the plant exhibited its antihyperlipidaemia activity at significant level (Poduri *et al.*, 2013). Initially by treating with the statins was a cause of an increased level of AST and ALT. Treatment with statins was a cause of obesity that is related with fatty liver that caused the elevated levels of ALT (Athyros *et al.*, 2010). The extract of plant led to decreased level of ALT and AST in comparison to the standard simvastatin showing its hepatoprotective activity.

CONCLUSIONS

The plant *C. lanatus* has shown to exert significant antihyperlipidaemia effects in hyperlipidaemia rats induced by Triton X-100. This shows the presence of active phytochemicals which are responsible for hyperlipidaemia effect. This plant besides having antihyperlipidaemic effect has fewer side effects of abnormal liver enzymes and weight gain. However, further studies are recommended to investigate the exact mechanism(s) by which the plant shows antihyperlipidaemia effect. There is need for establishing the efficacy and safety for further clinical use. Also, there is the requirement for activity-oriented fractionation to isolate and identify the active constituent(s) of this well-known medicinal plant.

ACKNOWLEDGEMENT

Authors are grateful to Professor Dr. Muhammad Shahzad, Department of Pharmacology, UHS, Lahore for helpful guidance and support.

Statement of conflicts of interest

The authors have declared no conflict of interest.

REFERENCES

- Athyros, V.G., Tziomalos, K., Gossios, T.D., Griva, T., Anagnostis, P., Kargiotis, K. and GREACE Study Collaborative Group. 2010. Safety and efficacy of long-term statin treatment for cardiovascular events in patients with coronary heart disease and abnormal liver tests in the Greek Atorvastatin and Coronary Heart Disease Evaluation (GREACE) Study: a post-hoc analysis. *Lancet*, **376**: 1916-1922. [https://doi.org/10.1016/S0140-6736\(10\)61272-X](https://doi.org/10.1016/S0140-6736(10)61272-X)
- Calixto, J.B., 2005. Twenty-five years of research on medicinal plants in Latin America: A personal view. *J. Ethnopharmacol.*, **100**: 131-134. <https://doi.org/10.1016/j.jep.2005.06.004>
- Carleton, H.M., Drury, R.A.B. and Wallington, E.A., 1980. *Carleton's histological technique*. Oxford University Press. <https://catalogue.nla.gov.au/Record/2864951>
- Catapano, A.L., Farnier, M., Foody, J.M., Toth, P.P., Tomassini, J.E., Brudi, P. and Tershakovec, A.M., 2014. Combination therapy in dyslipidemia: Where are we now? *Atherosclerosis*, **237**: 319-335. <https://www.sciencedirect.com/science/article/pii/S0021915014014282>, <https://doi.org/10.1016/j.atherosclerosis.2014.09.026>
- Davidson, M.H., Voogt, J., Luchoomun, J., Decaris, J., Killion, S., Boban, D. and Neese, R., 2013. Inhibition of intestinal cholesterol absorption with ezetimibe increases components of reverse cholesterol transport in humans. *Atherosclerosis*, **230**: 322-329. <https://doi.org/10.1016/j.atherosclerosis.2013.08.006>
- Friedewald, W.T., Levy, R.I. and Fredrickson, D.S., 1972. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin. Chem.*, **18**: 499-502. <https://doi.org/10.1093/clinchem/18.6.499>
- Gramza, A. and Korczak, J., 2005. Tea constituents (*Camellia sinensis* L.) as antioxidants in lipid systems. *Trends Fd. Sci. Tech.*, **16**: 351-358. <https://doi.org/10.1016/j.tifs.2005.02.004>
- Hassan, L.E.A., Koko, W.S., Osman, E.E., Dahab, M.M. and Sirat, H.M., 2011. *In vitro* anti-giardial activity of *Citrullus lanatus* var. *citroides* extracts and cucurbitacins isolated compounds. *J. med. Pl. Res.*, **5**: 3338-3346. <http://www.academicjournals.org/JMPR>
- Hong, M.Y., Hartig, N., Kaufman, K., Hooshmand, S., Figueroa, A. and Kern, M., 2015. Watermelon consumption improves inflammation and antioxidant capacity in rats fed an atherogenic diet. *Nutri. Res.*, **35**: 251-258. <https://doi.org/10.1016/j.nutres.2014.12.005>
- Koba, S. and Hirano, T., 2011. Dyslipidemia and atherosclerosis. *Nihon Rinsho. Jpn. J. clin. Med.*, **69**: 138-143. <https://europepmc.org/article/med/21226274>
- Kumar, A., Mazumder, A. and Saravanan, V., 2008. Antihyperlipidemic activity of *Camellia sinensis* leaves in Triton WR-1339 induced albino rats. *Pharmacogn. Mag.*, **4**: 60. https://www.researchgate.net/publication/333603466_Pharmacognosy_Magazine
- Li, H., Horke, S. and Förstermann, U., 2014. Vascular oxidative stress, nitric oxide and atherosclerosis. *Atherosclerosis*, **237**: 208-219. <https://doi.org/10.1016/j.atherosclerosis.2014.09.026>

- [org/10.1016/j.atherosclerosis.2014.09.001](https://doi.org/10.1016/j.atherosclerosis.2014.09.001)
- Lucky, O.O., John, U.O., Kate, I.E., Peter, O.O. and Jude, O.E., 2012. Quantitative determination, metal analysis and antiulcer evaluation of methanol seeds extract of *Citrullus lanatus* Thunb (*Cucurbitaceae*) in rats. *Asian Pac. J. trop. Biomed.*, **2**: S1261-S1265. [https://doi.org/10.1016/S2221-1691\(12\)60396-7](https://doi.org/10.1016/S2221-1691(12)60396-7)
- Mendis, S., Puska, P., Norrving, B. and World Health Organization, 2011. *Global atlas on cardiovascular disease prevention and control*, WHO, Geneva.
- OECD, 2008. *OECD guidelines for the testing of chemicals, Test 425*. OECD Publishing, Paris.
- Omoboyowa, D.A. and Ajayi, A.A., 2016. Effect of methanolic extract of *Citrullus lanatus* seeds on lipid profile and oxidative stress in acetaminophen intoxicated rats. *Adv. Biomed. Pharm.*, **3**: 87-93. <https://doi.org/10.19046/abp.v03i02.02>
- Poduri, A., Rateri, D.L., Saha, S.K., Saha, S. and Daugherty, A., 2013. *Citrullus lanatus* 'sentinel' (watermelon) extract reduces atherosclerosis in LDL receptor-deficient mice. *J. Nutri. Biochem.*, **24**: 882-886. <https://doi.org/10.1016/j.jnutbio.2012.05.011>
- Ridker, P.M., Rifai, N., Cook, N.R., Bradwin, G. and Buring, J.E., 2005. Non-HDL cholesterol, apolipoproteins AI and B100, standard lipid measures, lipid ratios and CRP as risk factors for cardiovascular disease in women. *J. Am. med. Assoc.*, **294**: 326-333. <https://jamanetwork.com/>, <https://doi.org/10.1001/jama.294.3.326>
- Saravanan, R., Rajendra Prasad, N. and Pugalendi, K.V., 2003. Effect of piper betle leaf extract on alcoholic toxicity in the rat brain. *J. med. Fd.*, **6**: 261-265. <https://doi.org/10.1089/10966200360716689>
- Sathya, J. and Shoba, F.G., 2014. Assessment of antimicrobial efficacy of *Citrullus lanatus* methanolic seed extract. *J. chem. pharm. Res.*, **6**: 640-643. <https://pdfs.semanticscholar.org/d781/b57cd2744b301e7bfc6d526cadc2cc5c008e.pdf>
- Sharma, S., Sarvesh, P., Dwivedi, J. and Amita, T., 2011. First report on laxative activity of *Citrullus lanatus*. *Pharmacologyonline*, **2**: 790-797. <http://www.phytopharmajournal.com/V2issue606.pdf>
- Shinde, S., Chivate, N., Kulkarni, P. and Naikwade, N., 2013. Hypolipidemic activity of *Psidium guajava* Linn leaves extracts in hyperlipidemic rats. *Int. J. Pharm. Pharm. Sci.*, **5**: 70-72. <https://pdfs.semanticscholar.org/7b1a/a5d4c218bdd7fa79cf6c068fa909a1dbbb5c.pdf>
- Singh Gill, N., Sood, S., Muthuraman, A., Bali, M. and Dev Sharma, P., 2011. Evaluation of antioxidant and anti-ulcerative potential of *Citrullus lanatus* seed extract in rats. *Lat. Am. J. Pharm.*, pp. 30. <http://sedici.unlp.edu.ar/handle/10915/8158>
- Sudha, S.S., Karthic, R. and Naveen, J.R., 2011. Anti hyperlipidemic activity of *Spirulina platensis* in Triton x-100 induced hyperlipidemic rats. *Hygeia J. D. Med.*, **3**: 32-7. <http://www.hygeiajournal.com/downloads/2337269785.sudha.pdf>
- Sudhop, T., Lütjohann, D., Kodak, A., Igel, M., Tribble, D.L., Shah, S. and Von Bergmann, K., 2002. Inhibition of intestinal cholesterol absorption by ezetimibe in humans. *Circulation*, **106**: 1943-1948. <https://www.ahajournals.org/doi/abs/10.1161/01.cir.0000034044.95911.dc>, <https://doi.org/10.1161/01.CIR.0000034044.95911.DC>
- Terunuma, S., Kumata, N. and Osada, K., 2013. Ezetimibe impairs uptake of dietary cholesterol oxidation products and reduces alterations in hepatic cholesterol metabolism and antioxidant function in rats. *Lipids*, **48**: 587-595. <https://link.springer.com/article/10.1007/s11745-013-3790-6>, <https://doi.org/10.1007/s11745-013-3790-6>
- Valiathan, M.S., 1998. Healing plants. *Curr. Sci.*, **75**: 1122-1127. <https://scholar.google.com.pk/citations?user=tyV77i8AAAA&Jandhl=en&doi=sra>
- Winsor, L., 1998. Collection, handling, fixation, histological and storage procedures for taxonomic studies of terrestrial flatworms (*Tricladida: Terricola*). *Pedobiologia*, **42**: 405-411. <https://researchonline.jcu.edu.au/2715s>