



# Expression of the ZFX Gene in Mouse Kidney During Postnatal Development

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## ABSTRACT

The relationship of cell proliferation and apoptosis is important in normal kidney development. The ZFX gene is important for regulation of growth, proliferation and differentiation in tissue cells while the Bcl-2 and BAX genes regulate apoptosis. This experiment adopted the real-time fluorescence quantitative technique to test ZFX, Bcl-2, and BAX expression in mice kidney tissue on 1 d, 7 d, 14 d, 21 d, 42 d, and 90 d after birth. The results showed that with the maturation of mouse kidney, the cell proliferation activity decreased, and the expression of ZFX, Bcl-2, and BAX were down regulated. The ZFX gene may participate in the developmental process of the kidney through the balance of cell proliferation and apoptosis regulation.

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## Authors' Contribution

BJ designed the project. XW performed the research and wrote the manuscript. JX conducted the experiments, collected and analysed the data. YZ and CL contributed to RT-PCR.

## Key words

ZFX, Bcl-2, BAX, RT-qPCR, Kidney development, Cell proliferation and apoptosis, Mice.

## INTRODUCTION

Zinc finger protein X-linked (ZFX) is a member of the zinc finger protein super family and contains 13 C2H2-type zinc finger structures. The ZFX gene is on the x chromosome in mammals, and its expression is highly conservative. The structure of ZFX contains an acidic transcriptional activation domain (AD), a nuclear localization sequence (NLS), and a DNA binding domain (DBD). Early research suggested the ZFX gene was associated with sex determination of mammals (Schneider-Gadicke *et al.*, 1989). In recent years, research documented that ZFX plays an important role in self-renewal and anti-apoptotic mechanisms of embryonic stem cells, adult hematopoietic stem cells, lymphoid cells and several other types of stem cells (Galan-Cardad *et al.*, 2007; Arenzana *et al.*, 2009). Considering the characteristics of the tumor and stem cell, oncology studies found ZFX was abnormally expressed in laryngeal carcinoma's, glioma's, gastric cancer, prostate cancer, breast cancer and other common human malignancies, and it may also be involved in the regulation of tumor cell proliferation and anti-apoptotic processes (Jiang *et al.*, 2012; Nikpour *et al.*, 2012; Zhou *et al.*, 2011). Since the zinc finger protein is a transcription factor which has a

finger-like domain, it can combine with the promoter region thereby regulating gene expression, and also plays an important role in the growth, proliferation and differentiation of tissue cells (Chandrasekharan *et al.*, 2009; Schnidar *et al.*, 2009). There are two categories of apoptosis regulatory genes which contain apoptosis inhibition genes and apoptosis promotion genes. Bcl-2 is the most important anti-apoptotic gene while BAX is a major pro-apoptotic gene (Kallio *et al.*, 2004; Yamamoto *et al.*, 2004).

The development of the mammalian kidney includes stages of pronephros, mesonephros and metanephros, and the three stages are consecutive in the timing and location (Lipschutz, 1998; Davies, 2002). The appearance of mouse pronephros is at embryonic day 8 (E8), mesonephros is initiated at E9.5, and metanephros begins at E11 with the appearance of a small epithelial buds or diverticulum from the lower end of the mesonephric duct near its entry into the cloaca; the completion of the number of glomeruli in mice is on day 7 postpartum, with completion of kidney medulla on day 21, and by day 42 the structure of kidney is the same as the adult (Nyengaard, 1993).

Real-time, reverse transcription quantitative PCR (RT-qPCR) is a highly sensitive and reproducible technology for the analysis of gene expression patterns. In this study, by quantitative real-time technique ZFX, Bcl-2 and BAX expression at different stages of kidney development in mice after birth were evaluated, to determine the expression of ZFX during kidney development.

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**Table I.- Conditions of PCR and oligonucleotide primer pairs.**

Target gene	GenBank No.	Sequence of primer (5'-3')	Annealing temp.	Products
ZFX	NM_011768.2	F: CAGATCCGTTTGTACTAAGTCGC R: ACTGATACACTTTCCGGCCACT	61°C	147 bp
Bcl-2	NM_011768.2	F: GTCGCTACCGTCGTGACTTC R: CAGACATGCACCTACCCAGC	61°C	284 bp
BAX	NM_007527.3	F: CGGCGAATTGGAGATGAACTG R: GCAAAGTAGAAGAGGGCAACC	61°C	161 bp
GAPDH	NM_008084.2	F: ACCCAGAAGACTGTGGATGG R: CACATTGGGGGTAGGAACAC	61°C	171 bp

## MATERIALS AND METHODS

### Animals

Clean adult mice were obtained from the Experimental Animal Center, Shihezi University. Mice were housed in clean, controlled environment, with food and water available *ad libitum*. Room conditions included maintenance of temperature at 24°C, humidity at 50%, with lighting from 8:00-20:00. After adaptation for 7 days, mice were paired and bred. The next morning females with vaginal plugs were separated for production of experimental litters. Kidneys of progeny aged 1, 7, 14, 21, 42 and 90 days were removed immediately after the animals were killed by cervical dislocation. The tissues stored in liquid nitrogen for reverse transcription-quantitative real-time polymerase chain reaction (RT-qPCR).

### Primary reagents

Trizol was purchased from Invitrogen Life Technologies, Carlsbad, CA, USA, reverse transcription kits were purchased from Takara Co., Dalian, China, diethyl pyrocarbonat was purchased from Sigma (Santa Clara, CA, USA), agarose was purchased from GeneTech Co., Shanghai, China, DNA Marker was purchased from Tiangen Co., Beijing, China, SYBR® Premix Ex Taq™ (TaKaRa), 96-well plates and cover films were purchased from Applied Biosystems Inc., CA, USA and SYBR Green SuperMix was acquired from Qiagen, Valencia, CA, USA.

### RNA extraction and reverse transcription

Total RNA was extracted from kidneys cryopreserved with liquid nitrogen using Trizol following the manufacturer's instructions. Reverse transcription of RNA was carried out using the gDNA Eraser PrimerScript RT reagent kit according to the manufacturer's instructions.

### Primer design

According to the principles of primer design (Rodríguez *et al.*, 2015; Livak *et al.*, 2001), Primer Premier 5.0 software was applied to design the primer sequences

of ZFX, BAX, Bcl-2, and GAPDH, The primer sequences was synthesized by BGI (Table I).

### Conventional PCR assays

To detect primer specificity, and the quality of the template, PCR was conducted in advance. The amplification system was as follows: cDNA 1 µl, PCR reaction mixture (Mix) 10µl, upstream and downstream of each primer, 0.5 µl, ddH<sub>2</sub>O 8µl. Reaction conditions were: 94°C denaturation for 5 min; 94°C denaturation 30 s, 61°C annealing 30 s, 72°C extension 50 s, 30 cycles; 72°C extension 10 min. Finally, 10 µl of PCR products were run on 2.0% agarose gel, and then analyzed by a gel imager analysis system (need to specify the system).

### RT-qPCR

Reverse transcription-quantitative real-time polymerase chain reaction (RT-qPCR) was conducted using a LightCycler 2.0 instrument (Roche, Basel, Switzerland), with SYBR Green SuperMix, following the manufacturer's instructions. Each sample was tested three times to insure accuracy. The amplification system was as follows: cDNA 2 µl, SYBR® Premix Ex Taq™ (2×) mixture (Mix) 10 µl, upstream and downstream of each primer, 0.4 µl, sterile distilled water to 20 µl. Reaction conditions were: initial denaturation step (30 s at 95°C), 45 cycles of 95°C for 10 s, 61°C for 30 s, and 72°C for 30 s.

### Statistical analysis

Numerical analysis was done using the 2<sup>-ΔΔCT</sup> method using the Gapdh gene as an internal standard. Data were analyzed using SPSS 17.0 (city, state, USA). Tests of hypotheses were done using t-tests with P<0.05 considered statistically significant.

## RESULTS

### Amplified products of ZFX, Bcl2, BAX and GAPDH

The cDNA were amplified with ZFX, Bcl<sub>2</sub>, BAX and GAPDH primers (Table I) to obtain 147 bp, 284 bp, 161 bp

and 171 bp products (Figs. 1, 2), respectively. According to the results of the gel imaging system, primers had excellent specificity and the qualities of the templates were reliable.



Fig. 1. PCR amplification of ZFX and Bcl-2.



Fig. 2. PCR amplification of BAX and GAPDH.

#### Expression analysis of ZFX

RT-qPCR was performed to determine if ZFX mRNA displayed an age-dependent expression pattern and results indicated an age-dependent change of ZFX mRNA expression in the postnatal mouse kidney. ZFX mRNA expression was greatest on the day 1, and then dropped gradually from day 1 through day 21 with the least expression on day 21 ( $P < 0.01$ ). Day 42 had similar expression to day 21 ( $P > 0.05$ ) while day 90 was slightly greater ( $P < 0.05$ ) (Fig. 3A).

#### Expression analysis of Bcl-2

The mRNA level of Bcl-2 also displayed an age-dependent expression pattern. On day 1, the expression of Bcl-2 was greatest, but lesser on days 7-90 ( $P < 0.01$ ). The least expression occurred day 21 compared to days 1-14 ( $P < 0.01$ ) with days 42 and 90 similar to day 21 ( $P > 0.05$ ) (Fig. 3B).

#### Expression analysis of BAX

The mRNA level of BAX in different periods of postnatal mouse kidney showed a trend similar to Bcl-2. Expression of BAX was greatest on day 1, and then declined on day 7 ( $P < 0.01$ ) and day 14 ( $P < 0.01$ ) until reached the least expression on day 21 ( $P < 0.01$ ), Expression of BAX increased slightly on day 42 ( $P < 0.05$ ) but expression on day 90 was similar to day 21 ( $P > 0.05$ ) (Fig. 3C).

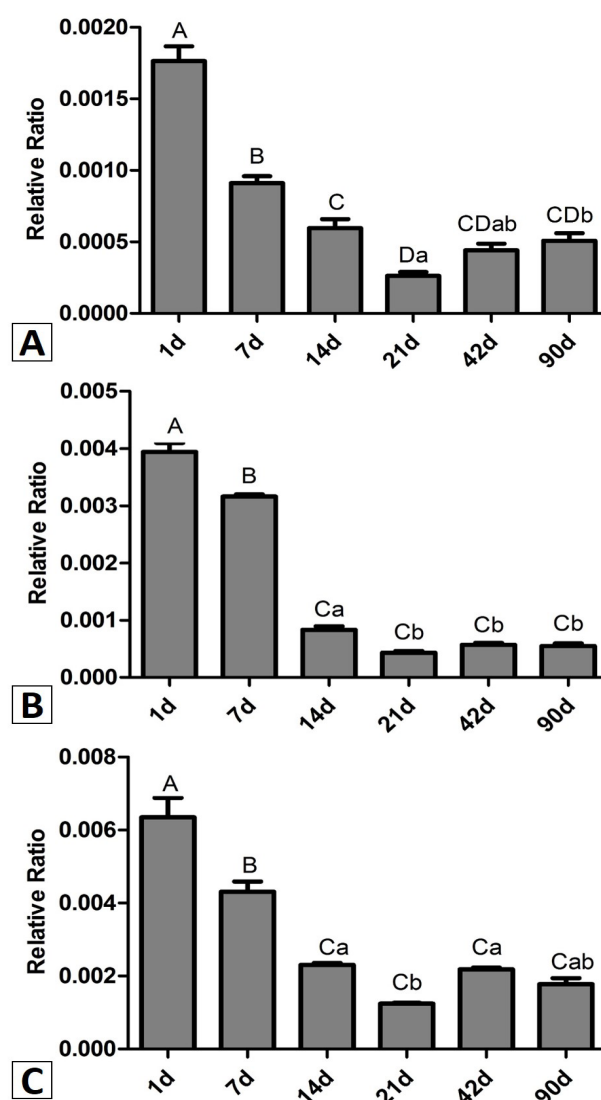


Fig. 3. mRNA expression of ZFX (A), Bcl-2 (B) and BAX (C) detected by RT-qPCR. Columns with the same letters indicate no difference between the two groups ( $P > 0.05$ ). Columns with differing capital letters differ ( $P < 0.01$ ) and columns with differing lowercase letters differ ( $P < 0.05$ ).

## DISCUSSION

Zinc finger protein plays an important role in the proliferation and differentiation of cells. The ZFX gene products are necessary for maintaining embryonic stem cells and adult stem cell self-renewal, and are a common molecular basis of embryonic stem cells development and adult stem cell regeneration (Cellot *et al.*, 2007; Ouyang *et al.*, 2009). Inactivation of the ZFX gene has been shown to cause increased mortality of neonatal mice and decreased reproductive capacity of surviving maternal mice (Luoh *et*

*et al.*, 1997). These studies also demonstrated that ZFX was closely related to cell proliferation.

Cell proliferation and apoptosis in mouse kidney development is regulated by a number of genes (Terzi *et al.*, 2000) and normal growth and development involves a balance relationship of cell proliferation and apoptosis. On day 1 of kidney development, the cell proliferation index of medullary collecting duct and renal tubular tissue was greater than the apoptosis index, with proliferation dominant. On day 7, cell apoptosis was dominant and cells begin to differentiate into mature cells. On day 14, the cell number and size of renal tubular and collecting duct tissue was not predominant with proliferation and apoptosis both at lesser levels. On day 21, the renal cortex and medulla were thickened with the development of the inner medulla, and cell proliferation decreased. At 42 days after birth, the adult kidney structure was largely formed and cell proliferation and apoptosis were in a relatively balanced state (Li *et al.*, 2006).

The Bcl-2 and BAX genes belong to the Bcl-2 family. The Bcl-2 gene is an anti-apoptotic gene and BAX is a pro-apoptotic gene, and are important genes in regulating cells apoptosis. The mechanism of Bcl-2 is through production of antioxidant, inhibiting the release of pro-apoptotic protein and inhibiting the cytotoxicity of BAX to achieve an anti-apoptotic effect. The BAX gene can form homodimers, damaging mitochondrial membranes, and the appearance of apoptosome in the cytoplasm, activating Caspase cascade activation, triggers the fragmentation of DNA and damage to a variety of cellular proteins components, which accelerates cell death (Saikumar *et al.*, 1998; Yang *et al.*, 1997). During postnatal kidney development Bcl-2 and BAX are both necessary for regulating cells apoptosis, and Bcl-2 deficient mice demonstrate polycystic kidneys (Veis *et al.*, 1993).

SYBR Green reverse transcriptase quantitative PCR (RT-qPCR) is the most widely method used to investigate gene expression in ruminants. The increase in the fluorescent signal is directly proportional to the number of PCR product molecules generated. It possible to simultaneously characterize different genes, using small quantities of sample with high specificity, sensitivity and accuracy (Giulietti *et al.*, 2001; Bustin, 2000).

In our study ZFX, Bcl-2, and BAX mRNA was confirmed to display an age-dependent expression pattern by RT-qPCR, mRNA expression figures were done to determine: (i) the relationship between the mRNA expression of ZFX, Bcl-2 and BAX; and (ii) whether ZFX mRNA age-dependent expression tendency was consistent with those of Bcl-2 and BAX. Results showed that mRNA expression of the ZFX, Bcl-2, and BAX genes was detected in all stages of postnatal kidney development

and displayed similar age-dependent changes. On day 1 of early development of medulla ZFX was strongly expressed, and with the maturation in medulla development, ZFX expression decreased gradually until 21 days when the medulla was fully developed and the expression of ZFX reached the minimum. While the expression trend of Bcl-2 and Bax during the development of postnatal kidney showed the cell proliferation and apoptosis were have the same patterns which were consistent with Zang *et al.* (2007). These results suggested that during the development of postnatal kidney, ZFX might ensure cell proliferation and viability, to promote cell further maturation. However, after day 42 when the kidney is mature, ZFX expression was increased so that renal cell proliferation and apoptosis were in relative equilibrium.

It was reported that ZFX was significantly up-regulated in renal cell carcinoma (RCC) tissues (Li *et al.*, 2015), and knockdown of ZFX suppresses renal carcinoma cell growth and induces apoptosis (Fang *et al.*, 2014). In addition congenital anomalies of the kidney and urinary tract (CAKUT)/ cystic kidney disease is the first primary cause of the end-stage renal disease (ESRD) in China's children (Tang *et al.*, 2014). According the expression pattern of ZFX in the postnatal kidney in our research, ZFX may be involved in the process of developing renal cell proliferation and balanced regulation of apoptosis.

## CONCLUSION

In this study, ZFX gene expression in the different maturation state of mouse kidney was tested using the real-time fluorescence quantitative technique, meanwhile Bcl-2, and BAX expression was tested. Through the comparison, it showed the ZFX gene may participate in the developmental process of the kidney through the balance of cell proliferation and apoptosis regulation. However, the role of ZFX in the balance of cell proliferation and apoptosis in kidney development needs further protein test to confirm or refute this hypothesis, and it maybe provide some information to research on the end-stage renal disease.

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### *Statement of conflict of interest*

Authors have declared no conflict of interest.



## REFERENCES

- Arenzana, T.L., Smith-Raska, M.R. and Reizis, B., 2009. Transcription factor ZFX controls BCR-induced proliferation and survival of B lymphocytes. *Blood*, **113**: 5857-5867. <https://doi.org/10.1182/blood-2008-11-188888>
- Bustin, S.A., 2000. Absolute quantification of mRNA using real-time reverse transcription polymerase chain reaction assays. *J. mol. Endocrinol.*, **25**: 169-193. <https://doi.org/10.1677/jme.0.0250169>
- Cellot, S. and Sauvageau, G., 2007. ZFX: At the crossroads of survival and self-renewal. *Cell*, **129**: 239-241. <https://doi.org/10.1016/j.cell.2007.04.002>
- Chandrasekharan, S., Kumar, S., Valley, C.M. and Rai, A., 2009. Proprietary science, open science and the role of patent disclosure: The case of zinc-finger proteins. *Nat. Biotechnol.*, **27**: 140-144. <https://doi.org/10.1038/nbt0209-140>
- Davies, J.A., 2002. Morphogenesis of the metanephric kidney. *Scient. World J.*, **2**: 1937-1950. <https://doi.org/10.1100/tsw.2002.854>
- Fang, Q., Fu, W.H., Yang, J., Li, Y., Zhou, Z.S., Chen, Z.W. and Pan, J.H., 2014. Knockdown of ZFX suppresses renal carcinoma cell growth and induces apoptosis. *Cancer Genet.*, **207**: 461-466. <https://doi.org/10.1016/j.cancergen.2014.08.007>
- Galan-Caridad, J.M., Harel, S., Arenzana, T.L., Hou, Z.E., Doetsch, F.K., Mirny, L.A. and Reizis, B., 2007. ZFX controls the self-renewal of embryonic and hematopoietic stem cells. *Cell*, **129**: 345-357. <https://doi.org/10.1016/j.cell.2007.03.014>
- Giulietti, A., Overbergh, L., Valckx, D., Decallonne, B., Bouillon, R. and Mathieu, C., 2001. An overview of real-time quantitative PCR: Applications to quantify cytokine gene expression. *Methods*, **25**: 386-401. <https://doi.org/10.1006/meth.2001.1261>
- Jiang, H., Zhang, L., Liu, J., Chen, Z., Na, R., Ding, G., Zhang, H. and Ding, Q., 2012. Knockdown of zinc finger protein X-linked inhibits prostate cancer cell proliferation and induces apoptosis by activating caspase-3 and caspase-9. *Cancer Gene Ther.*, **19**: 684-689. <https://doi.org/10.1038/cgt.2012.53>
- Kallio, J.P., Hirvikoski, P., Helin, H., Luukkaala, T., Tammela, T.L., Kellokumpu-Lehtinen, P. and Martikainen, P.M., 2004. Renal cell carcinoma MIB-1, BAX and Bcl-2 expression and prognosis. *J. Urol.*, **172**: 2158-2161. <https://doi.org/10.1097/01.ju.0000144334.97639.bf>
- Li, C.Y., Li, H.J., Zhang, T., Li, J.M., Ma, F.L., Li, M., Sui, Z.F. and Chang, J.W., 2015. ZFX is a Strong Predictor of Poor Prognosis in Renal Cell Carcinoma. *Med. Sci. Monit.*, **21**: 3380-3385. <https://doi.org/10.12659/MSM.894708>
- Li, X.M., Zang, D.Y. and Guo, M., 2006. Cell proliferation and apoptosis in mouse kidney during postnatal development. *Chinese J. Anat.*, **29**: 446-449.
- Lipschutz, J.H., 1998. Molecular development of the kidney: A review of the results of gene disruption studies. *Am. J. Kidney Dis.*, **31**: 383-397. <https://doi.org/10.1053/ajkd.1998.v31.pm9506676>
- Livak, K.J. and Schmittgen, T.D., 2001. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. *Methods*, **25**: 402-408. <https://doi.org/10.1006/meth.2001.1262>
- Luoh, S.W., Bain, P.A., Polakiewicz, R.D., Goodheart, M.L., Gardner, H., Jaenisch, R. and Page, D.C., 1997. ZFX mutation results in small animal size and reduced germ cell number in male and female mice. *Development*, **124**: 2275-228.
- Nikpour, P., Emadi-Baygi, M., Mohammad-Hashem, F., Maracy, M.R. and Haghjooy-Javanmard, S., 2012. Differential expression of ZFX gene in gastric cancer. *J. Biosci.*, **37**: 85-90. <https://doi.org/10.1007/s12038-011-9174-2>
- Nyengaard, J.R., 1993. The quantitative development of glomerular capillaries in rats with special reference to unbiased stereological estimates of their number and sizes. *Microvasc. Res.*, **45**: 243-261. <https://doi.org/10.1006/mvres.1993.1022>
- Ouyang, Z., Zhou, Q. and Wong, W.H., 2009. ChIP-Seq of transcription factors predicts absolute and differential gene expression in embryonic stem cells. *Proc. natl. Acad. Sci. USA*, **106**: 21521-21526. <https://doi.org/10.1073/pnas.0904863106>
- Rodríguez, A., Rodríguez, M., Córdoba, J.J. and Andrade, M.J., 2015. Design of primers and probes for quantitative real-time PCR methods. *Methods Mol. Biol.*, **1275**: 31-56. [https://doi.org/10.1007/978-1-4939-2365-6\\_3](https://doi.org/10.1007/978-1-4939-2365-6_3)
- Saikumar, P., Dong, Z., Weinberg, J.M. and Venkatachalam, M.A., 1998. Mechanisms of cell death in hypoxia/reoxygenation injury. *Oncogene*, **17**: 3341-3347. <https://doi.org/10.1038/sj.onc.1202579>
- Schneider-Gadicke, A., Beer-Romero, P., Brown, L.G., Mardon, G., Luoh, S.W. and Page, D.C., 1989. Putative transcription activator with alternative isoforms encoded by human ZFX gene. *Nature*, **342**: 708-711. <https://doi.org/10.1038/342708a0>
- Schnidar, H., Eberl, M., Klingler, S., Mangelberger, D., Kasper, M., Hauser-Kronberger, C., Regl,

- G., Kroismayr, R., Moriggl, R., Sibilio, M. and Aberger, F., 2009. Epidermal growth factor receptor signaling synergizes with Hedgehog/GLI in oncogenic transformation via activation of the MEK/ERK/JUN pathway. *Cancer Res.*, **69**: 1284-1292. <https://doi.org/10.1158/0008-5472.CAN-08-2331>
- Tang, X.S., Xu, H., Shen, Q., Sun, L., Miao, Q.F., Zhou, L.J., Cao, Q., Liu H.M., Chen, J., Rao, J., Zhai, Y.H. and Fang, X.Y., 2014. The spectrum of causes for end-stage renal disease in 113 pediatric patients. *Med. J.*, **37**: 382-385.
- Terzi, F., Burtin, M. and Friedlander, G., 2000. Cellular proliferation and apoptosis in the development and progression of renal diseases: Pharmacological interventions in genetically modified animals. *Annl. Pathol.*, **20**: S3-51.
- Veis, D.J., Sorenson, C.M., Shutter, J.R. and Korsmeyer, S.J., 1993. Bcl-2-deficient mice demonstrate fulminant lymphoid apoptosis, polycystic kidneys, and hypopigmented hair. *Cell*, **75**: 229-240. [https://doi.org/10.1016/0092-8674\(93\)80065-M](https://doi.org/10.1016/0092-8674(93)80065-M)
- Yamamoto, K., Tomita, N., Yoshimura, S., Nakagami, H., Taniyama, Y., Yamasaki, K., Ogihara, T. and Morishita, R., 2004. Hypoxia-induced renal epithelial cell death through caspase-dependent pathway: role of Bcl-2, Bcl-xL and BAX in tubular injury. *Int. J. mol. Med.*, **14**: 633-640. <https://doi.org/10.3892/ijmm.14.4.633>
- Yang, J., Lin, X., Kim, C.N., Ibrado, A.M., Cai, J., Peng, T.I., Jones, D.P. and Wang, X., 1997. Prevention of apoptosis by Bcl-2: Release of cytochrome C from mitochondria blocked. *Science*, **275**: 1129-1131. <https://doi.org/10.1126/science.275.5303.1129>
- Zang, D.Y., Li, X.M. and Guo, M., 2007. Relationship between the expressions of Bcl-2 and BAX and cell apoptosis in renal medullary of mice during postnatal development. *J. Clin. Rehab. Tissue Engineer. Res.*, **11**: 1473-1475.
- Zhou, Y., Su, Z., Huang, Y., Chen, S., Wu, T., Chen, G., Xie, X., Li, B. and Du, Z., 2011. The ZFX gene is expressed in human gliomas and is important in the proliferation and apoptosis of the human malignant glioma cell line U251. *J. exp. clin. Cancer Res.*, **30**: 114. <https://doi.org/10.1186/1756-9966-30-114>