



of this inflammatory disease and to identify potential genetic markers for resistance to mastitis in bovine. This article presents analytical study of bovine mammary gland genes to uncover underlying regulatory networks.

## MATERIALS AND METHODS

### *Literature survey and data collection*

This article presents analytical study of bovine mammary gland genes to uncover underlying regulatory networks. Network, functional and pathway analysis of 226 mammary gland genes were performed to find mastitis resistant genes. Data was collected from UniGene, NCBI, Ensembl and bovine genome databases. Network analysis was performed by STRING (Search Tool for the Retrieval of Interacting Genes/Proteins). For functional analysis DAVID (the Database for Annotation, Visualization and Integrated Discovery) was used and pathway analysis was performed through KEGG (Kyoto Encyclopedia of Genes and Genomes) (Kanehisa, 2013).

A list of genes representing part of the mammalian mammary gland transcriptome was assembled by an extensive literature survey and searching various databases. The UniGene which is an NCBI database of the

transcriptome was searched to identify genes expressed in bovine mammary gland. Literature published was reviewed by searching for the relevant publications through PubMed using key phrases: association, gene candidates, epigenetics, genetics, mammary gland, mastitis, methylation, milk, miRNA, QTL, SNP. A systematic review of both original research articles and reviews was performed for searching literature associated with bovine mammary gland genes. Data of genes was retrieved from the NCBI, Ensembl and bovine genome databases.

### *Network analysis*

Complete knowledge of all direct and indirect interactions between proteins in a given cell would represent an important milestone towards a comprehensive description of cellular mechanisms and functions. Network analysis of the list of mammary gland genes was completed by STRING. It sorted out our data according to predicted functional association among different genes and proteins. We used four views for our analysis which were, Co-expression, database, experiment and text mining at high confidence score of 0.700. This analysis helped in systematically making the group of genes performing similar functions and showing similar expression pattern.

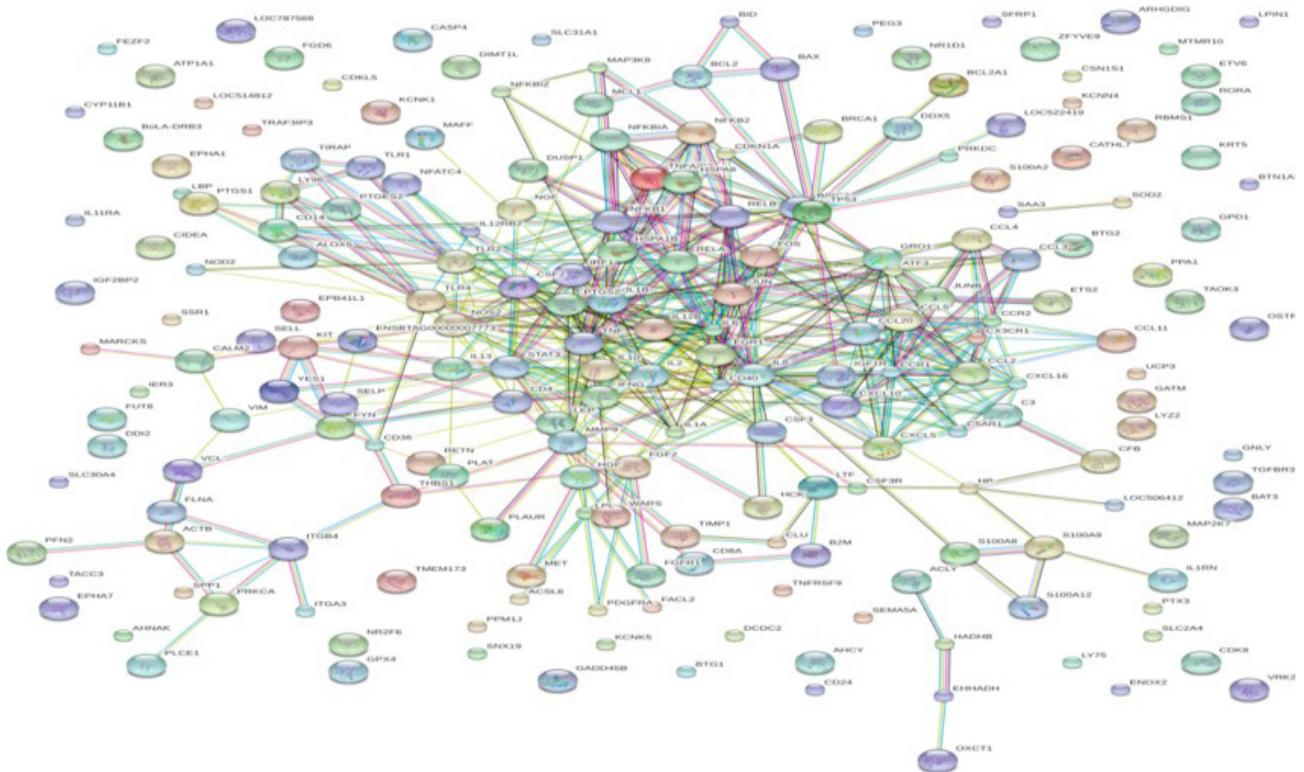


Fig. 1. The network showing 3 clusters: In the center there is the largest network of the dataset; in the middle there is a second largest network of genes and at the sides scattered genes are shown that are not interconnected to any other gene in the provided dataset.

**Table I.- Gene functional classification.**

<b>Gene Group 1</b>	<b>Enrichment Score: 14.712468824680236</b>
OFFICIAL_GENE_SYMBOL	Gene Name
CCL11	chemokine (C-C motif) ligand 11
IL13	interleukin 13
IL8	interleukin 8
CCL2	chemokine (C-C motif) ligand 2
CCL3	chemokine (C-C motif) ligand 3
CCL5	chemokine (C-C motif) ligand 5
CXCL16	chemokine (C-X-C motif) ligand 16
CXCL5	chemokine (C-X-C motif) ligand 5
CSF3	colony stimulating factor 3 (granulocyte)
CCL4	chemokine (C-C motif) ligand 4
IL1RN	interleukin 1 receptor antagonist
CCL20	chemokine (C-C motif) ligand 20
CXCL10	chemokine (C-X-C motif) ligand 10
<b>Gene Group 2</b>	<b>Enrichment Score: 6.197470798536051</b>
OFFICIAL_GENE_SYMBOL	Gene Name
IFNG	interferon, gamma
IL10	interleukin 10
IL1B	interleukin 1, beta
IL2	interleukin 2
TNF	tumor necrosis factor (TNF superfamily, member 2)
<b>Gene Group 3</b>	<b>Enrichment Score: 3.3291366663923707</b>
OFFICIAL_GENE_SYMBOL	Gene Name
RELA	v-relreticuloendotheliosis viral oncogene homolog A (avian); similar to v-relreticuloendotheliosis viral oncogene homolog A, nuclear factor of kappa light polypeptide gene enhancer in B-cells 3, p65
NFKB2	nuclear factor of kappa light polypeptide gene enhancer in B-cells 2 (p49/p100)
NFKB1	nuclear factor of kappa light polypeptide gene enhancer in B-cells 1
RELB	v-relreticuloendotheliosis viral oncogene homolog B
<b>Gene Group 4</b>	<b>Enrichment Score: 2.1814765223414283</b>
OFFICIAL_GENE_SYMBOL	Gene Name
IGF1R	insulin-like growth factor 1 receptor
YES1	v-yes-1 Yamaguchi sarcoma viral oncogene homolog 1
FGFR1	fibroblast growth factor receptor 1
MAP3K8	mitogen-activated protein kinase kinase 8
MET	met proto-oncogene (hepatocyte growth factor receptor)
HCK	hemopoietic cell kinase
KIT	v-kit Hardy-Zuckerman 4 feline sarcoma viral oncogene homolog
<b>Gene Group 5</b>	<b>Enrichment Score: 1.8117924213734606</b>
OFFICIAL_GENE_SYMBOL	Gene Name
S100A2	S100 calcium binding protein A2
S100A8	S100 calcium binding protein A8
S100A9	S100 calcium binding protein A9
S100A12	S100 calcium binding protein A12 (calgranulin C)
<b>Gene Group 6</b>	<b>Enrichment Score: 1.6666772544237716</b>
OFFICIAL_GENE_SYMBOL	Gene Name
CX3CR1	chemokine (C-X3-C motif) receptor 1
C5AR1	complement component 5a receptor 1
CCR2	chemokine (C-C motif) receptor 2
CCR1	chemokine (C-C motif) receptor 1

### Functional analysis

DAVID was used to perform analysis of functionally related genes. Functional annotation and gene functional classification tools were used for the functional analysis of mammary gland genes at  $p < 0.05$ . This gave us an in-depth understanding of the biological themes in list of genes that are enriched in genome-scale studies.

### Pathway analysis

Pathway analysis of mastitis resistant genes was performed through KEGG which gave an insight into pathways where the genes were involved and it helped to interpret the data in the context of biological processes, pathways and networks. This analysis helped to understand the mechanisms of mastitis, the unaffected pathways and the pathways that were are affected most. These findings ultimately helped to find resistance mechanism and genes involved in resistance.

## RESULTS

### Network analysis

A list of total 226 mammary gland genes was created by systematic and integrated review of literature and provided to STRING. Two hundred and five genes matched with STRING database for *Bos Taurus* and a network was generated of these 205 genes at high confidence score of 0.700 (Fig. 1). One hundred and thirty four genes form the largest network. Figure 2 shows the genes that are co-expressed.

### Functional analysis

DAVID was used for functional analysis of the short listed genes by network analysis. 134 genes that were the part of the largest network were selected for functional analysis to uncover the meaning of interaction of these genes. Functional annotation clustering resulted in a total of 72 clusters. The majority of gene ontology (GO) terms were related to immune response, for example, GO identifiers were related to inflammatory response (GO:0006954), chemotaxis (GO:0006935), immune response (GO:0006955), leukocyte migration (GO:0050900), response to lipopolysaccharide (GO:0032496), and TLR signalling pathway (GO:0002224). To get significant results, we applied an enrichment score cutoff criterion of  $>2$ . The gene list was further sorted out by using another DAVID tool of gene functional classification which sorted the gene list into 6 gene groups as shown in Table I.

### Pathway analysis

Pathway analysis of the functionally related genes was conducted by using KEGG database. Thirty five significant pathways related to these genes were identified among which 11 pathways were directly related to the immunity related pathways. Pathways other than immunity

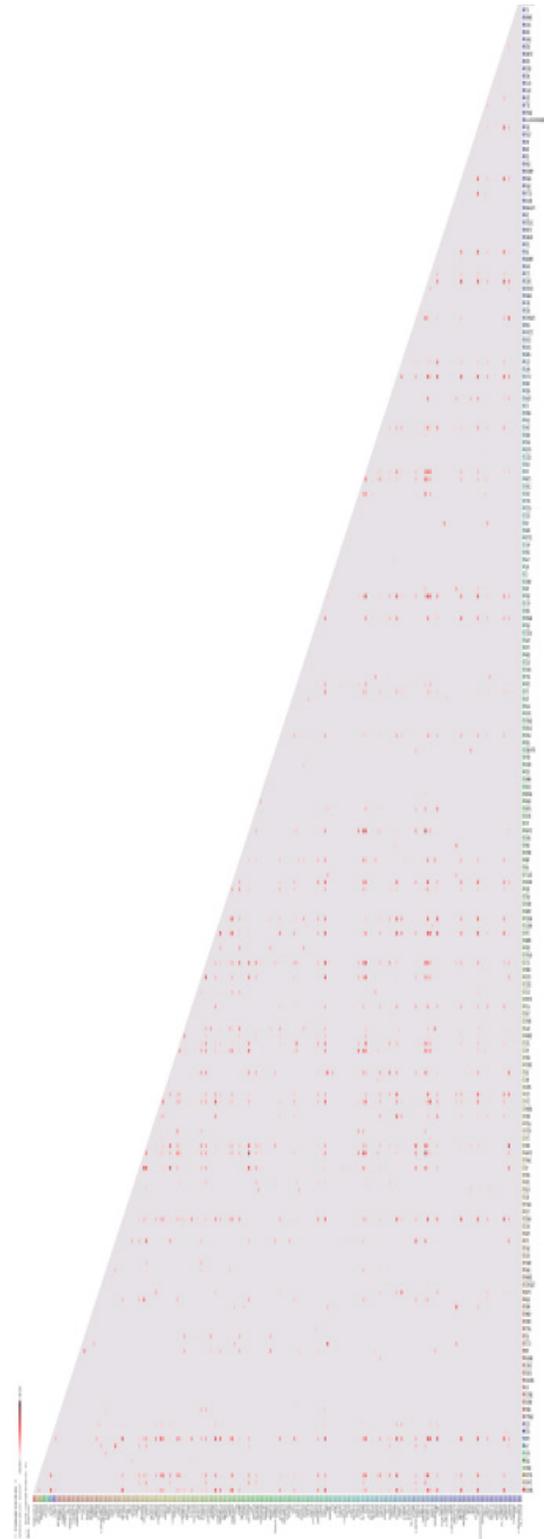


Fig. 2. Co-expressed genes in the dataset. The paler red color represent the less co-expressed genes, the dark red color shows the genes are more co-expressed.

**Table II.- KEGG pathway annotation.**

Category	Term
KEGG_PATHWAY	bta04060:Cytokine-cytokine receptor interaction
KEGG_PATHWAY	bta04620:Toll-like receptor signaling pathway
KEGG_PATHWAY	bta04621:NOD-like receptor signaling pathway
KEGG_PATHWAY	bta04660:T cell receptor signaling pathway
KEGG_PATHWAY	bta05200:Pathways in cancer
KEGG_PATHWAY	bta04010:MAPK signaling pathway
KEGG_PATHWAY	bta04062:Chemokine signaling pathway
KEGG_PATHWAY	bta04510:Focal adhesion
KEGG_PATHWAY	bta04640:Hematopoietic cell lineage
KEGG_PATHWAY	bta04210:Apoptosis
KEGG_PATHWAY	bta05222:Small cell lung cancer
KEGG_PATHWAY	bta04920:Adipocytokine signaling pathway
KEGG_PATHWAY	bta05215:Prostate cancer
KEGG_PATHWAY	bta05218:Melanoma
KEGG_PATHWAY	bta04623:Cytosolic DNA-sensing pathway
KEGG_PATHWAY	bta05330:Allograft rejection
KEGG_PATHWAY	bta04940:Type I diabetes mellitus
KEGG_PATHWAY	bta04622:RIG-I-like receptor signaling pathway
KEGG_PATHWAY	bta04630:Jak-STAT signaling pathway
KEGG_PATHWAY	bta04520:Adherens junction
KEGG_PATHWAY	bta05332:Graft-versus-host disease
KEGG_PATHWAY	bta05020:Prion diseases
KEGG_PATHWAY	bta05210:Colorectal cancer
KEGG_PATHWAY	bta05310:Asthma
KEGG_PATHWAY	bta04662:B cell receptor signaling pathway
KEGG_PATHWAY	bta04722:Neurotrophin signaling pathway
KEGG_PATHWAY	bta05214:Glioma
KEGG_PATHWAY	bta04810:Regulation of actin cytoskeleton
KEGG_PATHWAY	bta05219:Bladder cancer
KEGG_PATHWAY	bta04610:Complement and coagulation cascades
KEGG_PATHWAY	bta04650:Natural killer cell mediated cytotoxicity
KEGG_PATHWAY	bta05220:Chronic myeloid leukemia
KEGG_PATHWAY	bta04512:ECM-receptor interaction
KEGG_PATHWAY	bta05221:Acute myeloid leukemia
KEGG_PATHWAY	bta00590:Arachidonic acid metabolism

related pathways were apoptosis, small cell lung cancer, adipocytokine signaling pathway, prostate cancer, melanoma, cytosolic DNA-sensing pathway, allograft rejection, type I diabetes mellitus, adherens junction, graft-versus-host disease, prion diseases, colorectal cancer, asthma, glioma, regulation of actin cytoskeleton, bladder cancer, complement and coagulation cascades, chronic myeloid leukemia, ECM-receptor interaction, acute myeloid leukemia, arachidonic acid metabolism. Total pathways associated with the provided gene list are represented in [Table II](#).

## DISCUSSION

The biological complexity of animals unavoidably requires a systems biology approach, *i.e.*, a way to systematically study the complex biological interactions using a method of integration instead of reduction ([Loor and Cohick, 2009](#)). Important goals of systematic and integrated analysis are to uncover the underlying links (pathways, regulatory networks, and functional organization), and also to discover new emergent properties that may arise from examining the interactions between all components of a system ([Bruggeman and Westerhoff, 2007](#)). In the context of intra-mammary infection (IMI) and the mammary gland response, a “true” systems biology approach would encompass not only the intra-cellular networks of factors, but also communication between tissues ([Piantoni \*et al.\*, 2010](#)) and between organs/systems.

Most of the genes were related to functions including Immune Response, Immune Disease, Connective Tissue Disorders, Lipid and Carbohydrate Metabolism, Molecular Transport, Cell-To-Cell Signaling, Tissue Development, Cellular Development, and Immune and Lymphatic System Development and Function. These genes encoded cytokines (CSF3, CSF2, CCL3, TNF, CCR1, IL13, KIT, CCL5, CCL4, IL10, CXCL10, IL12RB2, IFNG, IL1B, IL1A, IL8, MET, CD40, HGF, LEP, CXCL16, CX3CR1, CCR2, PDGFRA, IL12B, IL2), lipid metabolism related genes (CD36, GPX4, LPIN1, LPL, LPB), transcription regulators (BCL3, FOS and NFKBIA), receptors (TNF, IL8, RELA, TLR1, TIRAP, TLR2, NFKBIA, NFKB1, TLR4, CD40, CCL5, CXCL10, FOS, JUN, MAP3K8, IL1B, LBP, IL12B, CD14, SPP1), and others such as SELP, SELL, and SOD1, all play a role in some aspect of the immune response including cytokine activity (IL10, TNF, IL8, and IL1B), cell adhesion (SELL and SELP), immune activation (CD14 and TLR2), acute phase reaction (TNF, IL1B, and SAA3), apoptosis (BCL2, BAX). Similar results were observed by Moyes *et al.* (2009) in their study on bovine mammary tissue challenged with *Streptococcus uberis*.

A French group examined the milk somatic cell transcriptome (predominantly PMN) of mastitis

susceptible and mastitis-resistant sheep infected successively by *Streptococcus epidermidis* (*S. epidermidis*) and *Staphylococcus aureus* (*S. aureus*) (Bonnefont *et al.*, 2011). The challenges per se (*i.e.*, *S. aureus* vs. *S. epidermidis*) resulted in >5,000 DEG (FDR= 0.05 and fold change >1.5 ratio between *S. aureus* and *S. epidermidis*), but the authors performed a functional analysis on those DEG with a >5-fold difference. The functional analysis of genes more expressed in *S. aureus* uncovered significantly-enrichment (EASE score  $\leq 0.05$ ) (da Huang *et al.*, 2009a), among DEG of functions related to the immune response; whereas, genes more expressed in *S. epidermidis* resulted in over representation of functions related to metabolism and cell growth, indicating that more T cells were recruited upon inoculation by *S. aureus* than *S. epidermidis*. Few DEG (57 at FDR=0.05 and fold-change >1.5) were found between the mastitis-susceptible and mastitis-resistant sheep. The functional analysis indicated that mastitis-resistant sheep had a more active immune response, as indicated by the significant enrichment of “leukocyte adhesion and activation” among DEG more expressed in mastitis-resistant relative to the mastitis-susceptible sheep (Bonnefont *et al.*, 2011).

Another German group performed direct transcriptomics comparison between primary bovine mammary epithelial cells isolated from cows with low- or high susceptibility to mastitis challenged either with *E. coli* or *S. aureus* (Brand *et al.*, 2011). Similarly to the French study mentioned above (Bonnefont *et al.*, 2011), the main finding from the ORA functional analysis was that the transcriptome of mammary epithelial cells from cows with low-susceptibility to mastitis challenged with either *S. aureus* or *E. coli* had a greater response to the bacterial challenge via up-regulation of genes associated with ‘acute phase response signaling’, *i.e.*, interleukin-6 (IL6) and nuclear factor NF-kappa-B p100 subunit (NFKB2), than cows with high-susceptibility to mastitis (Brand *et al.*, 2011). Both the German and the French studies appear to support the notion that the sensitivity of the mammary gland to bacteria plays an important role in the resistance to invading microorganisms. Perhaps the evolutionary transition of the mammary gland from a mucus-secreting surface epithelium serving a protective role to a milk-secreting gland with both protective and nutritional roles (Vorbach *et al.*, 2006) has, as a consequence, a negative impact on the importance of the tissue’s innate immune system.

## CONCLUSION

It was concluded that the stronger immune system of an animal would be more resistant to mastitis. In

future, the analysis of this study can be used to further our knowledge about this disease which is causing a huge economical lose worldwide, however, a deeper understanding of its fundamentals and development of more adequate bioinformatics tools is critically needed in order to really benefit from such an approach.

In conclusion, our network-based gene prioritization approach provides a general framework for identifying and ranking genes associated with complex diseases. To our knowledge this is the first time that protein interactions, orthologue mapping, gene expression, and literature mining were integrated for ranking candidate genes in any livestock species. The approaches and techniques that were systematically implemented in the present study are general and not confined by specific trait or species and can be applied to various complex diseases in different organisms.

## Conflict of interest statement

We declare that we have no conflict of interest.

## REFERENCES

- Bauman, D.E., Mather, I.H., Wall, R.J. and Lock, A.L., 2006. Major advances associated with the biosynthesis of milk. *J. Dairy Sci.*, **89**: 1235-1243. [http://dx.doi.org/10.3168/jds.S0022-0302\(06\)72192-0](http://dx.doi.org/10.3168/jds.S0022-0302(06)72192-0)
- Bionaz, M., Periasamy, K., Rodriguez-Zas, S.L., Everts, R.E., Lewin, H.A., Hurley, W.L. and Loor, J.J., 2012. Old and new stories: Revelations from functional analysis of the bovine mammary transcriptome during the lactation cycle. *PLoS One*, **7**: e33268. <http://dx.doi.org/10.1371/journal.pone.0033268>
- Bonnefont, C.M.D., Toufeer, M., Caubet, C., Foulon, E., Tasca, C., Aurel, M., Bergonier, D., Boullier, S., Robert-Granie, C., Foucras, G. and Rupp, R., 2011. Transcriptomic analysis of milk somatic cells in mastitis resistant and susceptible sheep upon challenge with *Staphylococcus epidermidis* and *Staphylococcus aureus*. *BMC Genomics*, **12**: 208. <http://dx.doi.org/10.1186/1471-2164-12-208>
- Brand, B., Hartmann, A., Repsilber, D., Griesbeck-Zilch, B., Wellnitz, O., Kuhn, C., Ponsuksili, S., Meyer, H.H. and Schwerin, M., 2011. Comparative expression profiling of *E. coli* and *S. aureus* inoculated primary mammary gland cells sampled from cows with different genetic predispositions for somatic cell score. *Genet. Sel. Evol.*, **43**: 24. <http://dx.doi.org/10.1186/1297-9686-43-24>
- Bruggeman, F.J. and Westerhoff, H.V., 2007. The nature of systems biology. *Trends. Microbiol.*, **15**: 45-50.

- <http://dx.doi.org/10.1016/j.tim.2006.11.003>  
Huang da, W., Sherman, B.T. and Lempicki, R.A., 2009a. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nature Protoc.*, **4**: 44-57.
- Jensen, K., Gunther, J., Talbot, R., Petzl, W., Zerbe, H., Schuberth, H., Seyfert, H. and Glass, E.J., 2013. *Escherichia coli* and *Staphylococcus aureus* induced mastitis differentially modulate transcriptional responses in neighboring uninfected bovine mammary gland quarters. *BMC Genomics*, **14**: 36. <http://dx.doi.org/10.1186/1471-2164-14-36>
- Kanehisa, M., 2013. Molecular network analysis of diseases and drugs in KEGG. *Methods Mol. Biol.*, **939**: 263-275. [http://dx.doi.org/10.1007/978-1-62703-107-3\\_17](http://dx.doi.org/10.1007/978-1-62703-107-3_17)
- Khan, M.Z. and Khan, A., 2006. Basic facts of mastitis in dairy animals: a review. *Pakistan Vet. J.*, **26**: 204-208.
- Lemay, D.G., Neville, M.C., Rudolph, M.C., Pollard, K.S. and German, J.B., 2007. Gene regulatory networks in lactation: identification of global principles using bioinformatics. *BMC Syst. Biol.*, **1**: 56. <http://dx.doi.org/10.1186/1752-0509-1-56>
- Loor, J.J. and Cohick, W.S., 2009. ASAS centennial paper: lactation biology for the twenty-first century. *J. Anim. Sci.*, **87**: 813-824. <http://dx.doi.org/10.2527/jas.2008-1375>
- Moyes, K.M., Drackley, J.K., Morin, D.E., Bionaz, M., Rodriguez-Zas, S.L., Everts, R.E., Lewin, H.A. and Loor, J.J., 2009. Gene network and pathway analysis of bovine mammary tissue challenged with *Streptococcus uberis* reveals induction of cell proliferation and inhibition of PPAR $\gamma$  signaling as potential mechanism for the negative relationships between immune response and lipid metabolism. *BMC Genomics*, **10**: 542. <http://dx.doi.org/10.1186/1471-2164-10-542>
- Paape, M.J., Bannerman, D.D., Zhao, X. and Lee, J., 2003. The bovine neutrophil: structure and function in blood and milk. *Vet. Res.*, **34**: 597-627. <http://dx.doi.org/10.1051/vetres:2003024>
- Piantoni, P., Bionaz, M., Graugnard, D.E., Daniels, K.M., Everts, R.E., Rodriguez-Zas, S.L., Lewin, H.A., Hurley, H.L., Akers, M. and Loor, J.J., 2010. Functional and gene network analyses of transcriptional signatures characterizing preweaned bovine mammary parenchyma or fat pad uncovered novel inter-tissue signaling networks during development. *BMC Genomics*, **11**: 331. <http://dx.doi.org/10.1186/1471-2164-11-331>
- Rudolph, M.C., McManaman, J.L., Hunter, L., Phang, T. and Neville, M.C., 2003. Functional development of the mammary gland: use of expression profiling and trajectory clustering to reveal changes in gene expression during pregnancy, lactation, and involution. *J. Mammary Gland Biol. Neoplasia*, **8**: 287-307. <http://dx.doi.org/10.1023/B:JOMG.0000010030.73983.57>
- Schukken, Y.H., Günther, J., Fitzpatrick, J., Fontaine, M.C., Goetze, L., Holst, O., Leigh, J., Petzl, W., Schuberth, H.J., Sipka, A., Smith, D.G., Quesnell, R., Watts, J., Yancey, R., Zerbe, H., Gurjar, A., Zadoks, R.N. and Seyfert, H.M., 2011. Host-response patterns of intramammary infections in dairy cows. *Vet. Immunol. Immunopathol.*, **144**: 270-289. <http://dx.doi.org/10.1016/j.vetimm.2011.08.022>
- Viguiier, C., Arora, S., Gilmartin, N., Welbeck, K. and O'Kennedy, R., 2009. Mastitis detection: current trends and future perspectives. *Trends Biotech.*, **27**: 486-493. <http://dx.doi.org/10.1016/j.tibtech.2009.05.004>
- Vorbach, C., Capecchi, M.R. and Penninger, J.M., 2006. Evolution of the mammary gland from the innate immune system? *Bioassays*, **28**: 606-616. <http://dx.doi.org/10.1002/bies.20423>
- Wiley, A.S., 2007. The globalization of cow's milk production and consumption: Biocultural perspectives. *Ecol. Fd. Nutri.*, **46**: 281-312. <http://dx.doi.org/10.1080/03670240701407657>