



Research Article

Effect of Selenium on Growth and Chemical Properties of Tomato Hybrid Salar F1

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Abstract | An experiment was carried out in plots during 2016 and 2017 to evaluate the effect of different levels of selenium application in irrigation water and foliar spray on physicochemical parameters of tomato hybrid Salar F1 grown in simple plastic tunnels using randomized complete block design. The data showed that plant height in centimeter, buds, flower, leaves and leaflets in numbers per plants with a range of 131.33 to 176.67 cm, 60 to 105, 41.5 to 60.0, 50.66 to 77.167 and 7.607 to 15.33, 30.33 to 82.0, respectively were significantly affected ($P < 0.05$) by the interaction of selenium application in irrigation and foliar spray in relation to season. The effect was also significant ($P < 0.05$) on some minerals like Cu, Mn, Zinc, Mg and Se that ranged from 0.2080 to 0.3150, 0.1260 to 0.2520, 0.2012 to 0.2970 and 18.04 to 32.09, 0.2147 to 0.5257 mg/Kg, respectively. Most of the proximate parameters of leaves like moisture, ash, crude fiber, crude protein, crude fat ranged from 92.082 to 92.317, 3.097 to 3.85, 4.9133 to 5.2717, 5.0133 to 5.2867, 1.08 to 1.24, 0.0236 to 1.9567 g/100g and that of fruits with lesser values were also affected significantly ($P < 0.05$). From the present study it was concluded that Se applied in the form of sodium selenite in irrigation and foliar spray considerably affected the physical parameter of tomato hybrid Salar F1, followed by proximate composition while minerals content was less affected. It is recommended that selenium may be added in moderate amount to plants for their physical well-being and for improvement of some chemical parameters.

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Introduction

Tomato is edible red berry fruit, belongs to South America region and it is used in daily foods in various forms. Tomato is rich in Lycopene and other essential nutrients. Tomato family is Solanaceae also called Nightshade that can reach a height of 1 to 5 meters i.e. 3–16 ft. The stem of tomato is tender and mostly creeps on ground or climb on other plants or supports. It is perennial in its native habitat, although often grown in temperate climates as annual crop.

Interest in the biological impacts of selenium (Se) is escalating because of its essentiality for humans and animals. Se in food mainly comes from plants sources and focus on effect of selenium uptake will prove its essentiality for fortification in plants. The deficiency of selenium causes keshan disease (fatal cardiomyopathy), kashin-beck disease. The selenium deficiency also badly affects the thyroid and immune system functions (Combs, 2001). In human, the nutritional function of Se is fulfilled by the selenoenzymes/ selenoproteins such as glutathione

peroxidase, thioredoxin reductase and iodothyronine 5'-deiodinase that are involved in hormonal regulations. Humans need Se in their diet for at least 25 different proteins, mostly antioxidant enzymes and the recommended dietary allowance (RDA) is 55- 200 µg/day for an adult. It is for these reasons; there is a resurgence of interest in Se fortification in higher plants during the last decade researches. The importance of Se for plant growth has not yet been fully understood and a lot of work has to be done but may be helpful in bio-accumulation.

As studies revealed that Se is very important in human diet and its deficiency causes various diseases that compel the scientists to work for overcoming its deficiency in human diets. Deficiency of Se prevailed in various countries like UK, Australia, New Zealand, China and many others (Chen *et al.*, 2002), where Se is deficient in their soils and crops grown in those countries contained negligible amount of Se (Combs, 2001).

Se in the plants mainly depends on soil concentration of selenium and soil properties e.g. Higher pH help in uptake of Se by plants (Chaney, 1994). Selenium concentration also related with plant sulphur content and with some of the soil components including CaCO₃ and the ratio of sand and silt in the soil (Dhillon *et al.*, 1992; Mayland *et al.*, 1990). The present work is the continuation of such type of research where effect of selenium has been studied on tomato hybrid Salar F1 applied in the form of Na selenite in irrigation water and foliar spray during two cropping season of tomato.

Materials and Methods

Study design and field layout

The experiment was carried out in Randomized Complete Block (RCB) factorial design with three 3 replications. The factors and their levels included foliar application of Se (F) with four levels (0, 5, 15, 20 mg/Kg) and Se application via irrigation water (I) with three (0, 50, 75 mg/Kg) levels replicated three times. The experiment was repeated over two seasons. Thus, the total numbers of treatment combinations in the experiment were 3×4×3= 36 per season. The plot size was 4x4 feet. All other agronomic practices for tomatoes cultivation were carried out as standard recommended in literature.

The following parameters were studied during the

experiment:

Moisture content

The drying method was used for moisture determination (AOAC, 2016). A sample of two gram (W₁) was weighted by electric balance in petri dishes with lids. Then the Petri plates with samples were completely dried in oven at a temperature of 105 °C. The samples were then covered with its lids and that was cooled in desiccator. When the plates with samples got cool then the sample was reweighted (W₂). The moisture in g/100g was calculated as follows.

$$\% \text{ moisture} = \frac{W_1 - w_2}{\text{weight of sample}} \times 100$$

Where as:

W₁ = Initial weight of Petri dish + sample; W₂ = Final weight of Petri dish + sample.

Crude protein

For the determination of nitrogen percentage in tomato, Kjeldhal method was used (AOAC, 2016). For digestion of samples sulfuric acid in concentrated form was used in addition of digestion mixture (7g K₂SO₄:1g CuSO₄). In the digestion tube, 2.0 g of samples was added with 15 mL of sulfuric acids and was heated up to 300 °C using digester. The greenish color digest was then cooled diluted to 100 mL with distilled water in volumetric flasks. For distillation 10 mL of the samples was taken and 40% Sodium hydroxide was added in the reaction tube of micro kjeldhal distillation apparatus. During heating with steam, ammonia was produced which was collected in the receiving flask containing boric acid (4%) with modified methyl red as indicator. Ammonia changed the colored yellow solution which then was titrated with 0.05 N HCl to calculate the percent nitrogen. The same procedure was repeated for blank and %N obtained was then multiplied with 6.25 as protein factor. The following formula was used.

$$\% N = \frac{(S - B) \times N \times 0.014 \times D \times 100}{\text{Wt. of sample} \times V}$$

Crude fat

Crude fat was determined by Soxhlet apparatus method (AOAC, 2016). The extraction solvent was petroleum ether (40-60°C). Each sample (1.0 g) was weighed and wrapped in filter paper, kept in the

thimble and transferred into the extraction tube. The round bottom flask was weighed and one third of it was filled with petroleum ether and connected to the extraction tube. After various siphonings, the flasks were dried and cooled and weighed. Crude fats were calculated as under.

$$\text{Crude fat, (\%)} = \frac{\text{Weight of, flask with fat} - \text{weight of, empty flask}}{\text{Weight of, original sample}} \times 100$$

Crude fiber

Crude fiber was determined by acid base digestion method (AOAC, 2016). For acid digestion, 200 ml of 2% H₂SO₄ was taken in a 500 mL beaker and 2.0 g sample was added into it. The samples were placed on water bath for 30 min. The digested was then filtered through muslin cloth. The acid digest was transferred into 500 mL beaker containing 200 mL of 2% NaOH. After alkali digestion the sample was filtered again with muslin cloth and weighted and dried in oven completely at 100 °C. After drying the digest was transferred into pre-weighted crucible and kept at 550 °C in muffle furnace. The crucible was weighted again and kept in desiccator for cooling. The crude fiber percentage was determined as under:

$$\text{Crude fiber, (\%)} = \frac{W1 - W2}{\text{Weight of sample}} \times 100$$

Total ash

Ash content was determined by combustion method (AOAC, 2016). Two grams of ground tomato sample was taken in a crucible and weight was taken (W1). The sample in crucible was charred with blowing flame and then ignited in muffle furnace at 550 °C into grayish white residues. The samples were cooled in desiccator. It was weighed again (W2) accurately and ash contents was calculated as.

$$\text{Ash, (\%)} = \frac{\text{Weight, of ash}}{\text{Weight of, sample}} \times 100$$

Nitrogen free extracts (NFE)

The nitrogen free extract (NFE) is the total digestible carbohydrate of tomato plant and fruit samples. It was calculated through subtraction as follows:

$$\text{NFE} = 100 - (\% \text{ ash} + \% \text{ crude fat} + \% \text{ crude fiber} + \% \text{ crude protein})$$

Minerals analysis

Tomato samples were analyzed for its minerals contents by following (AOAC, 2016) procedures.

Sodium and potassium were determined by flame photometer. Flame photometer was used for Sodium and Potassium determination. Other minerals i.e. Cu, Ca, Fe, Mg, Mn, P, Se and Zn were analyzed through atomic absorption.

Preparation of acid digests

The tomato samples (1.0 g) were digested to colorless liquid with conc. HNO₃ and perchloric acid (HClO₄) at the ratio of 7:3 in medium sized digestion tubes using hot plates. These digests were diluted to 100 ml with distilled water.

Na and K

Flame photo meter (PEP7) was used for determination of Na and K in the tomato samples.

Preparation of standard curve: Technical grade NaCl and KCl (Murk) was taken and 100 ppm solution was made for Na and K in double distilled water. These solutions were then diluted to 4 different concentrations of 20, 40, 80 and 100 ppm itself. The emission reading for these dilutions were noted and standard curves were developed using MS excel for further quantitative determination of Na and K in the samples.

Sample assay: The acid digest of tomato samples was taken in beakers and emission reading was noted by Flame photometer using Na and K filters. The amount of Na and K was calculated in mg/Kg which was then mathematically change into mg/100 g.

Phosphorus determination

Phosphorus was determined by molybdate method. For molybdate solution 7.5 g of ammonium paramolybdate (NH₄) Mo₇O₂₄·4H₂O was dissolved in 250 mL of deionized water. Similarly, for H₂SO₄ solution 70 mL of concentrated sulfuric acid was added to 450 mL of DI water. The ascorbic acid solution was prepared by dissolving 13.5 g of ascorbic acid in 250 mL of DI water. Potassium antimonyl-tartrate solution was prepared by adding 0.34 g in 250 mL of DI water. Then mixed reagent was obtained by adding together 100 mL ammonium molybdate, 250 mL sulfuric acid, 100 mL ascorbic acid, and 50 mL of potassium antimonyl-tartrate solutions. A stock solution of phosphorus 10 mM was prepared from 0.6805 g of KH₂PO₄ (fw = 136.09) in 100 mL DI water using a volumetric flask. For secondary stock solution of 50 mM, 100 µL of the primary stock solution was

diluted up to 100 mL with deionized water using a volumetric flask. Then 1.0 mL from diluted solution was taken and 4.0 mL coloring reagent and 15 mL of water were added. It was kept for 15 min. to develop color. Absorption reading was taken at 880 nm wave length using spectrophotometer. Similarly, absorption readings of all the samples were taken and the data were compared with standard curve as under.

$$\text{Concentration of P (mg/kg)} = \frac{\text{Graphical reading} \times \text{Dilution} \times 100 \times 1000}{\text{weight of sample}}$$

Micro mineral analysis

Micro mineral was determined by flame Atomic absorption spectrophotometer (Perkin Elmer) using the respective cathode lamp of the minerals. The sample was sucked through the flame and the reading was noted in ppm. For calibration of the instrument the standard solution of the required minerals was used provided with machine.

Agronomic characteristics

Plant height: Measuring tape was used to measure plant height from the base to the tip of the plant.

Number of flowering buds per plant: Buds as they are the initial form of flower were counted when it started growing on plant.

Number of flowers per plant: Buds then changed in the flower and when this phenomenon was started the flower count was started.

Number of fruits per plant: Flowers changed into fruits and as this phenomena was started the fruits were counted.

Chlorophyll analysis

For chlorophyll measurement the instrument used was AT leaf chlorophyll meter. The unit used was µg/g.

Lycopene analysis

For Lycopene measurement spectrophotometer sp 3000 was used. The reagents of HPLC grade was used where they were taken in the 2:1:1 including hexane, acetone and ethanol for lycopene extraction from tomato. After washing the tomato was turned into juice in blender. A portion of juice 100 µL was taken through pipette into 20 mL screw cap tube. In this juice, 8.0 mL of the mix solvents were mixed and vortexed. Then the sample was kept in the dark for

incubation at room temperature. 1.0 mL of distilled water was added and vortexed again. The sample was kept undisturbed for about 20 min. where two phases were formed. In this procedure distilled water was used as blank. The upper layer was taken into a prewashed cuvette. The maximum absorption was 503 nm. First the spectrophotometer was made zero with blank.

$$\text{Lycopene } \left(\frac{\text{mg}}{\text{Kg fresh wt.}} \right) = (A503 \times 537 \times 8 \times 0.55) / (0.10 \times 172)$$

The formula showed that 537 g/mole is the molecular weight of Lycopene, 8 mL is the volume of mixed solvent, 0.55 is the volume ratio of the upper layer to the mixed solvents, 0.10 g is the weight of tomato added, and 172 mM⁻¹ is the extinction coefficient for Lycopene in hexane

Statistical design

Data was analyzed using the statistical package statistix 8.1 (USA) and the significant differences between treatments was determined using least significant difference (LSD) test for main as well as interaction effects.

Results and Discussion

A field experiment was conducted in two consecutive seasons to study the effect of selenium on plant growth parameters, proximate composition, mineral and other chemical constituents of tomato cultivar Salar F1, in randomized complete block design with three factors factorial experiment.

The data of growth parameters (Table 1) showed that average height (cm), no of leaves, leaflets, buds, flowers and fruits per plant ranged from 131.33 to 176.67, 6.67 to 15.33, 30.33 to 82, 61.33 to 116.33, 50.67 to 77.17, 12 to 58.67, respectively. Different factors and their interaction (Table 5) showed that height was significantly affected (P<0.05) by all factors and their interaction except season and season into irrigation interaction. The number of leaves/plant was significantly affected by season and foliar versus irrigation interaction while number of leaflets/plant was affected by all the three factors and their interactions. Number of buds/plant were also affected by irrigation, season and by the interaction of irrigation with foliar and season. Number of flower/plant was affected by foliar, season and by the interaction of foliar with irrigation and season while

Table 1: Physical parameters of tomato hybrid Salar F1 as affected by Selenium application (mg/Kg) in irrigation and as foliar spray in two seasons.

IA	FA	SS	hieght/p	Leaves/Plant	Leaflets/Plant	Buds/Plant	Flowers/Plant	Fruits/plant
1	1	a	155 defg	7.667 ^{cd}	30.333 ^k	61.33 ^f	50.667 ^h	26.333 ^{cde}
		b	141 hij	8 ^{cd}	50 ^{fdg}	82.67 ^{cd}	66.333 ^{bcd}	50.667 ^{ab}
	2	a	166.33 abcd	8.000 ^{cd}	36.667 ^{jk}	65.67 ^{ef}	53.167 ^{gh}	38.667 ^{abcd}
		b	171.33 abc	10.333 ^{abcd}	52.333 ^{fd}	94.00 ^{bc}	62.500 ^{cdefg}	45 ^{abc}
	3	a	148 ghi	9.667 ^{bcd}	34 ^k	61.67 ^f	51.000 ^h	24 ^{cde}
		b	176.67 a	9.667 ^{bcd}	49 ^{ghi}	76.33 ^{de}	64.167 ^{bcd}	21 ^{de}
	4	a	140.67 hij	8.667 ^{cd}	38 ^{jk}	70.00 ^{def}	60.667 ^{cdefgh}	38.667 ^{abcd}
		b	137.33 ij	15.333 ^a	82 ^a	84.00 ^{cd}	77.167 ^a	49.667 ^{ab}
2	1	a	153.33 efgh	8.667 ^{cd}	39.667 ^{hijk}	82.00 ^{cd}	66.333 ^{bcd}	50.333 ^{ab}
		b	131.33 j	12 ^{abc}	60 ^{def}	116.33 ^a	71.000 ^{ab}	39.667 ^{abcd}
	2	a	160.67 bcdefg	9.000 ^{cd}	39.333 ^{hijk}	71.33 ^{def}	59.167 ^{cdefgh}	12 ^e
		b	153.33 efgh	9.667 ^{bcd}	46 ^{ghij}	114.33 ^a	67.833 ^{bcd}	58.667 ^a
	3	a	158.67 cdefg	6.667 ^d	34 ^k	65.00 ^{ef}	58.667 ^{defgh}	20 ^{de}
		b	161.33 bcdef	10 ^{bcd}	53.333 ^{efg}	102.67 ^{ab}	61.000 ^{cdefg}	31 ^{bcd}
	4	a	171 abc	7.667 ^{cd}	36.667 ^{jk}	67.00 ^{ef}	58.833 ^{defgh}	32 ^{bcd}
		b	173 ab	11.333 ^{abcd}	67.667 ^{bcd}	102.33 ^{ab}	63.500 ^{bcd}	39.333 ^{abcd}
	1	a	157.33 defg	8.667 ^{cd}	39 ^{ijk}	66.67 ^{ef}	56.000 ^{efgh}	30 ^{bcd}
		b	154 defg	11.667 ^{abcd}	78.667 ^a	92.00 ^{bc}	67.500 ^{bcd}	30 ^{bcd}
	2	a	165.67 abcde	7.667 ^{cd}	33 ^k	64.00 ^{ef}	54.833 ^{fgh}	33.667 ^{bcd}
		b	164.67 abcde	14.667 ^{ab}	71.333 ^{abc}	99.67 ^{bc}	56.000 ^{efgh}	41 ^{abcd}
3	3	a	163.67 bcde	8.333 ^{cd}	37.667 ^{jk}	66.00 ^{ef}	58.833 ^{defgh}	33.667 ^{bcd}
		b	164.67 abcde	12.333 ^{abc}	77.333 ^{ab}	104.00 ^{ab}	59.667 ^{defgh}	43.667 ^{abc}
	4	a	164.33 abcde	7.667 ^{cd}	37.333 ^{jk}	60.67 ^f	58.833 ^{defgh}	38 ^{abcd}
		b	149 fghi	9.667 ^{bcd}	64 ^{cde}	105.00 ^{ab}	64.500 ^{bcd}	34.667 ^{bcd}

number of fruits/plant was highly affected by season and by the interaction of the three factors.

The present study was in agreement with the work done by Nancy *et al.* (2014) who reported that the application of Se can increase fruit yield by increasing shoot length. Akbulut and Cakir (2010), and Djanaguirman *et al.* (2005) also concluded that selenium application increases leave numbers by decreasing leaf abscission which assisted the present study. Han-Wens (2010) reported that Se increase meiosis of meristematic cells that may result in higher leaves number. Increase in leaf numbers may be due to stimulation of cell division, increase photosynthesis, chlorophyll and carbon fixation (Malik *et al.*, 2011). Fu *et al.* (2011) reported that application of Se at moderate level can promote plant growth parameters like leaves, buds etc. Prins *et al.* (2011) reckoned to the increase of buds with application of the selenium. Xue *et al.* (2001) and Djanaguiraman *et al.* (2005) work on rice, lettuce and soybean and reported same results of increasing yield with application of selenium. Chi *et al.* (2017) also founded that Se application can

increase number of fruits.

The mineral content of tomato fruits (Table 2) showed considerable variation under the effect of different factors. Among different minerals in mg/kg Cu ranged from 0.186 to 0.324, Mn from 0.119 to 0.257, Zn from 0.125 to 0.301, Fe from 1.244 to 3.09, Ni from 0.211 to 0.92, Pb from 0.119 to 0.614, Cr from 0.011 to 0.331, Se from 0.19 to 0.564, Ca from 72.279 to 128.35 and Mg was ranged from 18.029 to 33.02, respectively. The effect of factors and their interaction (Table 6) on different mineral showed that Cu was significantly affected by foliar application of Se and its interaction with irrigation. Zn, Fe, Ni, Cr, Pb and Mg were significantly affected by Foliar, Irrigation application and also by season. The interaction of foliar and irrigation applications was also significant in case of these minerals. Mg was also affected by the interaction of Foliar and Season. Ca, Mn and Se were affected by foliar and irrigation applications and also by their interaction.

Table 2: Mineral content (mg/Kg*) of tomato hybrid Salar F1 as affected by Selenium application (mg/Kg) in irrigation and foliar spray in two seasons.

IA	FA	SS	Cu	Mn	Zn	Fe	Ni	Pb	Cr	Se	Ca	Mg
0	0	a	0.2190 ^{efgh}	0.1253 ^k	0.2367 ^{fghi}	1.2476 ⁱ	0.6633 ^{def}	0.1987 ^{ij}	0.1146 ^c	0.2147 ^h	72.285 ^l	18.04 ⁿ
		b	0.2110 ^{gh}	0.1610 ^{ghij}	0.2370 ^{fghi}	1.3053 ^{hi}	0.8000 ^{ab}	0.2287 ^{hi}	0.1206 ^c	0.2317 ^{fgh}	72.653 ^l	18.28 ⁿ
	5	a	0.2500 ^{cdef}	0.1507 ^{ghijk}	0.2717 ^{bc}	2.0866 ^{cde}	0.8200 ^a	0.5710 ^a	0.0163 ^l	0.2507 ^{fg}	128.027 ^a	32.42 ^b
		b	0.2350 ^{defgh}	0.1593 ^{ghij}	0.2843 ^{ab}	2.1876 ^{bcd}	0.8413 ^a	0.5850 ^a	0.0530 ^{hij}	0.2600 ^f	128.18 ^a	32.90 ^b
	10	a	0.2280 ^{defgh}	0.1700 ^{efghi}	0.2830 ^{ab}	2.0506 ^{de}	0.4800 ^{ij}	0.3480 ^{cd}	0.0270 ^{kl}	0.5200 ^a	112.036 ^d	28.24 ^c
		b	0.2730 ^{bc}	0.1737 ^{efgh}	0.2970 ^a	2.3250 ^{ab}	0.6247 ^{efg}	0.3723 ^{cd}	0.0340 ^{ijkl}	0.5257 ^a	112.386 ^d	28.68 ^d
	15	a	0.2316 ^{defgh}	0.1260 ^k	0.2407 ^{fghi}	1.4130 ^{ghi}	0.5467 ^{ghi}	0.2768 ^c	0.1723 ^d	0.2233 ^{gh}	86.817 ^j	21.66 ^{lm}
		b	0.2543 ^{cdef}	0.1510 ^{ghijk}	0.2253 ^{ij}	1.4616 ^{gh}	0.5767 ^{fgh}	0.2757 ^{ef}	0.1750 ^d	0.2293 ^{fgh}	87.453 ^j	21.97 ^l
	50	a	0.2290 ^{defgh}	0.2073 ^{bcd}	0.2012 ^k	2.1896 ^{bcd}	0.5088 ^{hi}	0.3482 ^{cd}	0.0813 ^{fg}	0.2413 ^{fgh}	106.228 ^f	26.97 ^g
		b	0.2336 ^{defgh}	0.2357 ^{ab}	0.2107 ^{jk}	2.2890 ^{abc}	0.5527 ^{ghi}	0.3690 ^{cd}	0.0823 ^{fg}	0.2388 ^{fgh}	106.446 ^{ef}	27.73 ^f
	5	a	0.2173 ^{fgh}	0.1610 ^{ghij}	0.2703 ^{bcd}	2.0473 ^{de}	0.5333 ^{ghi}	0.1877 ^j	0.0418 ^{ijk}	0.4410 ^b	107.205 ^{ef}	26.99 ^g
		b	0.2223 ^{efgh}	0.1487 ^{ghijk}	0.2813 ^{ab}	2.0650 ^{de}	0.6163 ^{efg}	0.2017 ^{ij}	0.0495 ^{hij}	0.4420 ^b	107.133 ^{ef}	27.15 ^g

IA: Irrigation Application; FA: Foliar Application; SS: Seasons; *: ppm; The means followed by same letters are not statistically significant at $P < 0.05$.

Copper is essential for cellular metabolism (Ivanova *et al.*, 2010). Mn is essential mineral and intervening in several metabolic processes, mainly in photosynthesis and as an enzyme antioxidant-cofactor (Millaleo *et al.*, 2010). Hu *et al.* (2015) also reported that Se application can improve Mn and Zn, Ni, Ca and other minerals in the plants. Arvy (1992) reported that Se when applied on moderate level affects Mn, Co, Zn, P and Mo. Zinc is present in many proteins so important for cellular metabolism (Ivanova *et al.*, 2010). Selenium can increase uptake of Zn when applied at moderate level (Fu *et al.*, 2011). Iron plays an important role in redox reaction and electron transport chain. It is also important for photosynthesis and respiration (Kim and Gueriot, 2007). Li *et al.* (2018) reported that when Se is applied to the plants, it increases the concentration of iron. He *et al.* (2007) also reported that application of Se in lettuce had increased mineral content. Soil pH, acidity, salinity and related factors

affect the mineral availability to the plants.

He *et al.* (2007) worked on application of Se in lettuce and Ying *et al.* (2014) worked on rice who reported that application of Se can decrease Pb concentration. Qing *et al.* (2015) reported that Se application in foliar form decrease chromium concentration. They also reported that Se can detoxify Cr by minimizing super oxide free radicals that are produced by Cr in leaves. Selenium helps in plant growth. It delays leaf senescence. It increases oxidative stress in plant cause due to UV light Germ *et al.* (2007). Application of Se can increase Se concentration in tomato fruit (Nancy and Indra, 2014). Lee *et al.* (2007) also found the same results. Smith and Watkinson (1984) also found that application of Se can increase Se in tomato. Se is essential for plants because it helps in holding together cell walls and have structural role in cell wall and membrane (White *et al.*, 2003).

Table 3: Proximate composition (%) of tomato hybrid Salar F1 fruits as affected by Selenium application (mg/Kg) in irrigation and foliar spray in two seasons.

IA	FA	SS	C. Fat	C. Prtn	Ash	C Fibr	Moisture	Lycopene
1	1	a	0.19 ^{ghi}	0.65 ^{ghi}	1.13 ^{abcde}	1.64 ⁱ	93.083 ^k	2.82 ^b
		b	0.22 ^{efg}	0.67 ^{fghi}	1.16 ^{ab}	1.8333 ^{bcd}	93.127 ^{jk}	2.3567 ^{mn}
	2	a	0.14 ^j	0.68 ^{efghi}	1.01 ^{ghijkl}	1.56 ^j	93.15 ^{ijk}	2.6833 ^c
		b	0.1667 ^{ij}	0.7133 ^{bcdef}	1.0667 ^{cdefgh}	1.6333 ⁱ	93.167 ^{ghij}	2.4233 ^{ijkl}
	3	a	0.2167 ^{efgh}	0.72 ^{abcdef}	1.09 ^{bcdefg}	1.76 ^{efg}	93.247 ^{cdef}	2.5733 ^d
		b	0.25 ^{cde}	0.7533 ^{abc}	1.14 ^{abc}	1.8233 ^{bcde}	93.283 ^{abcd}	2.4867 ^{fghi}
	4	a	0.24 ^{cdef}	0.6267 ⁱ	0.9767 ^{ijklm}	1.75 ^{fg}	93.157 ^{hij}	2.84 ^b
		b	0.2767 ^{bc}	0.6633 ^{fghi}	1.0633 ^{cdefgh}	1.8033 ^{cdef}	93.193 ^{fghij}	2.4167 ^{klm}
2	1	a	0.18 ^{hi}	0.6833 ^{defghi}	0.86 ^p	1.6833 ^{hi}	93.223 ^{defgh}	2.7333 ^c
		b	0.2167 ^{efgh}	0.7167 ^{bcdef}	0.95 ^{lmn}	1.72 ^{gh}	93.273 ^{abcd}	2.35 ⁿ
	2	a	0.2767 ^{bc}	0.65 ^{ghi}	1.05 ^{efghij}	1.8367 ^{bcd}	93.17 ^{ghij}	2.4967 ^{efg}
		b	0.3 ^{ab}	0.6633 ^{fghi}	1.0933 ^{bcdef}	1.88 ^{ab}	93.217 ^{defghi}	2.3267 ⁿ
	3	a	0.21 ^{fgh}	0.74 ^{abcd}	1.1967 ^a	1.7733 ^{defg}	93.257 ^{cdef}	2.8 ^b
		b	0.2367 ^{def}	0.7567 ^{ab}	1.1333 ^{abcd}	1.7967 ^{cdef}	93.303 ^{abc}	2.43 ^{hijk}
	4	a	0.2967 ^{ab}	0.6967 ^{cdefgh}	1.0433 ^{fghijk}	1.65 ⁱ	93.123 ^{jk}	2.92 ^a
		b	0.3267 ^a	0.72 ^{abcdef}	1.1067 ^{bcdef}	1.7233 ^{gh}	93.167 ^{ghij}	2.49 ^{efgh}
3	1	a	0.1867 ^{ghi}	0.7133 ^{bcdef}	0.92 ^{mnp}	1.8733 ^{abc}	93.2 ^{efghi}	2.9467 ^a
		b	0.21 ^{fgh}	0.7333 ^{abc}	0.9967 ^{hijklm}	1.91 ^a	93.227 ^{defgh}	2.3633 ^{lmn}
	2	a	0.2733 ^{bcd}	0.7433 ^{abc}	0.8633 ^{op}	1.7967 ^{cdef}	93.167 ^{ghij}	2.5467 ^{def}
		b	0.31 ^{ab}	0.7767 ^a	0.9433 ^{lmno}	1.8533 ^{abc}	93.23 ^{defg}	2.4567 ^{ghij}
	3	a	0.22 ^{efg}	0.6433 ^{hi}	0.9633 ^{klm}	1.6333 ⁱ	93.267 ^{bcde}	2.7233 ^c
		b	0.2533 ^{cde}	0.6667 ^{fghi}	1.0567 ^{defghi}	1.68 ^{hi}	93.337 ^{ab}	2.46 ^{ghij}
	4	a	0.21 ^{fgh}	0.6767 ^{efghi}	0.88 ^{nop}	1.8533 ^{abc}	93.283 ^{abcd}	2.5533 ^{de}
		b	0.2433 ^{cdef}	0.7033 ^{bcdefg}	0.97 ^{ijklm}	1.88 ^{ab}	93.343 ^a	2.3667 ^{klmn}

IA: Irrigation Application; FA: Foliar Application; SS: Seasons; The means followed by same letters are not statistically significant at $P < 0.05$.

It helps to decrease leaf senescence also plays an important role in photosynthesis and nucleic acid synthesis. Increasing Se application can increase Mg concentration (Kopsell *et al.*, 2000).

Table 3 showed the effect of selenium application on proximate composition of tomato fruits. It was examined that crude fat ranged from 0.14 to 0.3267%, crude protein from 0.6267 to 0.7767%, ash from 0.86 to 1.1967%, crude fiber from 1.56 to 1.91%, moisture 93.083 to 93.343 % and lycopene 2.3567 to 2.9467 $\mu\text{g/g}$. Factors and interaction showed their effect on these proximate composition of fruits. Crude fat, ash and moisture were significantly affected by all the three factors and by the interaction of foliar and irrigation applications. Crude protein was affected by season and by the interaction of foliar and irrigation. Crude fiber was affected by irrigation, season, interaction of foliar and irrigation and by the interaction of irrigation and season. Lycopene was affected by foliar and season

applications and by interaction all the three factors.

Table 4 showed the effect of selenium application on proximate composition of tomato leaves which showed that ash 3.1933 to 3.85%, crude fiber 5.03 to 5.2733%, crude fat 0.0296 to 0.0309%, crude protein 1.0967 to 1.24%, moisture from 92.083 to 92.317%, chlorophyll before flowering 0.024 to 0.0309 $\mu\text{g/g}$, chlorophyll after flowering 0.0352 to 0.043 $\mu\text{g/g}$. The factors effect on these parameters of tomato leaves showed that crude fat, ash, chlorophyll before and after flowing was significantly affected by all the three factors i.e. foliar and irrigation application and season and by the interaction of Foliar and irrigation. However, the chlorophyll of leaves after flowing was additionally affected by irrigation and season interaction and by overall interaction of the three factors. Crude proteins were affected by season and moisture by irrigation. These both parameters were also affected by the interaction of foliar and irrigation factors.

Table 4: Proximate composition (%) of tomato hybrid Salar F1 leaves as affected by Selenium application (mg/Kg) in irrigation and foliar spray in two seasons.

IA	FA	SS	C. Fat	C. Prtn	Ash	C. Fibr	Moisture	Chlor before flowers	Chlor after flowers
1	1	a	1.7333 ^{cde}	1.0967 ^{hi}	3.24 ^{hijk}	5.1067 ^{ab}	92.25 ^{ab}	0.0251 ^{kl}	0.0424 ^a
		b	1.78 ^{bcd}	1.14 ^{efgh}	3.4667 ^{cdefg}	5.1433 ^a	92.093 ^{cd}	0.024 ^{lm}	0.0382 ^{ef}
	2	a	1.77 ^{cd}	1.1533 ^{defg}	3.2933 ^{ghij}	5.2567 ^a	92.083 ^d	0.0322 ^{bc}	0.0391 ^{de}
		b	1.8067 ^{bc}	1.19 ^{abcde}	3.3867 ^{defgh}	5.2867 ^a	92.153 ^{bcd}	0.0314 ^{cd}	0.0362 ^{hij}
	3	a	1.9267 ^a	1.1133 ^{ghi}	3.7733 ^a	5.03 ^{ab}	92.18 ^{abcd}	0.0347 ^a	0.0382 ^{ef}
		b	1.9567 ^a	1.16 ^{defg}	3.81 ^a	5.0733 ^{ab}	92.213 ^{abcd}	0.0336 ^{ab}	0.0352 ^{jk}
	4	a	1.47 ⁱ	1.1733 ^{cde}	3.5867 ^{bc}	5.1333 ^{ab}	92.227 ^{abc}	0.0318 ^{cd}	0.0411 ^{bc}
		b	1.4967 ^{hi}	1.2233 ^{ab}	3.6933 ^{ab}	5.1767 ^a	92.267 ^{ab}	0.0309 ^{cde}	0.0377 ^{fg}
2	1	a	0.0296 ^{efg}	1.1533 ^{defg}	3.1933 ^{ijk}	5.2767 ^a	92.2 ^{abcd}	0.0296 ^{efg}	0.0385 ^{def}
		b	0.0288 ^{gh}	1.2033 ^{abcd}	3.2733 ^{hij}	5.02 ^{ab}	92.24 ^{ab}	0.0288 ^{gh}	0.0366 ^{gh}
	2	a	0.0253 ^{kl}	1.08 ⁱ	3.1367 ^{jk}	5.11 ^{ab}	92.193 ^{abcd}	0.0253 ^{kl}	0.0413 ^{bc}
		b	0.0236 ^m	1.1533 ^{defg}	3.2067 ^{ijk}	5.1633 ^a	92.243 ^{ab}	0.0236 ^m	0.0348 ^k
	3	a	0.0249 ^{klm}	1.1167 ^{fghi}	3.79 ^a	5.19 ^a	92.26 ^{ab}	0.0249 ^{klm}	0.0421 ^{ab}
		b	0.0239 ^{lm}	1.1733 ^{cde}	3.85 ^a	5.2333 ^a	92.293 ^a	0.0239 ^{lm}	0.0363 ^{hi}
	4	a	0.0286 ^{ghi}	1.17 ^{cdef}	3.2467 ^{hijk}	5.26 ^a	92.183 ^{abcd}	0.0286 ^{ghi}	0.043 ^a
		b	0.0280 ^{hi}	1.2333 ^{ab}	3.3133 ^{efghi}	5.0133 ^{ab}	92.237 ^{ab}	0.0280 ^{hi}	0.0391 ^{de}
3	1	a	0.0272 ^{ij}	1.1833 ^{bcde}	3.46 ^{cdef}	5.1633 ^a	92.277 ^{ab}	0.0272 ^{ij}	0.0410 ^c
		b	0.0257 ^{jk}	1.24 ^a	3.53 ^{bcd}	5.2467 ^a	92.317 ^a	0.0257 ^{jk}	0.037 ^{gh}
	2	a	0.0309 ^{cde}	1.1433 ^{efgh}	3.0967 ^k	4.7067 ^b	92.267 ^{ab}	0.0309 ^{cde}	0.0394 ^d
		b	0.0299 ^{efg}	1.19 ^{abcde}	3.1767 ^{ijk}	5.12 ^{ab}	92.303 ^a	0.0299 ^{efg}	0.0384 ^{def}
	3	a	0.0288 ^{gh}	1.1467 ^{efgh}	3.2433 ^{hijk}	5.14 ^a	92.18 ^{abcd}	0.0288 ^{gh}	0.0375 ^{fg}
		b	0.0279 ^{hi}	1.19 ^{abcde}	3.29 ^{ghij}	5.1567 ^a	92.203 ^{abcd}	0.0279 ^{hi}	0.036 ^{hij}
	4	a	0.0304 ^{def}	1.0933 ^{hi}	3.4467 ^{cdefg}	5.2233 ^a	92.23 ^{abc}	0.0304 ^{def}	0.0385 ^{def}
		b	0.0292 ^{fgh}	1.1433 ^{efgh}	3.5333 ^{bcd}	5.2733 ^a	92.237 ^{ab}	0.0292 ^{fgh}	0.0355 ^{ijk}

IA: Irrigation Application; FA: Foliar Application; SS: Seasons; The means followed by same letters are not statistically significant at $P < 0.05$.

Table 5: Effect of foliar, irrigation applications and Season and their interaction on chemical parameters of tomato hybrid Salar F1 Leaves.

Factors and Interactions	Hieght/ p	Leaves/P	Leaflets/P	Buds/P	Flowers/P	Fruits/P
Foliar	Sig.		Sig.		Sig.	
Irrigation	Sig.		Sig.	Sig.		
Season		Sig.	Sig.	Sig.	Sig.	Sig.
Foliar x Irrigation	Sig.	Sig.	Sig.	Sig.	Sig.	
Foliar x Season	Sig.		Sig.			
Irrigation x Season			Sig.	Sig.	Sig.	
Foliar x Irrigation x Season	Sig.		Sig.			Sig.

* Significant at 5% probability ($P < 0.05$).

Table 6: Effect of foliar, irrigation applications and Season and their interaction on mineral content of tomato hybrid Salar F1.

Factors and Interactions	Ca	Cr	Cu	Fe	Mg	Mn	Ni	Pb	Se	Zn
Foliar	Sig.*	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.
Irrigation	Sig.	Sig.		Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.
Season		Sig.		Sig.	Sig.		Sig.	Sig.		Sig.
Foliar x Irrigation	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.
Foliar x Season					Sig.					
Irrigation x Season										
Foliar x Irrigation x Season										

* Significant at 5% probability ($P < 0.05$).

Table 7: Effect of foliar, irrigation applications and Season and their interaction on chemical parameters of tomato hybrid Salar F1 fruits.

Factors and Interactions	Fruit C.Fat	FC. Prtn	Fruit Ash	Fruit Fibr	Moisture	Lycopene
Foliar	Sig.		Sig.		Sig.	Sig.
Irrigation	Sig.		Sig.	Sig.	Sig.	
Season	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.
Foliar x Irrigation	Sig.	Sig.	Sig.	Sig.	Sig.	Sig.
Foliar x Season						Sig.
Irrigation x Season				Sig.		Sig.
Foliar x Irrigation x Season						Sig.

* Significant at 5% probability ($P < 0.05$).

Table 8: Effect of foliar, irrigation applications and Season and their interaction on chemical parameters of tomato hybrid Salar F1 leaves.

Factors and Interactions	Leaves C. Fat	Leaves C. Prtn	Leaves ash	Leaves C. Fibr	Leaves Moisture	Chlor before flowers	Chlor after flowers
Foliar	Sig.		Sig.			Sig.	Sig.
Irrigation	Sig.		Sig.		Sig.	Sig.	Sig.
Season	Sig.	Sig.	Sig.			Sig.	Sig.
Foliar x Irrigation	Sig.	Sig.	Sig.		Sig.	Sig.	Sig.
Foliar x Season							
Irrigation x Season							Sig.
Foliar x Irrigation x Season							Sig.

* Significant at 5% probability ($P < 0.05$).

Results of [Abdullahi et al. \(2016\)](#) were similar with the present study. Crude fat provides energy for metabolic processes structural component in membrane and important in intracellular signals. [Song et al. \(2015\)](#) reported and suggested that selenium application at optimum level can increase crude fat by increasing linoleic acid and sterol. [Fernando et al. \(2018\)](#) and [Zhu et al. \(2017\)](#) also reported the same result. [Hu et al. \(2003\)](#) suggested that crude protein content might be increased due to increase in different amino acid. [Ježek et al. \(2011\)](#) studied that application of Se can increase total amino acid when applied at flowering stage of Chrysanthemum. They further reported that when Se is sprayed on potato it would increase phenylalanine. The work done by [Nancy and Arulselvi \(2014\)](#) also agreed with the present study who reported that application of Se increases minerals content so in turn increased ash content. [Hu et al. \(2015\)](#) also agreed with Chen and Arvy study. So in variation or increase in minerals may result in the increase or decrease of ash content. [Fu et al. \(2011\)](#) also reported similar result. [Turakainen et al. \(2004\)](#) reported that selenium application can increase starch content in potato edible part. So it may be possible

that Se application in tomato also increases fruit fiber which is a part of carbohydrate. [Godina et al. \(2016\)](#) reported that Selenium application has positive effect on moisture of leaves. All these study support the present data. [Godina et al. \(2016\)](#) also reported that application of Se on tomato plant improves the content of % total dry matter (TDM) in fruits. [Zhu et al. \(2017\)](#) reported that application of Se can increase total polysaccharide. [Turakainen et al. \(2004\)](#) reported that selenium application can improve total starches in potato leaves. So Selenium might cause increase in crude fiber content of tomato leaves. [Xue et al. \(2001\)](#) also reported that Se increase chlorophyll content at low concentration. [Pennanen et al. \(2002\)](#) also agreed with the present study and reported that Se protect Chloroplast enzymes and hence increase biosynthesis of photosynthetic pigments.

Lycopene affect the antioxidant activity of tomato ([Chang and Liu, 2008](#)). Lycopene act as antioxidant and promote decrease in DNA damage ([Yildiz and Baysal, 2007](#)). Results of the present study were contradicted with those reported by [Pezzarossa et al. \(2013\)](#). The reason may be the concentration and way

of application of Se. Lee *et al.* (2007) also reported that lycopene was increased in ripe fruit of tomato after selenium application. Secondary metabolism and composition may be affected by selenium accumulation in fruit at ripening. Lycopene content increases because carotenoid biosynthetic path way is affected by Se accumulation. Sams *et al.* (2011) reported that when *Arabidopsis* was treated with Se, it regulated a key step in the carotenoid biosynthesis i.e. phytoene synthetase. However, in tomato fruit, Se may affect other carotenoid genes and enzymes (Sams *et al.*, 2011).

Conclusions and Recommendations

Selenium affected most of the growth parameters of tomato plant including height, leaves, flowers and buds with its moderate level application through irrigation. Proximate composition was also affected by selenium application with its moderate level; However here the foliar application was more effective than irrigation, in relation to the effect of season. Selenium increased most of the minerals, including Cu, Zn and Se etc. By comparing irrigation versus foliar application of the selenium, the irrigation application proved to be more efficient. Selenium should be applied in moderate amount. Application should be conducted mainly through irrigation because it is more efficient than foliar application. Selenium also have adverse effect on chlorophyll concentration that may cause unhealthy plant growth, so should be avoided in higher doses. Se effect should be checked on other cultivars or hybrids for further investigation

Novelty Statement

Novelty of this research is the addition of selenium in moderate amount to tomato hybrid Salar F1 for their physical well-being and improvement of some chemical constituents.

Author's Contribution

Sadaqat Khan: Conducted the research.

Saleem Ullah: Supervised the whole study.

Muhammad Sajid: Monitored the experiments.

Conflict of interest

The authors have declared no conflict of interest.

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