Research Article



Ovarian and Hormonal Responses of Local Rabbits After Superovulation Induction with Bovine Pituitary Extract

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Abstract | Superovulation using gonadotropin hormones are characterized by several limitations, leading to a wide variation in the resultant response. To overcome these limitations, several studies have recommended the use of bovine pituitary extract (BPE) as an effective alternative in improving animal reproduction. Therefore, this study aims to determine the ovarian and hormonal responses of local rabbits after superovulation induction with BPE. The sample comprised of 1 local male rabbit and 9 local female rabbits with a history of giving birth and weighing between 1.8-2.2 kg. The animals were adapted for 30 days in separate cages and provided with feed and water ad libitum before treatment. The rabbits were divided into 3 treatment groups (n=3), the first group as a control group (R1) was injected with physiological NaCl, while group two (R2) and group three (R3) were injected with FSH and BPE, respectively. The total volume of NaCl in R1 and BPE in R3 was 2.6 ml each while the total dose in R3 was 28 mg. Intramuscularly injections in both groups were performed 5 times with a 12-hour interval. The administration of NaCl in R1 and BPE in R3 was 1; 0.5+0.5; 0.3+0.3 mL, respectively while the injection of FSH in R2 was 4; 4+4; 8+8 mg. Subsequently, rabbits in R2 and R3 were injected with 100 IU hCG and mated with males, while those in R1 were mated without hCG administration 24 hours after the last FSH and BPE treatment. On the 6th day after treatment, rabbits in R1, R2, and R3 were slaughtered for ovarian organ retrieval, while blood samples for progesterone, estrogen, LH, and FSH hormone concentration examination were collected on days 1, 3, and 5 after mating. The hormone concentration was analyzed using the enzyme-linked immunoassay (ELISA) method, and the data obtained were analyzed using analysis of variance. The macroscopic observation results showed that the ovaries of rabbits in R2 and R3 were larger than those in R1. Overall, the progesterone concentration in R3 was higher compared to R1 and R2 (P<0.05), and the levels in R2 were higher than R1 (P<0.05). In addition, the number of implantation site showed a significant difference between R1 with R2 and R3 (P<0.05). The concentration of estrogen in R3 increased on day 5 after mating, while FSH and LH showed no significant differences between all groups on days 1, 3, and 5 after mating. Based on these findings, the administration of BPE could be considered as an alternative superovulation preparation in local rabbits in terms of ovarian and hormonal positive responses.

Keywords | Bovine pituitary extract, Hormonal, Induction, Local rabbits, Ovarian, Superovulation

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Pregnant Mare Serum Gonadotropin (PMSG) and Follicle-Stimulating Hormone (FSH) are hormones that play an essential role in superovulation induction, each offering distinct advantages and disadvantages. During experimental animal superovulation, PMSG is widely used, but its prolonged half-life often leads to residual presence in the bloodstream for several days after treatment (Wang et al., 2015). Several studies have shown that this extended half-life significantly influences variations in superovulation responses and the persistence of follicles within the ovaries. This condition often leads to hormonal imbalance and renders the embryos unsuitable for transfer (Afriani et al., 2020). Meanwhile, FSH typically induces better ovarian responses than PMSG due to its ability to produce a greater number of ovulated ova, fewer anovulatory follicles, and more high-quality embryos. Despite its advantages, a notable limitation lies in the susceptibility of FSH to adverse effects when administered in improper or excessively high doses, particularly affecting follicle development (Karl et al., 2021).

The administration of FSH typically comprises a twicedaily regimen spanning 3-4 days (Demoustier *et al.*, 1988; Menchaca *et al.*, 2007). However, this regimen can induce stress in animals and elevate the overall cost of superovulation implementation. Previous reports on rabbit superovulation using the hormone have shown several variations in terms of dose and injection frequency. For instance, Zhang *et al.* (2017) used FSH at dose of 30 IU/ rabbit which was administered through 6 injections for 3 consecutive days. Kauffman *et al.* (1998) stated that FSH administration for 4 days in superovulation could enhance the efficiency of reproductive biotechnology after embryo cryopreservation. Meanwhile, Techakumphu *et al.* (2002) reported that FSH administration was performed only 5 times with a 12-hour interval.

To overcome the limitations associated with PMSG and FSH, several studies have explored alternative substances, with pituitary extract emerging as a viable option. Bovine Pituitary Extract (BPE) is a rich source of hormones, containing at least 10 bioactive substances, including FSH, Luteinizing Hormone (LH), Growth Hormone (GH), Luteotropic Hormone (LTH), Adrenal Corticotropic Hormone (ACTH), Thyroid Somatotropic Hormone (TSH), and Lipogenic hormone, all produced by the anterior pituitary lobe. Recent reports have shown the potential of pituitary extract in inducing estrus in dairy cows and enhancing reproductive efficiency in goats (Siregar et al., 2013). Kaligesing goats injected with pituitary extract intramuscularly exhibited early estrus behaviors, suggesting its potential to improve estrus behavior performance in Etawah goats (Setiawan

et al., 2019). Moreover, Hafizuddin *et al.* (2010) showed that it had similar effectiveness as PMSG in inducing superovulation in mice. The use of pituitary extract has been reported to present several advantages over synthetic hormones due to its ability to produce gonadotropins, such as FSH, LH, and GH, which play a role in fetal growth and maternal milk production. The natural gonadotropins and GH in pituitary extract do exert negative impacts on the reproductive performance of female donor animals (Nalley *et al.*, 2017).

The use of pituitary extract for superovulation purposes remains limited (Arum et al., 2013; Sayuti et al., 2022), with precise methods and doses yet to be firmly established. In a study by Sayuti et al. (2022), BPE was administered to rabbits once daily for 3 days, with decreasing doses of 1, 0.5, and 0.3 ml. The results showed that the number of fetuses as well as the concentration of estrogen and progesterone showed no significant differences compared to the control group. The study also suggested that BPE administration tends to increase progesterone concentration in local rabbits, showing a hormonal increase compared to the control. On day 7 after mating, the progesterone levels in BPE-injected and control groups were 10.00±5.30 and 3.72±1.04 ng/ mL, while the values on day 14 were 11.95±5.38 and 1.01±0.65 ng/ml, respectively. The progesterone increase can be attributed to the effect of the increased ovulation caused by BPE. Siregar et al. (2020) also reported that BPE administration in white rats could enhance folliculogenesis activity, as showed by an increase in the number of follicles and estrogen concentration. Based on these findings, it was presumed that its ability in superovulation induction was similar to FSH. Therefore, treatment protocols must follow the FSH administration pattern.

The success of superovulation can be showed by an increased number of offspring (Nur et al., 2016), the concentration of estrogen and progesterone hormones (Amiruddin et al., 2013), and an increase in the number of corpus luteum (CL) (Siregar et al., 2020). Several studies have shown that ovulation and superovulation induction responses in rabbits can increase FSH and LH levels. FSH concentration has been reported to fluctuate more than LH after induction. In New Zealand White (NZW) rabbits induced ovulation with 50 µg hCG, the concentration of the hormone was 6.0±1.6 ng/ml and then decreased to pre-mating levels 72 hours after mating (Mills et al., 1981). Hashimoto et al. (2007) reported that the FSH peak concentration in rabbits super ovulated occurred 50 hours after the start of treatment. LH levels in GnRHsuper ovulated rabbits increased significantly compared to the controls. Furthermore, the concentration peaked at 24 hours after treatment and then gradually decreased until day 6 (Cervantes et al., 2015). This study aims to determine the ovarian and hormonal responses of local rabbits after

superovulation induction with BPE.

MATERIALS AND METHODS

This study was conducted after approval from the ethics committee of Faculty of Veterinary Medicine, Universitas Syiah Kuala, Banda Aceh, Indonesia with certificate No. 139/KEPH/VII/2020. A total of 9 local female rabbits that had previously given birth with a weight of 2-3 kg, and 1 local male rabbit were used in this study. The animals were adapted for 30 days in separate cages (Kanayama *et al.*, 1995) and provided with feed and water ad libitum. Subsequently, the rabbits were divided into 3 treatment groups (n=3). The rabbit in group 1 (R1) was injected with 2.6 mL physiological NaCl, while rabbit in group 2 (R2) and 3 (R3) were injected with 28 mg FSH and 2.6 mL BPE. The intramuscularly injections were performed 5 times with a 12-hour interval. The preparation of BPE was carried out based on the method by Isnaini *et al.* (1999).

SUPEROVULATION

The initial injection of physiological NaCl in groups R1, FSH in R2, and BPE in R3 was conducted at 08:00 PM and repeated every 12 hours until the 5th injection. In group R2, each injection comprised FSH with doses of 4 mg (1st injection), 4 mg (2nd injection), 4 mg (3rd injection), 8 mg (4th injection), and 8 mg (5th injection) (Techakumphu *et al.*, 2002). In group R3, each injection consisted of BPE with doses of 1 ml (1st injection), 0.5 ml (2nd injection), 0.5 ml (3rd injection), 0.3 ml (4th injection), and 0.3 ml (5th injection) (Sayuti *et al.*, 2022). Rabbits in R2 and R3 were then injected with 100 IU hCG and mated with males, while those in R1 were mated without hCG injection, 24 hours after the last FSH and BPE injections.

OVARIAN COLLECTION

On day 6 after treatment, rabbits in R1, R2, and R3 were slaughtered for ovarian organ retrieval. Dexter et sinister ovaries were then prepared for macroscopic morphological observations. Morphological observations focused on the superficial part to identify follicle and CL types, as well as to count the number of follicles and CL in the dexter et sinister ovaries.

BLOOD SAMPLING AND HORMONAL DETERMINATION

Blood samples for examining the concentration of progesterone, estrogen, LH, and FSH were collected on days 1, 3, and 5 after mating at the same time. Rabbit blood was collected using a 1 ml disposable syringe through the auricular vein. The blood was collected in a 3 ml EDTA vacutainer and centrifuged for 15 minutes at 3500 rpm. The plasma was then transferred to coded microtubes and stored in a -20 °C freezer until examination.

Hormone concentration analysis was performed using ELISA method. The measurement procedures for progesterone and estrogen concentration followed the ELISA kit manual (DRG, International Inc., Germany), while the concentration of FSH and LH was determined using the ELISA kit manual (Bioenzy, Indonesia). The P4 kit analytical sensitivity was 0.045 ng/mL and the variance of coefficients for intra-assay and inter-assay were $\leq 5.43\%$ and $\leq 4.3\%$, respectively. For the E2 kit, the limit of detection was 9.714 pg/mL, while the intra-assay and inter-assay were $\leq 2.71\%$ and $\leq 6.72\%$, respectively. The FSH and LH kit analytical sensitivity was 0.12 mIU/mL, while the variance of coefficients for intra-assay and inter-assay and inter-assay were $\leq 8\%$ and $\leq 10\%$, respectively.

DATA ANALYSIS

The data obtained in this study were analyzed using oneway analysis of variance (ANOVA), followed by Duncan's test.

RESULTS AND DISCUSSION

MACROSCOPIC OBSERVATIONS OF OVARIAN RESPONSES IN LOCAL RABBITS

Macroscopic observations of ovarian responses in the 3 rabbit groups focused on the changes in the superficial part of the ovaries, including follicles at various stages and CL. Based on the observations on the rabbits ovaries in R1, R2, and R3 showed differences in morphology as presented in Figure 1.

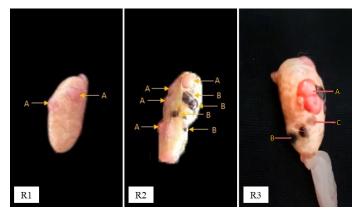


Figure 1: Morphology of the ovaries in local female rabbits (R1, physiological NaCl; R2, FSH; R3, Bovine pituitary extract; A, corpus luteum; B, corpus haemorrhagicum; C, follicle).

Macroscopically, the ovaries of rabbits in R2 and R3 were larger compared to R1. This size difference was attributed to the abundance of follicles and CL induced by superovulation with FSH and BPE. The formed follicles included primary, secondary, and tertiary (antral) variants, with CL and corpus haemorrhagicum (CH) observed among them. The macroscopic calculation results for the

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number of follicles and CL on the surface of the dexter et sinister ovaries in both groups are presented in Table 1.

Table 1: The number of follicles and CL observedmacroscopically in local rabbits induced with physiologicalNaCl and BPE.

Measurement	Treatment			
parameters	R1 (Phys- iological NaCl)	R2 (FSH)	R3 (BPE)	
Number of follicles	13.00±1.00ª	48.23±19.54 ^b	44.33 ± 28.57^{b}	
Number of CL	6.00±5.29ª	18.7 ± 7.8^{b}	21.00 ± 3.00^{b}	
Number of implantation site	3.21±1.34ª	11.7±4.7 ^b	12.3±3.5 ^b	

^{a,b}Different superscript letters in the same row showed significant differences (P < 0.05).

The number of implantation site showed a significant difference in groups R1, R2 and R3 (P<0.05). This was consistent with the significant differences in the number of CL, as presented in Table 1. The decrease in the number of implantation site was also consistent with the decrease in ovulation. Several studies showed that this could be due to the failure of fertilization in the ovulated oocytes. Satheskumar (2006) reported a decrease in the number of fertilized oocytes compared to the number of ovulations. In NZW rabbits, the number of ovulations was 22.0±1.4, while the number of fertilized oocytes was only 6.8±3.1.

Based on the data in Table 2, there was a significant difference (P<0.05) in progesterone concentration among R1, R2, and R3. The highest progesterone concentration on days 1, 3, and 5 was found in R3. Furthermore, the

estrogen level in R1 and R2 tended to remain stable, while that of R3 tended to increase by days, as shown in Table 2.

Table 2 showed that the estrogen levels in R1 and R2 tended to be lower R3 (BPE injection) on days 1 and 3 after mating, even not significantly different (P>0.05). However, estrogen levels in group R1 and R2 on day 5 exhibited a significant difference (P<0.05) compared to R3. The levels of FSH and LH which were measured in all groups on days 1, 3, and 5 after mating did not differ significantly (P > 0.05).

OVARIAN RESPONSES

The results showed that superovulation induction using BPE could enhance ovarian follicle development in R2 and R3. Furthermore, the macroscopically observed follicles included small (secondary) and large (tertiary or antral) ovarian follicles that had not undergone ovulation. This finding was consistent with O'Callaghan *et al.* (2000) that superovulation induction with FSH in sheep increased the number of follicles compared to the control, with values of 12.3±1.4 and 3.5±0.3, respectively. The number of CL in R2 was higher than in R1, and this result was consistent with Ju *et al.* (2001) and Marhaeniyanto and Khastama (2007) who obtained similar results.

Ju *et al.* (2001) reported an increase in the average number of ovulations in rabbits injected with FSH superovulation compared to the control, with respective values of 37.2 ± 16.1 and 10.4 ± 2.6 . Marhaeniyanto and Khastama (2007) also showed an increase in the number of CL in rabbits injected with a combination of PMSG and PGF2 α compared to the control, with respective values of 9.00 ± 1.15 and 6.25 ± 1.95 .

Table 2: The progesterone, estrogen, FSH, and LH concentration of local rabbits on days 1, 3, and 5 after mating and superovulation induction with BPE.

Parameters	Treatments	Hormone concentration on the day after mating		
		1	3	5
Progesterone (ng/mL)	R1 (physiological NaCl)	0.56±0.21ª	2.83±0.99ª	2.25±0.24ª
	R2 (FSH)	3.63 ± 0.83^{b}	8.49 ± 5.60^{b}	18.23±14.01 ^b
	R3 (BPE)	5.90±2.11°	13.15±2.37°	36.66±3.89°
Estrogen (pg/mL)	R1 (physiological NaCl)	18.80±4.36ª	18.00±4.19ª	23.53±5.09ª
	R2 (FSH)	22.71±1.47ª	21.59 ± 2.84^{a}	24.19±4.90 ^a
	R3 (BPE)	30.25±7.85ª	38.45±26.45 ^a	151.47±47.77 ^b
FSH (mIU/mL)	R1 (physiological NaCl)	40.66±5.29	45.73±8.12ª	43.52±5.22ª
	R2 (FSH)	38.89±3.96	42.01± 6.79	40.99±4.83
	R3 (BPE)	36.64±5.46 ^a	35.87±10.33ª	25.29±12.46 ^a
LH (mIU/mL)	R1 (physiological NaCl)	40.27±9.74 ^a	40.54±6.13ª	35.95±13.56ª
	R2 (FSH)	33.06±5.55	46.40±12.09	36.54±3.30
	R3 (BPE)	32.73±7.81ª	35.63±7.20ª	22.20±10.76 ^a

BPE: bovine pituitary extract.^{a,b} Different superscript letters in the same column indicated significant differences (P < 0.05).

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HORMONAL RESPONSE

The progesterone concentration in R2 and R3 showed that the rabbits had successfully became pregnant. Szendro *et al.* (2010) also reported that the progesterone concentration in pregnant rabbits was 9.4 ng/ml, while Kleden and Soetatto (2017) showed lower levels at 3.1 ng/ml. In New Zealand white rabbits, levels on days 7, 14, and 21 of pregnancy was 3.16±0.3, 4.00±0.32, and 3.94±0.32 ng/ml, respectively (Ashour and Abdel-Rahman, 2019). In R1, there was also a suspected pregnancy due to the tendency for an increase in progesterone concentration on day 3. Syafruddin *et al.* (2022) reported that the hormone levels in local pregnant rabbits on day 4 after mating was 2.10±1.41 ng/ml.

During the examination period, it was evident that the progesterone concentration in R3 was higher compared to R1 and R2 (P<0.05) and the levels in R2 was higher than R1 (P<0.05), primarily due to the increased number of ovulations. The formation of a higher number of CL led to an increased secretion of the hormone (Sudjatmogo et al., 2001; Adriani et al., 2004). Maertens and Luzi (1995) stated that superovulation caused an increase in the number of ovulating follicles. The rupture of follicles during ovulation formed a CH, which later became the CL. Furthermore, the CL was known as a temporary endocrine organ that predominantly produced progesterone. The levels of this hormone in blood serum were often used to predict the number of CL in domestic animals during the luteal phase (Scaramuzzi et al., 1993). Tjitosumirat (2009) reported that progesterone concentration could be used to predict the number of ovulations in small ruminant animals.

Based on Table 2, there was an increase in progesterone concentration in R2 and R3 over the examination period. The increase in progesterone concentration with time was also reported by Sayuti *et al.* (2022), who used BPE preparation for superovulation induction. The progesterone concentration on days 7 and 14 after mating was 10.00±5.30 and 11.95±5.38 ng/ml, respectively. Challis *et al.* (1973) reported a similar trend where the average levels during pregnancy increased from 5.3 ng/ml on day 3 to 17-19 ng/ml on days 12 to 15.

A relatively high increase in hormone concentration occurred on day 5, due to progression into the functional luteal phase, where the CL began to actively secrete progesterone in large quantities. Wijono and Didi (1998) stated that high levels of progesterone occurred in the pregnancy phase with maximum CL development. The lower progesterone concentration in R1 was believed to be related to the lower number of ovulations compared to R2 and R3. The absence of a linear increase in the hormone's level in R1 could be related to the regression of several CL after mating, which did not develop optimally. Arimbawa *et al.* (2012) stated that progesterone levels increased

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and decreased in line with CL development during the estrous cycle. Another possibility was that the mating was unsuccessful and only led to pseudo-pregnancy. Browning *et al.* (1980) stated that progesterone concentration in local pseudo-pregnant rabbits was lower compared to pregnant rabbits. However, Holt *et al.* (1976) stated that progesterone concentration in pregnant and pseudo-pregnant rabbits did not differ during the implantation period.

The increase in progesterone concentration in R2 and R3 showed that the administration of FSH and BPE as superovulation induction had an effect on increasing ovulation in the samples. The ability of BPE to induce folliculogenesis in R3, leading to the formation of numerous CL compared to R2, suggested that BPE was superior to FSH. This finding was in line with Isnaini et al. (1999) that BPE contained FSH hormone and could be used as a superovulation agent. Several studies also showed that PMSG and BPE had equal effectiveness in inducing twin birth in goats (Siregar *et al.*, 2013; Zulkarnain *et al.*, 2015). According to Sayuti et al. (2022), BPE administration in local rabbits led to a non-significant increase in progesterone concentration compared to the control on day 7 after mating at 10.00±5.30 and 3.72±4.04 ng/ml, respectively, while this current study obtained significant results. The method of administering BPE in this study was considered to be more appropriate compared to that of Sayuti et al. (2022). Possible factors affecting the higher progesterone concentration compared to previous reports included the injection method performed twice a day, the amount of gonadotropin hormone present in BPE, the good physiological condition of the experimental animals, and the addition of hCG, which played a role in controlling ovulation.

Based on Table 1, the estrogen levels in R3 increased on day 5 after mating. This was consistent with Suartini *et al.* (2013) who observed that repeated doses released gonadotropin hormones, leading to an increased number of growing follicles. Ashour and Abdel-Rahman (2019) reported that estrogen concentration in NZW rabbits tended to increase with advancing pregnancy age. The levels observed on days 14, 21, and 28 of pregnancy were 47.31±4.99, 56.33±5.34, and 88.99±5.34 pg/ml, respectively. According to Siregar *et al.* (2020), the increase in the hormone's level was due to the growth of follicles.

The possible increase in estrogen on day 5 in R3 could also be due to the administration of hCG. In addition, human Chorionic Gonadotropin (hCG) had a biological activity similar to LH. Mattioli *et al.* (2021) as cited by Mattioli *et al.* (2021), stated that LH could induce earlier ovulation and increase estrogen secretion. The peak increase in the hormone coincided with the wave of follicle growth (Fortune *et al.*, 1991). The occurrence of increased growth of ovulating

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follicles often led to significant production of estrogen due to the effect of PMSG administration (Maertens and Luzi, 1995). This was consistent with Sugiyatno *et al.* (2001) that the number of ovulating follicles increased the amount of the hormone in the blood serum.

In R3, there was a wide standard deviation that was suspected to affect the non-significance of the statistical test results for estrogen concentration on days 1, 3, and 5. This wide standard deviation showed a large variation among individuals. A possible cause of this observation was that rabbits in R3 came from different (proliferation) strains. In sheep, it had been reported that the effectiveness of PMSG administration was dependent on the fertility level of the strain and the treatment had no impact when given to high-fertility strains (Langford et al., 1983). Lehloenva and Greyling (2010) reported that the age and parity of Boer goats affected ovarian responses after superovulation. Song et al. (2012) also reported that the superovulation responses in Hanwoo cows were determined by their parity. The superovulation responses, marked by a higher embryo yield, were observed more in cows with parity 3-5 compared to parity 1-2 and parity >6. Based on these findings, parity had an effect on the responses to gonadotropin, as there was a difference in ovarian sensitivity to gonadotropin induction at different ages. In this study, the sample criteria used included animals that had previously given birth, without limiting the number of calving periods. The limited number of samples was also suspected to affect the non-significant results.

Sumiyoshi *et al.* (2014) reported that estrogen levels played a crucial role in endocrine control. Furthermore, Lestari and Ismudiono (2014) stated that an increase in estrogen had both positive and negative feedback effects on the hypothalamus, leading to the release of FSH and LH. According to Fortune (1993), the processes of follicle growth, ovulation, and CL formation were affected by the circulation of reproductive hormones in the body. The hypothalamus produced GnRH, which stimulated the anterior pituitary to release FSH and LH in response to estrogen. This suggested that the significant increase in estrogen was likely caused by a positive feedback effect on the hypothalamus in producing GnRH.

The non-significant differences in FSH and LH concentration between R1, R2, and R3 on days 1, 3, and 5 after mating were likely due to inaccuracies in the time of hormone concentration measurements. Blood sampling in this study was performed on days 1, 3, and 5 after mating and it was suspected that FSH and LH concentrations returned to their baseline levels. Mills *et al.* (1981) reported that FSH levels in NZW rabbits induced with 50 IU hCG peaked between 16-36 hours after mating and returned to baseline levels 72 hours after

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mating. Osteen and Mills (1979) also showed that the first peak increase in its concentration occurred 2-3 hours after mating, and the second was observed at 24-48 hours. Hashimoto et al. (2007) stated that the peak increase in rabbits super ovulated with FSH occurred 50 hours after the initiation of treatment, while LH concentration, as reported by Osteen and Mills (1979), peaked 2 hours after mating and returned to estrus levels within 12 hours. According to Argente (2021), the highest LH levels were observed 2 hours after natural mating, which was 5 times higher compared to the value at 48 hours before mating, and gradually decreased each day afterward. Furthermore, Cervantes et al. (2015) showed that LH concentration was peaked 1 hour after treatment and then gradually decreased after 3-6 hours. The measurement of FSH and LH concentration in this study was conducted on days 1, 3, and 5, at the point where both hormones were likely to return to their baseline levels, leading to non-significant differences between the 3 treatment groups.

The LH concentration in R1 was relatively stable, while fluctuations were observed in R2 and R3. Furthermore, statistical analysis of LH levels showed non-significant differences (P>0.05). The non-significant differences in FSH and LH concentration could be attributed to the growth of new follicles in the ovaries (Osteen and Mills, 1980) in preparation for the next cycle after the ovulation of de Graaf follicles. These new follicles released inhibin, which could reduce FSH secretion (Mills et al., 1981). The condition could also be due to the repetition and inappropriate interval of BPE injections. In this study, rabbit injections were repeated only 5 times over 3 days, in the morning and evening. Sayuti et al. (2022) stated that non-significant differences in hormone levels were often caused by the repetition factor in treatment and the interval of BPE injections. Similarly, Siregar and Hamdan (2007) mentioned that one factor to be considered for achieving optimal superovulation was repetition in treatment.

Another factor that contributed to the low levels of FSH and LH concentration was the low levels of specific hormones contained in BPE used for superovulation induction in the experimental rabbits (Sayuti *et al.*, 2022). Solihati (2006), stated that in addition to being affected by hormone levels in the blood, hormone activity also depended on the presence of receptors on target cells and transport proteins in the blood. The selection of the age of experimental animals could affect superovulation response, and this was consistent with Zhang *et al.* (2017).

CONCLUSIONS AND RECOMMENDATIONS

In conclusion, the administration of BPE could be considered as an alternative superovulation preparation in

local rabbits based on ovarian and hormonal responses.

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NOVELTY STATEMENT

The use of BPE for superovulation preparation as an alternative to PMSG and FSH.

AUTHOR'S CONTRIBUTION

H and SW wrote the initial manuscript, performed manuscript revision, and data analysis. RT, SHH, MFS, NS, MM, E, RPA, and PHN participated in performing, selecting samples, sample collection, and performed practical experiments. TNS and AS developed the original idea and protocol, conducted the research, and revised the final manuscript

CONFLICT OF INTEREST

The authors have declared no conflict of interest.

REFERENCES

- Adriani A, Sutama IK, Sudono A, Sutardi T, Manalu W (2004). The effects of superovulation prior to mating and zinc supplementation on milk yields in Etawah-grade does. J. Anim. Prod., 6: 86-94.
- Afriani T, Rahim F, Rachmat A, Mundana M, Farhana A (2020). Follicle stimulating hormone and gonadotropin releasing hormone administration to the superovulation of buffalo (*Bubalus bubalis*). Am. J. Anim. Vet. Sci., 15: 113-117. https://doi.org/10.3844/ajavsp.2020.113.117
- Amiruddin A, Siregar TN, Armansyah T, Hamdan H, Rifki M (2013). steroid level of aceh's cattle induced by pregnant mare's serum gonadotropin (PMSG) and follicle stimulating hormone (FSH). J. Kedokt. Hewan, 7: 120-124. https://doi. org/10.21157/j.ked.hewan.v7i2.923
- Argente MJ, Muelas R, Garcia ML (2021). Role of FSH, LH and prolactin in the components of litter size in female rabbits. In: Fábio Alessandro Pieri, editor. Prime Archives in Veterinary Sciences, Spanyol.
- Arimbawa IWP, Trilaksana IGNB, Pemayun TGO (2012). Profile progestrone hormone Bali cattle in one oestrus cycle. J. Indonesia Med. Vet., 1: 330-336.
- Arum WP, Siregar TN, Melia J (2013). The effect of bovine pituitary extract on superovulatory response of Aceh cattle.

J. Med. Vet., 7: 71-74. https://doi.org/10.21157/j.med.vet.. v7i2.2932

- Ashour G, Abdel-Rahman SM (2019). Hormonal changes in relation to productivity of pregnant rabbit does. World Vet. J., 9: 37-45. https://doi.org/10.36380/scil.2019.wvj6
- Browning JY, Keyes PL, Woli RC (1980). Comparison serum progesteron, 20α- dihydroprogesterone, and estradiol-17β in pregnant and pseudopregnant rabbits, evidence for post implantation recognition of pregnancy. Biol. Reprod., 23: 1014-1019. https://doi.org/10.1095/biolreprod23.5.1014
- Cervantes MP, Palomino JM, Adams GP (2015). In vivo imaging in the rabbit as a model for the study of ovulationinducing factors. Lab. Anim., 49(1): 1-9. https://doi. org/10.1177/0023677214547406
- Challis JR, Davies IJ, Ryan KJ (1973). The concentrations of progesterone, estrone and estradiol-17β in the plasma of pregnant rabbits. Endocrinology, 93: 971-976. https://doi. org/10.1210/endo-93-4-971
- Demoustier MM, Beckers JF, Zwalmen PVD, Closset J, Gillard JL, Ectors F (1988). Determination of porcine plasma follitropin levels during superovulation treatment in cows. Theriogenology, 30: 379-386. https://doi. org/10.1016/0093-691X(88)90185-9
- Fortune JE (1993). Follicular dynamics during the bovine estrous cycle: A limiting factor in improvement of fertility. Anim. Reprod. Sci., 33: 111-125. https://doi.org/10.1016/0378-4320(93)90110-D
- Fortune JE, Sirois J, Turzillo AM, Lavoir M (1991). Follicle selection in domestic ruminants. J. Reprod. Fertil., 43(1):187-98.
- Hafizuddin H, Suryani S, Yusmadi Y, Siregar TN, Armansyah T (2010). Respon superovulasi mencit dengan ektrak hipofisa sapi: suatu upaya menemukan agen superovulasi dengan biaya murah. Prosiding Seminar Nasional Peternakan Berkelanjutan. Fakultas Peternakan Universitas Padjajaran.
- Hashimoto S, Kimura K, Kuramochi T, Aoyagi K, Hirako M, Kawaguchi M, Iwata H, Hirao M, Kitada K, Hirasawa K, Ueda M (2007). Responsiveness of rabbits to superovulation treatment by a single injection of follicle-stimulating hormone with aluminum hydroxide gel. Mol. Reprod. Dev., 74(9): 1208-1212. https://doi.org/10.1002/mrd.20672
- Holt JA, Heise WF, Wilson SM, Keyes PL (1976). Lack of gonadotropin activity in the rabbit blastocyst prior to implantation. Endocrinology, 98(1): 904-909. https://doi. org/10.1210/endo-98-4-904
- Isnaini N, Wahyuningsih S, Ciptadi G (1999). *In vitro* maturasi oosit sapi dalam TCM199 yang disuplementasikan dengan ekstrak hipofisa sapi. J. Saintek, 7(1): 75-85.
- Ju JC, Chang YC, Huang WT, Tang PC, Cheng SP (2001). Superovulation and transplantation of demi-and aggregated embryos in rabbits. Asian-Australasian J. Anim. Sci., 14: 455-461. https://doi.org/10.5713/ajas.2001.455
- Kanayama K, Nariai K, Sankai T, Endo T (1995). Superovulation induction by human menopausal gonadotrophin in rabbits. J. Vet. Med. Sci., 57: 543-544. https://doi.org/10.1292/ jvms.57.543
- Karl KR, Jimenez-Krassel F, Gibbings E, Ireland JLH, Clark ZL, Tempelman RJ, Latham KE, Ireland JJ (2021). Negative impact of high doses of follicle-stimulating hormone during superovulation on the ovulatory follicle function in small ovarian reserve dairy heifers. Biol. Reprod., 104: 695-705. https://doi.org/10.1093/biolre/ioaa210
- Kauffman RD, Schmidt PM, Rall WF, Hoeg JM (1998).

Advances in Animal and Veterinary Sciences

OPEN OACCESS

Superovulation of rabbits with FSH alters in vivo development of vitrified morulae. Theriogenology, 50: 1081-1092. https://doi.org/10.1016/S0093-691X(98)00209-X

- Kleden MM, Soetanto H (2017). Concentration of progesterone and prolactin hormons and milk production of New Zealand White rabbits doe fed moringa leaves meal. Mediterr. J. Soc. Sci., 8(3): 79-85. https://doi.org/10.5901/mjss.2017. v8n3p79
- Langford GA, Marcus GJ, Batra TR (1983). Seasonal effects of PMSG and number of inseminations on fertility of progestogen-treated sheep. J. Anim. Sci., 57: 307-312. https://doi.org/10.2527/jas1983.572307x
- Lehloenya KC, Greyling JPC (2010). The effect of embryo donor age and parity on the superovulatory response in Boer goat does. South Afr. J. Anim. Sci., 40: 65-69. https://doi. org/10.4314/sajas.v40i1.54130
- Lestari TD, Ismudiono I (2014). Ilmu Reproduksi Ternak. Airlangga University Press, Surabaya.
- Maertens L, Luzi F (1995). Catatan tentang pengaruh stimulasi PMSG pada angka kematian saat lahir dan distribusi ukuran anak dalam dosis inseminasi buatan. Sains Kelinci Dunia, 1(3): 57-61.
- Marhaeniyanto E, Kasthama IGP (2007). Pengaruh induksi PGF2A dengan hormon PMSG pada jumlah korpus luteum, embrio dan jumlah anak kelinci. Buana Sains, 7: 105-112.
- Mattioli S, Maranesi M, Castellini C, Dal BA, Arias-Alvarez M, Lorenzo PL, Rebollar PG, Garcia-Garcia RM (2021). Physiology and modulation factors of ovulation in rabbit reproduction management. World Rabbit Sci. Assoc., 29: 1-10. https://doi.org/10.4995/wrs.2021.13184
- Menchaca A, Vilarino M, Crispo M, Pinczak A, Rubianes E (2007). Day 0 protocol: Superstimulatory treatment initiated in the absence of a large follicle improves ovarian response and embryo yield in goats. Theriogenology, 68: 1111-1117. https://doi.org/10.1016/j.theriogenology.2007.07.020
- Mills T, Copland A, Osteen K (1981). Factors affecting the postovulatory surge of FSH in the rabbit. Biol. Reprod., 25: 530-535. https://doi.org/10.1095/biolreprod25.3.530
- Nalley WM, Kune P, Hine TM (2017). Tampilan estrus dan angka kebuntingan sapi bali pascapemberian ekstrak pituitari. Acta Vet. Indones., 5: 74-80. https://doi.org/10.29244/ avi.5.2.74-80
- Nur NA, Siregar TN, Hamdan H, Nasution I, Thasmi CN, Dasrul D (2006). Effect of PMSG and hCG to increase birth number in Campbell's hamsters (*Phodopus campbelli*). J. Med. Vet., 10: 59-61.
- O'callaghan D, Yaakub H, Hyttel P, Spicer LJ, Boland MP. 2000. Effect of nutrition and superovulation on oocyte morphology, follicular fluid composition and systemic hormone concentrations in ewes. Journal of reproduction and fertility. 118(2):303-314.
- Osteen K, Mills T (1980). Changes in the size, distribution, and steroid content of rabbit ovarian follicles during early pseudopregnancy. Biol. Reprod., 22(5): 1040-1046. https:// doi.org/10.1095/biolreprod22.5.1040
- Osteen KG, Mills TM (1979). Serum LH and FSH levels in the pregnant rabbit. Proc. Soc. Exp. Biol. Med., 162: 454-457. https://doi.org/10.3181/00379727-162-40703
- Sayuti A, Hamdan H, Jannah M, Safriadi S, Nasri T, Siregar TN (2022). Superovulation induction with bovine pituitary extract in local rabbits. Punjab Univ. J. Zool., 37: 29-34. https://doi.org/10.17582/journal.pujz/2022.37.1.29.34

- Scaramuzzi RJ, Adams NR, Baird DT, Campbell BK, Downing JA, Findlay JK, Henderson KM, Martin GB, McNatty KP, McNeilly AS, Tsonis CG. 1993. A model for follicle selection and the determination of ovulation rate in the ewe. Reproduction, fertility and development. 5(5):459-478.
- Setiawan D, Iskandar F, Arifin HD (2019). Tingkah laku awal berahi kambing kaligesing (*Capra aegagrus hircus*) betina yang diinjeksi ekstrak hipofisa dengan level berbeda. J. Riset Agribisnis dan Peternakan, 4: 49-55.
- Siregar IK, Siregar TN, Akmal M, Wahyuni S, Nazaruddin N, Hafizudin H (2020). Efek pemberian ekstrak pituitary sapi terhadap peningkatan jumlah folikel ovarium dan konsentrasi estradiol pada tikus putih (*Rattus norvegicus*). Livest. Anim. Res., 18: 171-179. https://doi.org/10.20961/ lar.v18i2.42956
- Siregar TN, Hamdan H (2007). Teknologi reproduksi pada ternak. Universitas Syiah Kuala, Banda Aceh.
- Siregar TN, Siregar IK, Armansyah T, Sayuti A (2013). The reproductive performance of local does following induced superovulation with cattle pituitary extracts. J. Vet., 14: 91-98.
- Solihati N (2006). Treatment superovulation before animal slaughter. J. Ilmu Ternak Univ. Padjadjaran, 6: 145-149.
- Song S, Jang D, Min C, Park J, Joo Y, Lee J, Chung K (2012). Effects of parity and season on production of embryos in superovulated Hanwoo. J. Embryo Transfer, 27: 127-131.
- Stheskumar S (2006). Superovulation and embryo recovery in two breeds of rabbits. Indian J. Biotechnol., 5: 54-57.
- Suartini NK, Trilaksana IGNB, Pemayun TGO (2013). The levels of estrogen and the onset of estrous after administration of buserelin (GnRH agonist) on Bali cattle experiencing anestrous postpartum due to ovarian hypofunction. J. Ilmu dan Kesehatan Hewan, 1: 40-44.
- Sudjatmogo B, Utomo, Subiharta, Manalu W, Ramelan (2001). Tampilan produksi susu akibat peningkatan pertumbuhan ambing sapi perah Friesian Holstein yang disuntik PMSG pada program perkawinannya. J. Trop. Anim. Developm., 26: 8-13.
- Sugiyatno, Sumaryadi MY, Haryati (2001). Konsentrasi estrogen serum kaitannya dengan lama birahi domba ekor tipis yang diinduksi PMSG. Anim. Prod., 3: 40-44.
- Sumiyoshi T, Tanaka T, Kamomae H (2014). Relationships between the appearances and changes of estrous signs and the estradiol-17ß peak,luteinizing hormone surge and ovulation during the periovulatory period inlactating dairy cows kept in tie-stalls. J. Reprod. Dev., 60: 106-114. https:// doi.org/10.1262/jrd.2013-119
- Syafruddin S, Wahyuni S, Gholib G, Siregar TN (2022). Comparison of four methods of inducing pseudopregnancy in rabbits. Med. Weterinariya, 78: 85-90. https://doi. org/10.21521/mw.6608
- Szendro Z, Matics Z, Brecchia G, Theau-Clement M, Nagy Z, Princz Z, Biro-Nemeth E, Radnai I, Nagy I (2010). Milk production of pseudo pregnant and multiparous does. World Rabbit Sci., 18: 77-82. https://doi.org/10.4995/ wrs.2010.18.10
- Techakumphu M, Numchaisrika P, Suktrakun V, Phanitkitcharoen S, Kaewnanuer A, Suwajanakorn S (2002). Superovulation responsesin rabbits to different does of follicle stimulating hormone. Thai J. Vet. Med., 32: 53-58. https://doi.org/10.56808/2985-1130.1875
- Tjitosumirat T (2009). Kajian hubungan konsentrasi hormon progesteron dengan jumlah corpora lutea pada kambing. J.

Advances in Animal and Veterinary Sciences

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Ilmiah Aplikasi Isotop dan Radiasi, 1: 43-53.

- Wang Y, Zhang JMX, Xu L (2015). Control of PMSG-based wind turbines for system inertial response and power oscillation damping. Inst. Elect. Electron. Eng., 6: 1-10. https://doi.org/10.1109/TSTE.2015.2394363
- Wijono Didi B (1998). Peran kadar progesteron dalam plasma darah untuk deteksi estrus dan aktivitas ovarium. Semin. Nasional Petern. Vet., pp. 268-269.
- Zhang H, Cheng GH, Li YJ, Cai MY, Guo HY, Le-Qin K (2017). Superovulation and expression of follicle-stimulating hormone receptor in young rabbit females. World Rabbit Sci., 25: 167-172. https://doi.org/10.4995/wrs.2017.4485
- Zulkarnain, Sutiyono, Setiatin ET (2015). Pemanfaatan ekstrak hipotalamus kambing sebagai upaya optimalisasi kesuburan kambing Kejobong betina. J. Vet., 16: 343-350. https://ojs. unud.ac.id/index.php/jvet/article/view/16248