



Effect of Metformin on P53 and its Interacting Partners in Breast Cancer Cell Line MCF-7

Mehroze Amin¹, Afifa Yaqub¹, Qindeel Fatima¹, Rabail Hassan Toor^{1,3} and Abdul Rauf Shakoori^{1,2,*}

¹School of Biological Sciences, University of the Punjab, Quaid-i-Azam Campus, Lahore 54590, Pakistan

²Cancer Research Centre, University of the Punjab, Quaid-i-Azam Campus, Lahore 54590, Pakistan

³UVM Cancer Center, Larner College of Medicine, Department of Biochemistry, University of Vermont, Burlington, VT 05405, USA

ABSTRACT

Metformin is known to favor p53 anti-cancer activity and promote apoptosis and cell senescence in cancer cells. It is also known to decrease the incidence of several cancers though it is not entirely true for breast cancer. In the present study, the effect of metformin has been studied on the expression of p53 and its interacting partners MDM-2, PIRH2 and Δ Np73 in breast cancer cell line MCF-7 and non-cancerous kidney cell line HEK2093. Analysis of the cytotoxicity and proliferation parameters showed that higher concentrations of metformin were cytotoxic for the breast cancer cells with EC_{50} of 22.75 μ M. Analysis of expression of p53 and the interacting partner genes showed that metformin upregulated p53 expression in a dose-dependent manner, while the expression of interacting partners is downregulated indicating that metformin exhibits the anti-cancerous effect by regulating the p53 pathway. Altogether this study reports the effect of metformin on the expression of p53 and its interacting partners MDM2, PIRH2 and Δ Np73 in breast cancer cell line MCF-7, thereby providing evidence that metformin modulates p53 pathway to exhibit its anti-cancer property. Hence, metformin can be a potential therapeutic candidate for breast cancer treatment. However, further investigations are required to provide insight into the therapeutic potential of metformin for cancer.

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ARS conceptualization, fund acquisition, project administration, supervision, writing, review, editing and provided resources. MA methodology, writing original draft. AY help in methodology. QF methodology help in cell culture. RHT writing, review and editing.

Key words

Breast cancer, MDM2, PIRH2, ANP73, Metformin, Metastasis, P53

INTRODUCTION

Breast cancer is one of the most lethal and diversified malignant tumors in the world. It is a general health dilemma for women and accounts for 24.2% of female cancers up to 2018 (Akram *et al.*, 2017). Cancer recurrence and metastasis create problems that lead to a high mortality rate. There is a critical need for discovering unique target genes for the prediction and treatment of breast cancer (Zhang *et al.*, 2021). P53 also known as guardian of human genome induces senescence, growth arrest, and apoptosis to play the role of a tumor suppressor gene. In 50% of human cancers, it is known to be suppressed or inactivated

by mutations. As mutant p53 is frequently found involved in cancer proliferation it is known to be an exciting and promising target in cancer therapy (Wang and Sun, 2010).

Metformin, an anti-diabetic drug that is recently being studied for its anti-cancer properties, works with the p53 gene to exert its anti-tumor effects. Metformin limits oxidative phosphorylation that results in reduced ATP formation and ultimately AMP level in the cell rises. AMP-activated protein kinase (AMPK) phosphorylates the p53 gene as a result p53 attains stabilized structure and becomes activated. P53 being a tumor suppressor gene plays a major role in apoptosis and cellular senescence of the cancer cells. P53 acts in a positive feedback loop with AMPK by inhibiting mTOR (mammalian target of rapamycin) to arrest cell progression (Faria *et al.*, 2019). Isleem *et al.* (2020) showed that metformin combined with olive leaf extract resulted in reduced viability in the case of cancer cells through a powerful synergistic inhibitory effect.

Protein levels of p53 and its transcriptional activity are tightly regulated by E3 ubiquitin Mdm2-protein ligase Mdm2, encoded by MDM2 gene, the expression of which is transcriptionally regulated by p53 in a negative feedback loop (GarcíaCano *et al.*, 2020). In 2021, studies

* Corresponding author: arshakoori.sbs@pu.edu.pk, arshaksbs@yahoo.com
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were carried out to find the effect of 3, 4-dihydroxy-5, 4'-dimethoxybibenzyl (DS-1) on targeting MDM2 and restoring p53 function in lung cancer cells, as targeting MDM2, a negative regulator of p53 has recently attracted interest in cancer drug research as it may restore tumor suppressive function (Putri *et al.*, 2021).

The oncogenic Δ Np73 controls cell survival and self-renewing properties of stem cells, and promotes increased susceptibility to carcinogenesis. Δ Np73 overexpression has been commonly associated with lymph node metastases and vascular invasion, chemoresistance, and poorer patient prognosis. Δ Np73 also acts as a negative regulator of P53 thereby sustaining cancer cell proliferation (Ramos *et al.*, 2020).

Pirh2, a ubiquitin-protein ligase, has been reported to promote p53 degradation. Pirh2 overexpression could potentially cause degradation of p53 and reduce its tumor suppression function in the lung tumor cells (Duan *et al.*, 2006).

The aim of this study was to determine the effect of metformin on the expression level of the p53 gene and its interacting partners (PIRH2, MDM-2, and Δ Np-73) in breast cancer cell line MCF-7 and normal human embryonic kidney HEK-293 cell line. This study aimed to determine the anti-cancer effect of metformin concerning the p53 pathway.

MATERIALS AND METHODS

Maintenance of cell lines

In this study, breast cancer cell line MCF-7, and normal embryonic kidney cell line HEK-293 (CRL-1573TM) acquired from ATCC (American Type Culture Collection, Manassis, VA, USA (catalog No. 11686-029 and 30-2003, respectively) were used. The cell cultures were maintained in Dulbecco's Modified Eagle Medium (DMEM, Catalogue No. 11995065, GIBCO, Thermofisher Scientific, Waltham, MA, USA) with 10% heat-inactivated fetal bovine serum (FBS, Catalogue No. 10082147, GIBCO) and 1 % streptomycin/penicillin (catalog No. 15140148, GIBCO). Cells were maintained at 37°C in a humid atmosphere with 5% CO₂ maintained in air.

In vitro cytotoxicity assay

MCF-7 cells were seeded at the density of (3.0×10³ cells per cm²) in a 24-well culture plate. After 24 h of seeding, metformin treatment was given in a series of concentrations (1 μM, 5 μM, 10 μM, 20 μM, 30 μM). Untreated (control) cells were treated with a complete growth medium supplemented with 10 μl 1X PBS.

After 48 h of metformin treatment, neutral red assay was performed. The culture plates were incubated for 2 h in a CO₂ incubator following the procedure mentioned in

Repetto *et al.* (2008). The dye was extracted and its optical density was measured at 570 nm by using BioTek-808 ELISA absorbance reader. Since the assay is based on the lysosomal uptake of dye, the absorbance corresponded to the number of viable cells.

Percentage cell viability was calculated by using the following formula:

$$\text{Cell Viability (\%)} = \frac{\text{Average Absorbance of treated cells}}{\text{Average Absorbance of untreated cells}} \times 100$$

Growth inhibition and EC₅₀ were calculated by using Microsoft Excel.

In vitro cell proliferation assay

BrdU assay was carried out following the protocol provided by Sigma Aldrich (Sigma BrdU Cell Proliferation ELISA Kit Catalog no. 11296736001). Briefly, 1X BrdU reagent was added to each well for 4 h at 37 °C and 5% CO₂. Cells were fixed and DNA was denatured to make incorporated BrdU dye more accessible for detection by antibodies. The pre-diluted anti-BrdU primary or secondary peroxidase-conjugated monoclonal detector antibodies were added to each well (100 μl/well) and incubated for 1 h at room temperature. After washing thrice with washing buffer, the bound peroxidase was detected by substrate reaction, which was stopped by adding stop solution (100 μl/well), and then O.D of the yellow reaction product was measured at 450 nm using a BioTek-808 Elisa absorbance reader. The intensity of the color was proportional to the BrdU incorporation in the DNA. For metformin-treated cells, the percentage cell proliferation was calculated using Microsoft Excel taking untreated cells (control) as 100%.

The following formula was used to calculate percentage cell proliferation.

$$\text{Cell Proliferation (\%)} = \frac{\text{Av. Absorbance of treated cells}}{\text{Av. Absorbance of untreated cells}} \times 100$$

Quantitative real-time RT-PCR (qRT-PCR)

After 48 h of the treatment of metformin with a series of concentrations of metformin (1 μM, 5 μM, 10 μM, 20 μM, 30 μM) total RNA was isolated from treated and untreated cells using the TRIzol method. cDNA synthesis Kit (RevertAid First Strand cDNA synthesis kit-(catalog number: K1621) was used for cDNA synthesis. Primers used in this study are shown in Table I.

Each cDNA template was diluted to make the final concentration 40 ng/ μl. Maxima SYBR Green/ Low ROX qPCR Master Mix (catalog no. K0221) was used to set up the experiment of RT PCR where untreated samples were used as a calibrator and *HMBS* was used as a reference gene for normalization. The reaction was carried out in triplicates (biological replicates). Comparative gene expression was checked using the 2^{-ΔΔCt} method.

Statistical analysis

The graphs were plotted using Microsoft Excel and GraphPad Prism (v 7.03). Statistical analyses were carried out by using GraphPad Prism (V.7.03). For the cell-based assays, one-way ANOVA with Dunnett's test for multiple comparisons was carried out while for the real-time PCR, the statistical analysis was performed using two-way ANOVA with Tukey's test for multiple comparisons. In the case of both tests, analysis was performed with a 95% confidence interval with statistical significance at $P \leq 0.05$. Further indication of the level of statistical significance is shown as follows: *($p \leq 0.05$), **($p \leq 0.01$), ***($p \leq 0.005$), ****($p \leq 0.001$).

Table I. Primers for quantitative (real-time) PCR of target specific genes used in this study.

Target gene	Primer sequence 5' → 3'	Product size	Melting temperature TM
PIRH2	TGGGGTAAGTGTTCCTCA	137bp	58.9°C
	TGTGGTAAAAGTGGGTTTGG		58.3°C
$\Delta Np73$	CAGCCCATCAAGGAGGAGTT	127bp	61.5°C
	TGAGGCAGTTTTGGACACAC		59.7°C
MDM2	TGCCATTGAACCTTGTGTGA	127bp	60°C
	GCAGGGCTTATTCCTTTTCT		58°C
P53	AGAGTCTATAGGCCACCCC	140bp	60.04°C
	GCTCGACGCTAGGATCTGAC		59.96°C
HMBS	TACCCGAGAGGAGAGAACA	203bp	59°C
	CGAGCAGGAAGACCAGAAAC		60°C

*All primers were designed using Primer-Blast by NCBI (<https://www.ncbi.nlm.nih.gov/>).

RESULTS

Morphology of cells

The HEK-293 human embryonic kidney cell line was used as a model for non-cancerous cells in this study. Both MCF-7 and HEK-293 cell lines have epithelial-like morphology (Fig. 1). Morphologically it was observed that the untreated cells (control) and the cells treated with lower concentrations of metformin i.e., 1 μ M and 5 μ M exhibited their normal morphology, while the cells treated with higher metformin concentrations i.e., 10 μ M, 20 μ M, and 30 μ M exhibited stressed morphology, with smaller sizes and lesser number of cells compared to control cells (Fig. 2).

Cell viability and cell proliferation

Cell viability of MCF-7 cells treated with different concentrations of metformin was determined by lysosomal uptake of neutral red dye. Figure 3A shows that the

cells treated with 1 μ M metformin had comparable cell viability (97.8 %) to the control cells. The cell viability was significantly ($P \leq 0.05$) reduced in cells treated with 5 μ M metformin with 82.2 % viable cells. At higher concentrations, further reduction in cell viability was observed with 72 %, 69.7 %, and 33.9 % viable cells at 10 μ M, 20 μ M, and 30 μ M metformin-treated cells respectively. We observed a dose-dependent relation of metformin on cell viability.

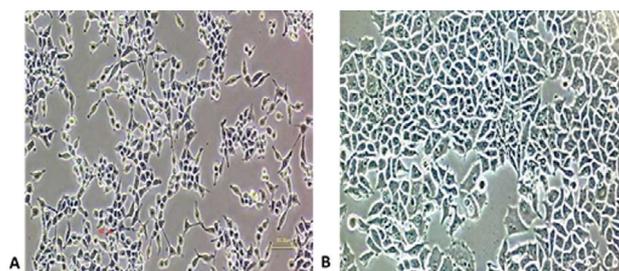


Fig. 1. Morphology of human embryonic kidney cell line HEK-293 (A) and human breast cancer cell line MCF-7. (B) Both cell lines showed normal epithelial-like morphology. The cells are 60 – 70 % confluence at the time images were taken under 100X magnification.

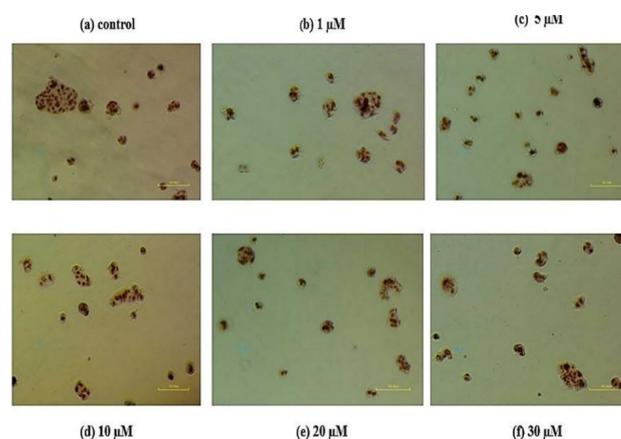
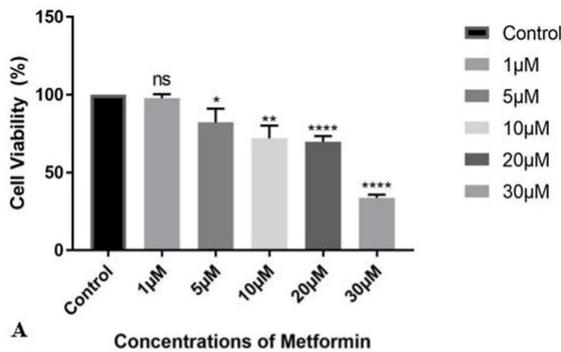


Fig. 2. MCF-7 cells stained with neutral red after treatment with different concentrations of metformin (1, 5, 10, 20 and 30 μ M) for 48h. Compared with control, cells treated with higher concentrations of metformin appear to be stressed with smaller size and reduced numbers.

BrdU, a synthetic thymidine analog, incorporation into DNA was determined to determine cell proliferation. Figure 3B shows the effect of metformin on the proliferation of MCF-7 cells. We observed a dose-dependent reduction in cell viability with increasing metformin concentrations in the MCF-7 cell line. The analysis of the percentage cell proliferation showed that the concentrations 1 μ M, 5 μ M, and 10 μ M had 86.4 %, 85.2 %, and 79.1 % proliferative

cells respectively, which although was less than the control, the difference was not found to be statistically significant with $P \geq 0.05$. However, at higher concentrations of metformin i.e., 20 μM and 30 μM the cell proliferation was found to be 64 % and 53.9 % which was significantly reduced ($P \leq 0.05$) compared to the control (Fig. 3B).

Cell Viability of Metformin Treated MCF-7 Cells



Cell Proliferation of Metformin Treated MCF-7 Cells

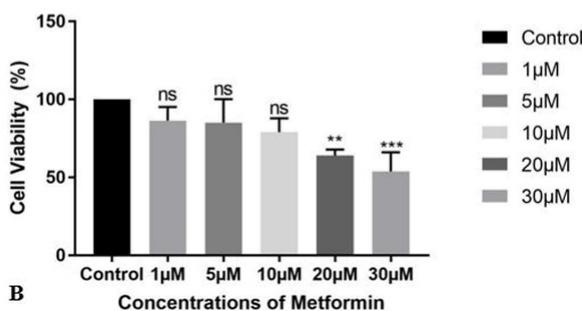


Fig. 3. Effect of different concentrations of metformin on cell viability (neutral red assay) and cell proliferation (BrdU assay) of breast cancer cell line MCF-7. Significant reduction was observed in these parameters at higher concentrations of metformin. ns= not significant, * = $P \leq 0.05$, ** = $P \leq 0.01$, **** = $P \leq 0.0001$.

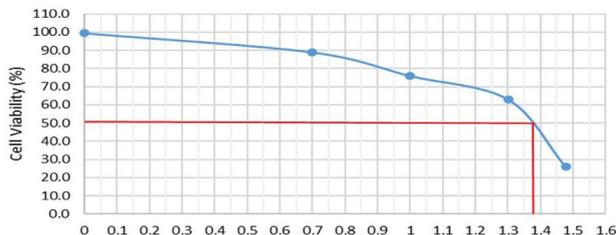


Fig. 4. A dose-dependent logarithmic curve for metformin treated MCF-7 cells. The log values for the concentrations of metformin were calculated and plotted against percentage cell viabilities. Antilog value corresponding to 50 % cell viability (EC_{50} of metformin) for MCF-7 cell line was 22.75 μM .

Figure 4 shows the effective concentration of metformin with 50 % cell viability of MCF-7 cells ($EC_{50} = 22.75 \mu\text{M}$) based on cell viability data.

According to the outcomes, metformin demonstrated a concentration-dependent anti-proliferative activity in MCF-7 cells.

Effect of metformin on p53 gene expression

Analysis of the expression of the p53 gene showed that in experimental controls of both MCF-7 and HEK cells, the expression of p53 was at the basal level. However, metformin treatment to MCF-7 upregulated the expression of the p53 gene in a concentration-dependent manner. In MCF7 cells, the p53 expression was upregulated 1.8-fold, 2.26-fold, and 3.20-fold after treatment with 1, 5, and 10 μM metformin. This increase was 4.3-fold and 8.8-fold, respectively after treatment with 20 μM and 30 μM metformin (Fig. 5A). A similar trend in the expression of p53 was observed in metformin-treated HEK-293 cells. The cells at 1 μM , 5 μM , and 10 μM had 1.16-fold, 1.35-fold, and 1.31-fold p53 expression respectively, which was upregulated to 12.8-fold and 11.47-fold in 20 μM and 30 μM metformin-treated HEK-293 cells, respectively.

Effect of metformin on the expression of interacting partner genes of p53 gene

Figure 5 shows the effect of different concentrations of metformin chloride on the interacting partner genes such as $\Delta Np73$, $PIRH2$, $MDM-2$ in MCF-7 and HEK-293 cell lines. The relative expression of $\Delta Np73$ was 10.79 in MCF7 and 7.37 in HEK-293 experimental controls. Compared to the experimental control, the expression of the $\Delta Np73$ gene was significantly ($P \leq 0.05$) downregulated in metformin-treated MCF-7 cells in a dose-dependent manner. The relative expression recorded was 4.16, 3.85-fold, 2.27, 2.2, and 1.41 after treatment of MCF-7 cells with 1, 5, 10, 20, and 30 μM metformin (Fig. 5B). HEK-293 cells also showed statistically significant ($P \leq 0.05$) downregulation in the expression of the $\Delta Np73$ gene compared to its experimental control, except for 1 μM metformin treatment), where the difference in expression was not significant compared to the experimental control sample.

$Pirh2$ is another interacting partner and negative regulator of p53. In MCF-7 cells, the relative expression of $PIRH2$ was 8.17 in the experimental control sample, which was downregulated after metformin treatment (Fig. 5C). An opposite trend was observed in the metformin-treated HEK-293 cells, where a slight, non-significant ($P \geq 0.05$) upregulation in $PIRH2$ gene expression was observed in the metformin-treated cells compared to its experimental control. As in the MCF-7 cells, the upregulation in the expression was not dose-dependent.

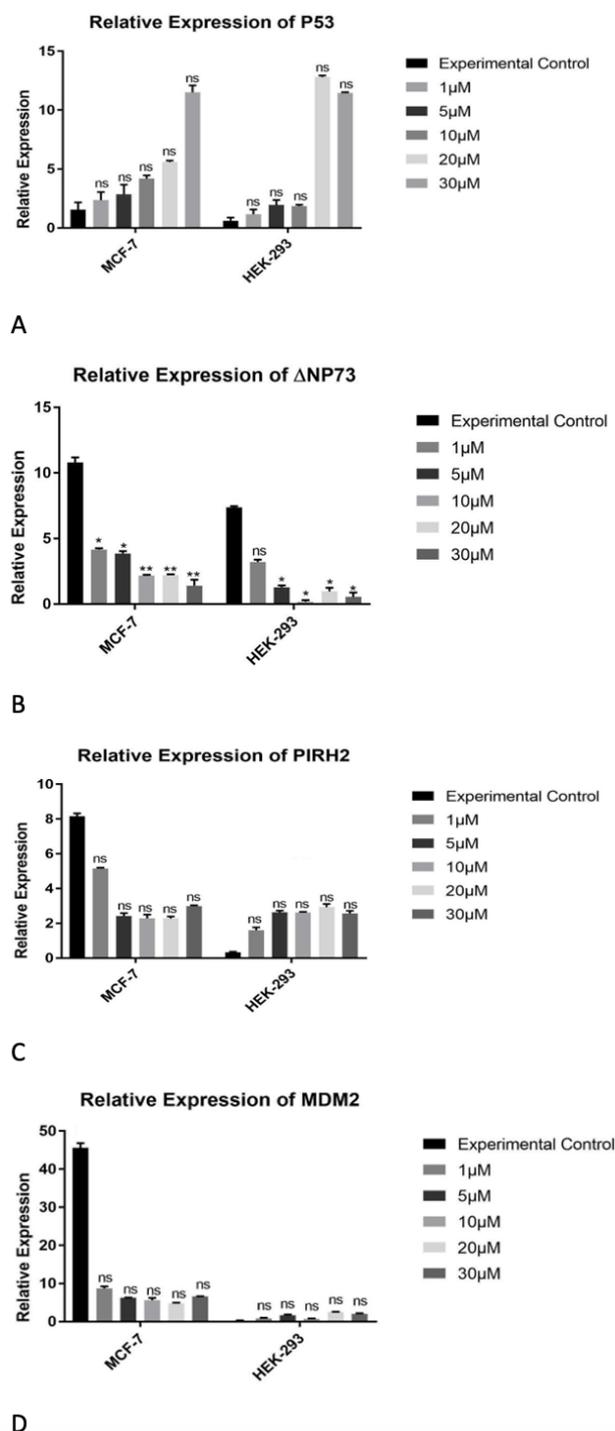


Fig. 5. Effect of metformin on expression of *P53*, *ΔNp73*, *PIRH* and *MDM2* in metformin treated MCF-7 and HEK-293 cells. Compared to control, dose-dependent upregulation was observed in *p53*, and downregulation in expression of other genes in MCF-7 and HEK-293 cell lines.

Like $\Delta Np73$ and *PIRH2*, *MDM2* is a negative regulator of the *p53* gene (Fig. 5D). It was observed that the relative expression of the *MDM2* gene was 45.6 in the experimental control MCF-7 cells. Treatment with different concentrations of metformin led to downregulation of *MDM2* gene expression, however, the dose-dependent downregulation was not much evident. Statistical analysis showed that compared to MCF-7 experimental control, the downregulation in the expression of *MDM2* was not significant ($P \geq 0.05$) which is indicated by “#” bar in Figure 5D. In the HEK-293 experimental control, the expression of *MDM2* was below 0, and metformin treatment slightly and non-significantly ($P \geq 0.05$) upregulated the expression of *MDM2* in HEK-293 cells. However, the fold change in expression was at the basal level, closer to 1 (Fig. 5D).

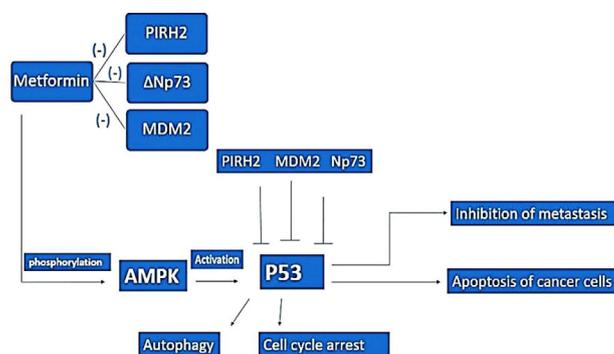


Fig. 6. Schematic representation of antitumor activity of metformin.

DISCUSSION

Metformin is a biguanide used for the treatment of type 2 diabetes mellitus for decades. It is right now the most prescribed anti-hyperglycemic drug used with a good safety profile as it improves glycemic control of the patient without any additional toxicity problem. Epidemiological studies depicted a correlation between T2DM (Type 2 diabetes mellitus) and the incidence of malignancies. Multiple studies linked breast cancer risk with T2DM until, in 2005, the first paper was published that showed reduced breast cancer risk in T2DM patients treated with metformin compared to the patients who were given some other treatment or therapy for diabetes. Since then, research is going on to find the mechanism behind the anti-tumor activity of metformin and its link to reduced risk of breast cancer. Finding the biological mechanisms of the metastatic phenomena is crucial to search for open therapeutic targets for successful interventions (Fares *et al.*, 2020)

Apart from other mechanisms that were proposed to

study metformin's antitumor activity, one important and promising mechanism is its relation to p53. Metformin limits oxidative phosphorylation that results in reduced ATP formation and ultimately AMP level in the cell rises. This AMPK is a key regulator of energy homeostasis of the body, it senses the energy state of the cell and shifts the metabolism of the cell accordingly. That is how metformin carries out its anti-hyperglycemic activity. AMPK phosphorylates the p53 gene as a result p53 attains stabilized structure and becomes activated. P53 being a tumor suppressor gene plays a major role in apoptosis and cellular senescence of the cancer cells. P53 acts in a positive feedback loop with AMPK by inhibiting mTOR to arrest cell progression (Faria *et al.*, 2019).

Until now BrdU assay was carried out to find the anti-proliferative effect of metformin on MCF-7 cells at 10 μ M concentration for different time durations 10 h, 24 h, and 72 h and the results showed that the metformin reduced the proliferation of cells in a time-dependent manner (Queiroz *et al.*, 2014).

The effect of metformin on the esophageal squamous carcinoma cell lines was checked to find the anti-tumor role of metformin. It was discovered that metformin could reduce cell proliferation in the case of squamous cancer cells and the rate of inhibition was found to be directly proportional to the concentration of metformin given and time administration. Proving that metformin can be used as a new approach to treat esophageal squamous carcinoma cases (Shao *et al.*, 2019)

Similar anti-proliferative activity of metformin on MCF-7 breast cancer cells was observed in the present study, where a direct dose-dependent reduction in cell viability and proliferation was observed. In the present study, the EC_{50} of metformin was found to be 22.75 μ M.

In a study carried out in 2019, cervical cancer cell line HeLa was treated with 60 mM, and 20 mM of metformin, and the expression of p53 and cyclin D1 was measured. The results showed that the dose was sufficient to increase the p53 expression significantly (Yudhani *et al.*, 2019).

Our studies showed an elevated p53 expression when breast cancer cell line MCF-7 were given the treatment of metformin in a range of concentrations (1 μ M, 5 μ M, 10 μ M, 20 μ M, 30 μ M). It showed that with the increase in the concentration of metformin the expression of p53 was upregulated in the MCF-7 as well as HEK-293 cells. While the upregulation of expression was 1.8, 2.2 and 3.2-fold were observed in 1 μ M, 5 μ M and 10 μ M treated MCF-7 cells, while treatment with 20 μ M and 30 μ M metformin upregulated the expression to 4.3-fold and 8.4-fold, respectively. A similar trend was observed in the case of HEK-293. Apparently metformin causes decrease in tumor cell proliferation since it phosphorylates and activates p53

which ultimately causes cell cycle arrest and apoptosis in cancer cells.

MDM-2 (murine double minute 2) is a principle negative regulator of the p53 gene and it maintains a low p53 level in the cell by targeting p53 to rapid proteolytic degradation thus inhibiting its transcriptional activity. That's why MDM-2 is a logical therapeutic target used for cancer treatment and therapies (Noon *et al.*, 2010).

Our findings showed that when given metformin treatment in the concentration of 1 μ M, 5 μ M, 10 μ M, 20 μ M and 30 μ M to breast cancer cells MCF-7, the rate of expression of MDM-2 was found to be decreasing as the concentration was increased.

Although there is a dose-dependent downregulation, the fold expression values are comparable to each other. In the case of HEK-293, however the fold change in expression was close to 1.

Presently, there is very limited information on the rate of expression of PIRH2 in the metformin-treated breast cancer cell lines. Our findings show that with the increasing metformin concentration the rate of expression of PIRH2 in MCF-7 cell line has downregulated. However, there were slight changes in fold differences that were observed among different concentrations of metformin used. In the case of metformin-treated HEK-293 cells; an opposite trend was observed where a slight, non-significant ($P \geq 0.05$) upregulation in PIRH2 gene expression was observed in the metformin-treated cells compared to its experimental control.

As Pirh2 is a negative regulator of p53, downregulation in the expression level of *PIRH2* in the case of MCF-7 after metformin treatment proves that metformin plays a role to inhibit or decrease the expression of cancerous factor Pirh-2 that ultimately enhanced the p53 expression leading to apoptosis and senescence.

$\Delta Np73$ is an oncogene that inhibits p53 activity and sustains cancer cell proliferation by suppressing apoptosis. The expression analysis of $\Delta Np73$ on esophageal squamous carcinoma cells has been done in the past and the results showed a reduced level of $\Delta Np73$ *in vivo* (Noon *et al.*, 2010).

However, further studies on the p53 pathway, with longer duration of treatment, and more factors involved in the p53 pathway will provide further insights into the mechanism anti-cancer effects of metformin cancer cells.

CONCLUSION

To conclude, by upregulating the expression of the p53 gene and by negatively regulating cancerous interacting partners Mdm-2, Pirh2, and $\Delta Np73$ involved in the p53 pathway, metformin showed anti-cancer properties and it

can be used as a target for therapeutic purposes in case of breast cancer. However, further investigation is required to find the exact mode of action of anti-tumor activity of metformin.

Statement of conflict of interest

The authors have declared no conflict of interest.

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