



# Thermal Physiology and Bioenergetics in Ambient or Seasonal Acclimatization in *Apodemus chevrieri*

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## ABSTRACT

Physiological adjustments to seasonal variations of the environment, such as changes in body mass and energy metabolism, are important adaptive strategies for small mammals. Ambient cues, as temperature and photoperiod, play important roles in animals' physiological adjustments. In order to determine the contributions of temperature and photoperiod to seasonal changes in body mass and thermogenesis in *Apodemus chevrieri*, we examined seasonal-acclimatized and lab-acclimated animals on several physiological, hormonal, and biochemical markers indicative of thermogenic capacity. The results showed that *A. chevrieri* adapt to winter or under cold condition by increasing thermogenesis and food intake. The present results suggest that the observed physiological regulation from the organismal, hormonal levels to the cellular levels of this alpine small mammal were important and allowed *A. chevrieri* to overcome the physiological challenges of an change to cold environment in winter successfully with the seasonal variations. Cold can induce an increase in mitochondrial protein contents and COX activity both in liver and brown adipose tissue, suggesting that *A. chevrieri* was more sensitive to cold than that of photoperiod. Together, these data suggested that *A. chevrieri* mainly depend on increasing thermogenic capacity to cope with cold or winter condition.

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## Authors' Contributions

W-L Z conceived and designed the study and wrote the article. GW executed the experimental work and participated in preparation of the manuscript.

## Key words

Bioenergetics, Thermogenesis, *Apodemus chevrieri*, Temperature, Photoperiod

## INTRODUCTION

Physiological adjustments, such as changes of body mass and energy metabolism, to seasonal variations of the environment are important adaptive strategies for small mammals (Gaston, 2009; Gaston and Spicer, 2001). It is well known that small mammals possess physiological adaptive strategies to cope with variations in the environment such as photoperiod and/or temperature changes (Heldmaier *et al.*, 1982). However, endotherms require appropriate responses and efficient thermoregulatory mechanisms (Klingenspor *et al.*, 2000) to regulate body mass and thermogenic capacity (nonshivering thermogenesis, NST) to cope with winter-like conditions (Chi and Wang, 2011; Concannon *et al.*, 2001; Król and Speakman, 2007; Li and Wang, 2005; Zhang *et al.*, 2011; Zhang and Wang, 2007; Zhu *et al.*, 2010; Nieminen and Hyvarinen, 2000). Liver metabolism accounts for 20–25% of RMR (Couture and Hulbert, 1995). Brown adipose tissue (BAT) is the structure mostly responsible for NST during cold climate or winter (Janský, 1973; Lowell and Spiegelman, 2000), during which the NST increases through a process that involves hypothalamic-pituitary-thyroid axis and

sympathetic nervous system (Cannon and Nedergaard, 2004). BAT capacity to produce NST entirely depends on uncoupling protein 1 (UCP1) (Cannon and Nedergaard, 2004). As the cytochrome *c* oxidase (COX) represents the terminal enzyme of oxidative phosphorylation in mitochondria, and is involved in mitochondrial energy metabolism (Kadenbach *et al.*, 2000). Triiodothyronine ( $T_3$ ), affects almost every physiological process in the body, including growth, development and metabolism (Kelly and Lieberman, 2009). Thyroxine ( $T_4$ ) is tyrosine-based hormone produced by the thyroid gland that is primarily responsible for regulation of metabolism (Kirkegaard and Faber, 1998). Thyroid-stimulating hormone (TSH) is a hormone that stimulates the thyroid gland to produce  $T_4$ , and then  $T_3$  which stimulates the metabolism of almost every tissue in the body (Parmentier *et al.*, 1989).

*Apodemus chevrieri* is a inherent species in Hengduan mountain region (Zhu *et al.*, 2016). Previous studies had demonstrated the presence of a seasonal variation in digestive tract morphology in *A. chevrieri* (Zhu *et al.*, 2012a). Specifically, cold environment reduced serum leptin levels but enhance thermogenic capacity in *A. chevrieri* (Zhu *et al.*, 2011), whereas the short photoperiod enhance thermogenic capacity (Zhu *et al.*, 2013a). However, we know nothing about seasonal changing, both temperature and photoperiod influenced its physiological adaptation from the hormonal levels to

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the cellular levels. The aim of the present study was to determine the response of body mass, enzyme activity and hormone concentrations in *A. chevrieri* under seasonal acclimatized and lab acclimated. We hypothesized that *A. chevrieri* showed seasonal changes of body mass, and temperature and photoperiod may play roles in its body mass regulation. We predicted that *A. chevrieri* would show significant seasonal changes in body mass, enzyme activity and hormone concentrations among different seasons, and temperature and photoperiod play important roles in its physiological adjustments.

## MATERIALS AND METHODS

### Samples

*A. chevrieri* were captured in farmland (26°15′–26°45′N; 99°40′–99°55′E; altitude 2,590m) in Jianchuan County, Yunnan province. All animals were healthy adults. *A. chevrieri* was housed individually in a wire cage (350×300×250mm<sup>3</sup>). Food and water were provided *ad libitum*. All pregnant, lactating or young individuals were excluded. All animal procedures were licensed under the Animal Care and Use Committee of School of Life Sciences, Yunnan Normal University (Permit No.: 13-0901-011).

### Experiment 1

The experiment was performed in 2014–2015. *A. chevrieri* were wild-captured in mid-July and October 2014, mid-January and late April 2015 (referred to as the summer, autumn, winter and spring groups hereafter). Between capture and food intake analysis, the animals were kept individually in plastic cages (350×300×250 mm) in a room with natural temperature and photoperiod, spring (5.4°C), summer (23.9°C), fall (16.6°C) and winter (−3.8°C). After the measurement of body mass and food intake, subjects were sacrificed by puncture of the posterior vena cava within 4 days after capture for all the four seasons, and blood and tissue samples were taken for measurement of physiological parameters (Wang *et al.*, 2006). A total of 40 adult *A. chevrieri* were used in the present study (summer, n=10; autumn, n=10; winter, n=10; spring, n=10). Between capture and metabolic analysis, the animals were kept individually in a metabolic cage (40×40×40cm<sup>3</sup>) in a room with natural temperature and photoperiod. Food intake was calculated as the mass of food missing from the hopper, subtracting orts mixed in the bedding.

### Experiment 2

Animals used in Experiment 2 were offspring of adult

*A. chevrieri*, that were captured in mid-July 2014 and then transported to School of Life Sciences of Yunnan Normal University, housed in plastic box (350×300×250mm<sup>3</sup>) individually. In order to test singularly or associate the effects of ambient temperature, photoperiod on mitochondrial protein content and COX activity and serum hormones, 40 adult weight-matched *A. chevrieri* were housed individually (were maintained at 12L: 12D (light on at 08:00am), 25±1°C, respectively), and kept for at least 2 weeks to familiarize with the environment. After the acclimatizing period, the animals were randomly assigned to the following four groups: long photoperiod (LD, 16L: 8D) cold (5±1°C), LD, and warm (30±1°C), short photoperiod (SD, 8L: 16D) cold, SD and warm; each group included 10 individuals. Animals were acclimated for 4 weeks. Food intake was calculated as the mass of food missing from the hopper, subtracting orts mixed in the bedding.

### Measurement of mitochondria respiration

After the measurements of food intake (Zhu *et al.*, 2010), animals were killed between 1200 and 1400 by decapitation and blood was collected for hormone determination. The blood was centrifuged at 4,000 rpm for 30 min after a 30-min interval and the serum was collected and stored at −72°C for hormone determination. Liver, BAT and testicular were carefully and quickly removed and weighted (0.1mg), and their adhering tissues separated. Liver and BAT were blotted, weighed, and placed in ice-cold sucrose-buffered medium and then homogenized for the isolation of mitochondria (Cannon and Lindberg, 1979). The protein content of mitochondria was determined by the Folin phenol method with bovine serum albumin as standard (Lowry *et al.*, 1951). State 4 (ST<sub>4</sub>) of mitochondrial respiration of liver and BAT were measured by Hansatech Oxy-Lab Chloroab 2 oxygen electrode (Hansatech Instruments LTD., England).

### Measurement of enzyme activity

The COX (EC 1.9.3.1) activity of liver and BAT was measured with polarographic method using oxygen electrode (Hansatech Instruments LTD., England) (Sundin *et al.*, 1987), and the  $\alpha$ -glycerophosphate oxidase ( $\alpha$ -PGO; EC 1.1.3.21) was determined polarographically according to Steffen and Roberts (Steffen and Roberts, 1977). Thyroxine 5′-deiodinase (T<sub>4</sub> 5′-DII; EC 1.97.1.10) activity in BAT was assayed as previously described (Leonard *et al.*, 1983).

### Measurements of hormone concentration

The concentrations of triiodothyronine (T<sub>3</sub>), thyroxine (T<sub>4</sub>), Thyroid-stimulating hormone (TSH), and testosterone

(T) in the serum were determined by radioimmunoassay using RIA kits (China Institute of Atomic Energy). These kits were validated for all species tested by cross-activity. Intra- and inter-assay coefficients of variation were 2.4% and 8.8% for the  $T_3$ , 4.3% and 7.6% for  $T_4$ , 3.6% and 6.9% for TSH, and 7.6% and 8.1% for T, respectively.

#### Statistical analysis

Data were analyzed using SPSS 16.0 software (SPSS Inc., Chicago, IL, USA). Prior to all statistical analyses, data were examined for assumptions of normality and homogeneity of variance using the Kolmogorov–Smirnov and Levene tests, respectively. Seasonal data such as COX activity, mitochondrial protein content and thyroid hormones were analyzed by one-way analysis of variance (ANOVA). Two-way ANCOVA was used to detect the effect of photoperiod and temperature on thermogenic properties in liver, BAT and hormones, using body mass as the covariate. Differences among groups were detected by Duncan. Since no gender effects were found on almost

all measured parameters, data from females and males were combined. Results are presented as mean  $\pm$  SE (n sample size) in the test, and  $P < 0.05$  was considered to be statistically significant.

## RESULTS

#### *Changes in mitochondrial protein content and COX activity*

There was a significant difference in the absolute mass of BAT among different seasons ( $F=5.32$ ,  $P < 0.05$ , Table I). The mitochondrial protein content (M<sub>P</sub>) and BAT COX activity varied significantly among seasons (M<sub>P</sub>:  $F=2.35$ ,  $P < 0.01$ ; COX:  $F=6.54$ ,  $P < 0.01$ ). The M<sub>P</sub> and COX activity in winter were significantly higher than that of other seasons. There was a significant difference in the absolute mass of liver ( $F=4.59$ ,  $P < 0.01$ , Table I). The M<sub>P</sub> and COX activity in liver varied significantly among seasons (M<sub>P</sub>:  $F=6.25$ ,  $P < 0.01$ ; COX:  $F=5.89$ ,  $P < 0.01$ , Table I). The M<sub>P</sub> and COX activity in winter were markedly higher than that of other seasons.

**Table I.- Thermogenic properties in liver, brown adipose tissue and hormones in *Apodemus chevrieri* under different seasons**

	Spring(n=10)	Summer(n=10)	Autumn(n=10)	Winter(n=10)
Body mass	31.23 $\pm$ 2.8 <sup>b</sup>	38.65 $\pm$ 3.8 <sup>a</sup>	36.24 $\pm$ 1.6 <sup>a</sup>	30.01 $\pm$ 2.3 <sup>b</sup>
<b>Brown adipose tissue</b>				
Mass (g)	0.20 $\pm$ 0.07 <sup>a</sup>	0.13 $\pm$ 0.07 <sup>b</sup>	0.12 $\pm$ 0.06 <sup>b</sup>	0.23 $\pm$ 0.05 <sup>a</sup>
% body mass	0.58 $\pm$ 0.02 <sup>b</sup>	0.34 $\pm$ 0.02 <sup>c</sup>	0.33 $\pm$ 0.01 <sup>c</sup>	0.77 $\pm$ 0.04 <sup>a</sup>
TP (mg·g <sup>-1</sup> )	226.41 $\pm$ 11.23 <sup>a</sup>	165.39 $\pm$ 8.97 <sup>b</sup>	176.59 $\pm$ 8.02 <sup>b</sup>	236.69 $\pm$ 10.81 <sup>a</sup>
M <sub>P</sub> (mg·g <sup>-1</sup> )	32.21 $\pm$ 1.94 <sup>b</sup>	26.92 $\pm$ 1.02 <sup>c</sup>	27.58 $\pm$ 0.84 <sup>c</sup>	39.98 $\pm$ 2.95 <sup>a</sup>
ST <sub>4</sub> (nmol O <sub>2</sub> mg <sup>-1</sup> MtP min <sup>-1</sup> )	35.89 $\pm$ 3.45 <sup>a</sup>	28.36 $\pm$ 3.15 <sup>b</sup>	29.11 $\pm$ 2.36 <sup>b</sup>	36.95 $\pm$ 4.26 <sup>a</sup>
COX (μg atoms O <sub>2</sub> min <sup>-1</sup> mg <sup>-1</sup> MtP)	84.63 $\pm$ 5.12 <sup>b</sup>	54.33 $\pm$ 3.25 <sup>c</sup>	58.69 $\pm$ 4.69 <sup>c</sup>	95.41 $\pm$ 5.21 <sup>a</sup>
T <sub>4</sub> 5'-DII (pmol O <sub>2</sub> ·mg <sup>-1</sup> ·MtP min <sup>-1</sup> )	34.38 $\pm$ 2.51 <sup>a</sup>	24.61 $\pm$ 2.23 <sup>b</sup>	25.64 $\pm$ 1.78 <sup>b</sup>	38.45 $\pm$ 2.36 <sup>a</sup>
<b>Liver</b>				
Mass (g)	2.32 $\pm$ 0.16 <sup>b</sup>	1.67 $\pm$ 0.22 <sup>c</sup>	1.73 $\pm$ 0.21 <sup>c</sup>	2.78 $\pm$ 0.16 <sup>a</sup>
% body mass	7.42 $\pm$ 0.56 <sup>a</sup>	4.32 $\pm$ 0.15 <sup>b</sup>	4.77 $\pm$ 0.35 <sup>b</sup>	9.26 $\pm$ 0.95 <sup>a</sup>
TP (mg·g <sup>-1</sup> )	231.25 $\pm$ 12.47 <sup>a</sup>	182.81 $\pm$ 13.26 <sup>b</sup>	186.59 $\pm$ 12.64 <sup>b</sup>	237.52 $\pm$ 15.23 <sup>a</sup>
M <sub>P</sub> (mg·g <sup>-1</sup> )	49.61 $\pm$ 3.21 <sup>b</sup>	35.41 $\pm$ 3.95 <sup>c</sup>	36.11 $\pm$ 2.22 <sup>c</sup>	55.33 $\pm$ 4.11 <sup>a</sup>
ST <sub>4</sub> (nmol O <sub>2</sub> mg <sup>-1</sup> MtP min <sup>-1</sup> )	37.27 $\pm$ 2.96 <sup>a</sup>	25.57 $\pm$ 1.61 <sup>b</sup>	26.98 $\pm$ 1.84 <sup>b</sup>	39.25 $\pm$ 3.48 <sup>a</sup>
COX (μg atoms O <sub>2</sub> min <sup>-1</sup> mg <sup>-1</sup> MtP)	69.24 $\pm$ 3.34 <sup>b</sup>	56.64 $\pm$ 4.12 <sup>c</sup>	60.14 $\pm$ 2.36 <sup>c</sup>	75.02 $\pm$ 2.42 <sup>a</sup>
<b>Hormones</b>				
Tri-iodothyronine (T <sub>3</sub> , ng·ml <sup>-1</sup> )	1.25 $\pm$ 0.11 <sup>ab</sup>	0.85 $\pm$ 0.05 <sup>c</sup>	0.88 $\pm$ 0.13 <sup>b</sup>	1.36 $\pm$ 0.12 <sup>a</sup>
Thyroxine (T <sub>4</sub> , ng·ml <sup>-1</sup> )	13.25 $\pm$ 0.78 <sup>a</sup>	14.26 $\pm$ 0.32 <sup>a</sup>	14.33 $\pm$ 0.16 <sup>a</sup>	14.69 $\pm$ 0.51 <sup>a</sup>
T <sub>3</sub> /T <sub>4</sub> (×100)	9.43 $\pm$ 0.34 <sup>a</sup>	5.96 $\pm$ 0.15 <sup>b</sup>	6.14 $\pm$ 0.12 <sup>b</sup>	9.26 $\pm$ 0.61 <sup>a</sup>
Thyroid-stimulating hormone (TSH, ng·ml <sup>-1</sup> )	0.91 $\pm$ 0.05 <sup>a</sup>	0.88 $\pm$ 0.07 <sup>a</sup>	0.89 $\pm$ 0.05 <sup>a</sup>	0.92 $\pm$ 0.09 <sup>a</sup>

Different superscripts in each row means significantly different ( $P < 0.05$ ).

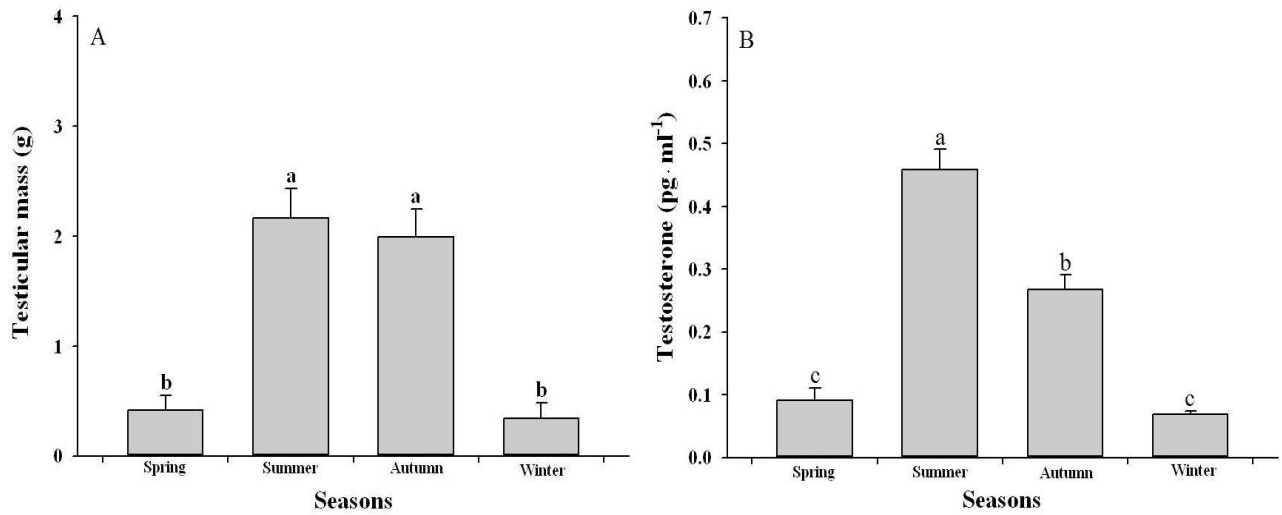


Fig. 1. Seasonal variations of testicular mass (A) and testosterone (B) in *Apodemus chevrieri*. Different superscripts in each row means significantly different ( $P<0.05$ ).

#### Changes in serum hormones

Seasonal alterations were also detected in testicular mass ( $F=5.36$ ,  $P<0.01$ , Fig. 1A), which was significantly lower in winter than that in summer. Seasonal alterations were also detected in  $T_3$  and testosterone (T) levels ( $T_3$ :  $F=5.35$ ,  $P<0.01$ , Table I; T:  $F=6.16$ ,  $P<0.01$ , Fig. 1B). Serum  $T_4$  and TSH levels gradually decreased from warmer to colder months, but showed no significant differences. Food intake showed significant differences among different seasons (Fig. 2).

#### Effects of photoperiod and/or temperature on mitochondrial protein content and COX activity

Both photoperiod and temperature, or by the interaction of the two parameters had significant effects on  $ST_4$  and COX activity in liver (Table II). In BAT, TP contents, MP contents,  $ST_4$  levels, COX activity,  $T_4$  5'-DII activity and UCP 1 contents were affected by both photoperiod and temperature, or by the interaction of the two parameters (Table II). But BAT mass was influenced significantly only by temperature (photoperiod:  $F=0.64$ ,  $P>0.05$ ; temperature:  $F=4.21$ ,  $P<0.01$ , interaction:  $F=0.95$ ,  $P>0.05$ , Table II).

#### Effect of photoperiod and/or temperature on serum hormones

Temperature, but not photoperiod or the interaction of photoperiod and temperature, had a significant effect on  $T_4$  levels for *A. chevrieri* (Table II). Temperature or photoperiod and the interaction of both parameters had a significant effect on  $T_3$  levels for *A. chevrieri* (photoperiod:  $F=4.31$ ,  $P<0.01$ ; temperature:  $F=5.45$ ,  $P<0.01$ ; interaction:

$F=2.85$ ,  $P<0.05$ , Table II). Temperature and photoperiod had significant effects on TSH levels for *A. chevrieri*, but no effect of the interaction of photoperiod and temperature was found (photoperiod:  $F=2.92$ ,  $P<0.05$ ; temperature:  $F=4.25$ ,  $P<0.05$ ; interaction:  $F=0.225$ ,  $P>0.05$ , Table II).

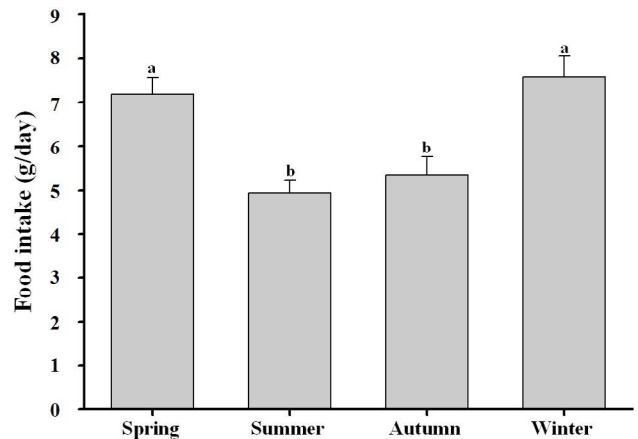


Fig. 2. Seasonal variations of food intake in *Apodemus chevrieri*.

Different superscripts in each row means significantly different ( $P<0.05$ ).

## DISCUSSION

Changes in body mass and energy metabolism are important adaptive strategies for small mammals under seasonal variations of the environment. Ambient cues,

**Table II.- Influences of photoperiod and/or temperature on thermogenic properties in liver, brown adipose tissue and hormones in *Apodemus chevrieri*.**

	Short photoperiod		Long photoperiod	
	5°C (n=10)	30°C (n=10)	5°C (n=10)	30°C (n=10)
Body mass	26.35±1.54 <sup>c</sup>	32.36±1.21 <sup>a</sup>	29.35±0.89 <sup>b</sup>	35.24±1.25 <sup>a</sup>
Food intake(g/d)	9.23±1.02 <sup>a</sup>	6.25±0.54 <sup>b</sup>	8.62±0.82 <sup>a</sup>	6.03±0.46 <sup>b</sup>
<b>Brown adipose tissue</b>				
Mass (g)	0.23±0.02 <sup>a</sup>	0.17±0.04 <sup>b</sup>	0.21±0.05 <sup>a</sup>	0.16±0.03 <sup>b</sup>
% body mass	0.97±0.02 <sup>a</sup>	0.52±0.01 <sup>b</sup>	0.53±0.02 <sup>b</sup>	0.45±0.01 <sup>c</sup>
TP (mg·g <sup>-1</sup> )	236.68±23.65 <sup>a</sup>	156.32±10.23 <sup>c</sup>	212.32±15.32 <sup>b</sup>	145.35±11.02 <sup>c</sup>
M <sub>I</sub> P (mg·g <sup>-1</sup> )	37.12±1.81 <sup>a</sup>	18.65±1.12 <sup>c</sup>	32.1±1.91 <sup>b</sup>	16.73±1.56 <sup>c</sup>
ST <sub>4</sub> (nmol O <sub>2</sub> ·mg <sup>-1</sup> MtP min <sup>-1</sup> )	45.25±2.36 <sup>a</sup>	28.36±1.54 <sup>b</sup>	43.78±4.21 <sup>a</sup>	25.36±2.21 <sup>c</sup>
COX (μg atoms O <sub>2</sub> ·min <sup>-1</sup> ·mg <sup>-1</sup> MtP)	102.36±9.56 <sup>a</sup>	70.02±6.35 <sup>c</sup>	94.36±6.35 <sup>b</sup>	65.32±4.21 <sup>c</sup>
T <sub>4</sub> 5'-DII (pmol O <sub>2</sub> ·mg <sup>-1</sup> MtP min <sup>-1</sup> )	31.21±2.35 <sup>a</sup>	18.54±2.36 <sup>c</sup>	35.26±2.12 <sup>b</sup>	16.98±1.92 <sup>c</sup>
<b>Liver</b>				
Mass (g)	2.06±0.53 <sup>a</sup>	1.58±0.22 <sup>c</sup>	1.89±0.34 <sup>b</sup>	1.32±0.33 <sup>c</sup>
% body mass	7.81±0.25 <sup>a</sup>	4.88±0.26 <sup>c</sup>	6.43±0.33 <sup>b</sup>	3.74±0.21 <sup>d</sup>
TP (mg·g <sup>-1</sup> )	231.23±11.25 <sup>a</sup>	88.61±7.93 <sup>c</sup>	202.36±11.63 <sup>b</sup>	84.36±6.32 <sup>c</sup>
M <sub>I</sub> P (mg·g <sup>-1</sup> )	43.72±5.31 <sup>a</sup>	28.51±6.42 <sup>b</sup>	40.91±3.51 <sup>a</sup>	27.11±4.12 <sup>b</sup>
ST <sub>4</sub> (nmol O <sub>2</sub> ·mg <sup>-1</sup> MtP min <sup>-1</sup> )	40.67±2.31 <sup>a</sup>	22.51±3.21 <sup>c</sup>	39.72±4.11 <sup>b</sup>	18.75±3.72 <sup>c</sup>
COX (μg atoms O <sub>2</sub> ·min <sup>-1</sup> ·mg <sup>-1</sup> MtP)	85.61±10.11 <sup>a</sup>	55.82±9.82 <sup>c</sup>	78.52±8.42 <sup>b</sup>	52.91±4.82 <sup>d</sup>
<b>Hormones</b>				
Tri-iodothyronine (T <sub>3</sub> , ng·ml <sup>-1</sup> )	1.53±0.12 <sup>a</sup>	0.52±0.02 <sup>b</sup>	1.23±0.06 <sup>a</sup>	0.51±0.03 <sup>b</sup>
Thyroxine (T <sub>4</sub> , ng·ml <sup>-1</sup> )	13.53±0.94 <sup>a</sup>	6.47±0.58 <sup>c</sup>	11.41±1.12 <sup>b</sup>	6.24±0.56 <sup>c</sup>
T <sub>3</sub> /T <sub>4</sub> (×100)	11.30±0.12 <sup>a</sup>	8.03±0.15 <sup>c</sup>	10.78±0.13 <sup>b</sup>	8.17±0.16 <sup>c</sup>
Thyroid-stimulating hormone (TSH, ng·ml <sup>-1</sup> )	1.18±0.15 <sup>a</sup>	0.77±0.14 <sup>b</sup>	0.89±0.11 <sup>b</sup>	0.69±0.12 <sup>c</sup>

Different superscripts in each row means significantly different ( $P<0.05$ ).

such as temperature and photoperiod, play important roles in animals' physiological adjustments (Zhang and Wang, 2006). Data from our study suggested that *A. chevrieri* showed seasonal adaptations of body mass regulation. For example, thermogenic capacity to adapt to winter like conditions (increased of COX activity with cold and short photoperiod), regulation of the appetite (increased food intake), variation of modification of biochemical and hormonal markers indicated that animals' abilities to adapt the variations of the environment in its natural habitat. In particular, temperature and photoperiod were important factors that influences the enzyme activity and hormone concentrations in *A. chevrieri*.

#### Seasonal variations in biochemical and hormonal markers

In the prior reports, *A. chevrieri* showed an decrease in body mass in winter (Zhu *et al.*, 2012a). In the current

study, *A. chevrieri* also showed a significant seasonal variation in body mass, which was highest in summer and lowest in winter. Similar results were found in other species, such as *Microtus maximowiczii* (Chen *et al.*, 2012), *M. pennsylvanicus* (Dark and Zucker, 1986), *M. ochrogaster* (Kriegsfeld and Nelson, 1996) and *A. draco* (Zhu *et al.*, 2012b). Small mammals mainly increase thermogenic capacity to cope with cold stress (Heldmaier *et al.*, 1982). In the present study, there were significant seasonal variations in several biochemical markers indicative of thermogenic capacity, which was supported by the M<sub>I</sub>P contents, the COX activity in liver and BAT, showing a consistency with the findings of previous studies on *Ochotona curzoniae* (Wang *et al.*, 2006), *Acomys russatus* (Kronfeld-Schor *et al.*, 2000) and *A. draco* (Zhu *et al.*, 2013b). T<sub>4</sub>, the primary thyroid-product, is relatively inactive until deiodinated to T<sub>3</sub> (Tomasi, 1991), and TSH stimulates circulating concentrations of T<sub>3</sub> and T<sub>4</sub> (Flier



*et al.*, 2000). Thyroid hormones are major modulators of cold-induced NST (Silva, 2006):  $T_3$  is the only active form of thyroxines, and mainly affect the expression of UCP1 (Cannon and Nedergaard, 2004). In the present study,  $T_4$  5'-DII activity in winter was significantly higher than that in summer, it is responsible for the production of  $T_4$  by thyroid gland and its transformation in the peripheral tissues into  $T_3$ .  $T_3$  concentration in winter was significantly higher than that in summer, associated with seasonal changes in NST. In *A. chevrieri*, the testosterone concentration and testicular mass in summer and autumn (the breeding seasons) were significantly higher than in winter, suggesting the possible use in protecting the territory and fighting for mates (Soto-Gamboa *et al.*, 2005). Our findings may indicate that variations of thermogenesis may be not only subjected by endocrine regulation, but also depended on the changes of the physiology of breeding. This adjustment mechanism may be supported with "challenge hypothesis", which predicts that seasonal patterns in testosterone levels are a function of mating system (monogamy versus polygyny), paternal care, and male-male aggression in seasonal breeders (Wingfield *et al.*, 1990).

#### *Effects of photoperiod and/or temperature in enzyme activity and serum hormones*

In prior reports had been shown that both short photoperiod or cold temperature are important environmental cues that can influence the decrease in body mass, increase in food intake and thermogenic capacity in *A. chevrieri* (Zhu *et al.* 2012a). As a key element of the adipostatic mechanism for regulating body mass were influenced by both short photoperiods and cold in *A. chevrieri* (Table II). The variations in thermogenesis at individual level were supported by biochemical markers such as MP content and COX activity. We found differences in serum  $T_3$  and  $T_4$  contents, the serum  $T_3$  and  $T_4$  were higher in short photoperiod and cold group than that in long photoperiod and warm group, then short photoperiod and cold induced increases in TSH level (Table II). We also found that the  $T_4$  5'-DII was higher in the short photoperiod in cold group than that in other groups.  $T_3$  and  $T_4$  concentration is associated with the activity of  $T_4$  5'-DII. In fact, TSH may indicate that the process of transformation of  $T_4$  into  $T_3$ , and promoted by  $T_4$  5'-DII activity increasing, with a consequent higher sensitivity of *A. chevrieri* to cold than that of photoperiod.

## CONCLUSIONS

In conclusion, environmental factors play key roles in the regulation of seasonal adaptation of body mass, hormonal and biochemical markers, and food intake in *A.*

*chevrieri*. The present results suggested that the observed physiological regulation from the organismal, hormonal levels to the cellular levels of this alpine small mammal were important and allowed *A. chevrieri* to overcome the physiological challenges of an change to cold environment in winter successfully with the seasonal variations. Together, these data suggested that *A. chevrieri* mainly depend on increasing food take and thermogenic capacity to cope with cold or winter condition.

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#### *Competing interests*

The authors declare that they have no competing interests.

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