



Short Communication

Genetic Diversity Analysis of Four Bactrian Camel Varieties in China

J.Y. Bai^{1,*}, R.T.D. Wu^{2,*}, Q. Zhang², D. Bao², L.M. Dao² and X.H. Tian³

¹College of Animal Science and Technology, Henan University of Science and Technology, Luoyang 471023, China

²Institute of Animal, Alxa of Inner Mongolia, Bayanhaote 750306, China

³School of Life Sciences, Henan University, Kaifeng 475000, China

ABSTRACT

In this study, 17 microsatellite markers were used to analyze the genetic polymorphism, genetic differentiation, gene flow and genetic distance of Alashan desert Bactrian camel, Alashan Gobi Bactrian camel, Sunite Bactrian camel and Qinghai Bactrian camel. The results indicated that the number of effective alleles of the four Bactrian camel varieties ranged from 2.7302 to 3.0524, and average theoretical heterozygosity and average polymorphic information content were 0.6283 and 0.5546, respectively. Observational heterozygosity, expected heterozygosity and polymorphic information content of Qinghai Bactrian camel were all higher than those of the other three varieties, being 0.8922, 0.6490 and 0.5813, respectively, so Qinghai Bactrian camel was of rich genetic polymorphism. Average gene flow of the microsatellite markers was 12.3188 and average F_{st} value was 0.0199, namely 1.99% of genetic variation derived between subpopulations and 98.01% came inside the subpopulations, revealing that the genetic differentiation degree between Bactrian camel subpopulations was low. The genetic relationship between Sunite Bactrian camel and Alashan Gobi Bactrian camel was close, so it was classified into the first type and that between Sunite Bactrian camel and Alashan desert Bactrian camel as the second type, but the genetic relationship of Qinghai Bactrian camel and other three Bactrian camel varieties was distant.

Article Information

Received 06 November 2019

Revised 26 December 2019

Accepted 03 January 2020

Available online 04 September 2020

Authors' Contribution

JYB conceived and designed the study and conducted the lab work. RTDW and QZ analyzed the data and wrote the article. DB and LMD helped in sampling. XHT helped in analysis of data.

Key words

Bactrian camels, Microsatellite marker, Genetic diversity, Gene flow, Polymorphic information content

Bactrian camel, called “ship of the desert”, and also called camel in China, is a special variety formed through the long-time natural selection. Bactrian camels have been tamed by human beings long before, which are docile, easy to ride and suitable for carrying loads, so they are usually used as tools for riding instead of walk among the people in desert areas, and meanwhile, they can provide livestock products such as meat, milk and fur. Furthermore, they have played a significant role in the human development and desert conquering. In recent years, domestic and foreign researches regarding genetic diversity (Hedayat-Evrigh, *et al.*, 2018) and organization structure (Ye *et al.*, 2014a, 2014b; Wang *et al.*, 2016) of Bactrian camels have achieved progress. Microsatellite markers have been extensively applied to genetic diversity studies of cow (Ni *et al.*, 2018), sheep (Bai *et al.*, 2015) and poultry (Bai *et al.*, 2014, 2016a, 2016b, 2016c, 2017) by virtue of high abundance, good repeatability, co-dominance marker and selective neutrality. It is also widely used in

plant genetic diversity research (Guo *et al.*, 2018; Yang *et al.*, 2013). Polyacrylamide gel electrophoresis method was used in this study to detect the polymorphism of the 17 microsatellite markers in four Bactrian camel varieties in China, expecting to provide a theoretical basis for the protection of Bactrian camel variety resources and improvement of population productivity.

Materials and methods

Blood samples (10 ml) were taken from jugular vein 40 Sunite Bactrian Camels, 40 Alashan desert Bactrian camel, 40 Alashan gobi Bactrian camel and 40 Qinghai Bactrian camel. ACD was added for anticoagulation. DNA was extracted from the blood using the whole-blood genomic DNA extraction kit (Beijing Dingguo Changsheng) method and stored at -20°C. Seventeen microsatellite markers with high polymorphism were screened (Evdotchenko *et al.*, 2003; Sushma *et al.*, 2014). The primers were synthesized by Shanghai Shengong Bioengineering Technology Service Co., Ltd.

Thermal cycle for PCR comprised pre-denaturation at 94°C for 4 min, then denaturation at 94°C for 40 s, annealing at 60°C for 1 min, annealing at 72°C for 20 s, denaturation, annealing and elongation were carried out

* Corresponding author: byezi@163.com; Wuren1223@vip.sina.com

0030-9923/2021/0001-0001 \$ 9.00/0

Copyright 2021 Zoological Society of Pakistan

Table I. Population genetic diversity of four Bactrian camels.

Population	Na	Ne	O_Hom	O_Het	E_Hom	E_Het	PIC
Alashan Desert Bactrian camel	3.8235	2.8367	0.1309	0.8691	0.3908	0.6192	0.5461
Alashan Gobi Bactrian camel	3.5294	2.7302	0.1403	0.8597	0.3932	0.6068	0.5308
Qinghai Bactrian camel	3.8235	3.0524	0.1078	0.8922	0.3510	0.6490	0.5813
Sunite Bactrian camel	3.7647	2.8913	0.1235	0.8765	0.3618	0.6382	0.5602

Author: Please explain abbreviations used in this table.

Table II. Fixed index and gene flow estimation.

Microsatellite markers	Fis (inbreeding coefficient of total population)	Fit (inter-population differentiation coefficient)	Fst (intra-population inbreeding coefficient)	Nm (gene flow)
LCA33	-0.3437	-0.3362	0.0056	44.5645
LCA37	-0.8362	-0.8300	0.0034	73.1873
LCA63	-0.4801	-0.4738	0.0043	58.2326
LCA66	-0.3643	-0.3294	0.0256	9.5282
LCA71	-0.7024	-0.6869	0.0091	27.1327
LCA82	-0.5725	-0.5545	0.0114	21.6694
LCA90	-0.2459	-0.2114	0.0277	8.7863
CMS15	-0.4746	-0.4049	0.0473	5.0378
CMS18	-0.5894	-0.5846	0.0030	83.1109
CMS36	0.8525	0.8584	0.0401	5.9919
CMS104	-0.4715	-0.4208	0.0345	7.0058
CVRL101	-0.3397	-0.3263	0.0100	24.7006
YWLL29	-0.1197	-0.0927	0.0241	10.1027
YWLL36	-0.6685	-0.6201	0.0290	8.3595
YWLL44	-0.4361	-0.3897	0.0323	7.4981
VOLP08	-0.3489	-0.3478	0.0009	291.4601
VOLP32	-0.3702	-0.3281	0.0307	7.8965
Mean	-0.4093	-0.3813	0.0199	12.3188

Table III. Genetic distance and coherence of four Bactrian camel populations.

Populations	Alashan Desert Bactrian camel	Qinghai Bactrian camel	Sunite Bactrian camel	Alashan Gobi Bactrian camel
Alashan Desert Bactrian camel		0.9553	0.9563	0.9607
Qinghai Bactrian camel	0.0458		0.9408	0.9545
Sunite Bactrian camel	0.0446	0.0610		0.9695
Alashan Gobi Bactrian camel	0.0401	0.0466	0.0310	

Note: The upper triangle is genetic consistency and the lower triangle is Nei's genetic distance.

for 35 cycles, then elongation at 72°C and finally the reaction was completed and cooled and preserved at 4°C.

For SSCP, 15% non denaturing polyacrylamide gels were used to detect the products. Silver nitrate dyeing method is used for dyeing, mainly through fixation, oxidation, dyeing, color rendering, photography and other links.

Popgene32 software was used to calculate numbers

of effective alleles, allele frequencies and heterozygosity of microsatellites.

Results and discussion

Genetic polymorphisms of the 17 microsatellite markers in the four Bactrian camel varieties are seen in [Table I](#). It could be seen that number of effective alleles of the four varieties ranged from 2.7302 to 3.0524.

Observational heterozygosity, expected heterozygosity and polymorphic information content of Qinghai Bactrian camel were all higher than those of the other three varieties, being 0.8922, 0.6490 and 0.5813, respectively, so Qinghai Bactrian camel was of rich genetic polymorphism.

Fixed indices and gene flows of the 17 microsatellite markers in the four Bactrian camel populations are seen in Table II according to which F_{is} and F_{ts} values of microsatellites are negative except for CMS36. The average F_{is} and F_{ts} values were -0.409 and -0.3813, respectively. According to Wight (1978), if the population F_{st} value is within 0-0.05, no differentiation exists between the subpopulations. If F_{st} value is within 0.05-0.15, moderate differentiation is considered. If it is between 0.15-0.25, high differentiation is manifested. Average F_{st} value of the microsatellite markers in this study was 0.0199, namely 1.99% of genetic variation derived between subpopulations and 98.01% came inside the subpopulations, revealing that the genetic differentiation degree between Bactrian camel subpopulations was low or even no differentiation existed. Gene flows of the microsatellite markers were large with average value of 12.3188. The gene flow values in this study were larger than the study result by Tian *et al.* (2012) ($N_m=5.4869$). It was revealed in this study that gene exchange degree between the four Bactrian camel subpopulations in different areas was high, which resulted in their low genetic differentiation.

The genetic distance and identity values between the Bactrian camel subpopulations are shown in Table III. It could be observed that Nei's genetic distance between the subpopulations was small, ranging from 0.0310 to 0.0610, but the genetic identity was large (0.9408-0.9695). The cluster diagram of the four Bactrian camel populations is seen in Figure 1, which shows that the genetic relationship between Sunite Bactrian camel and Alashan Gobi Bactrian camel was close, so it was classified into the first type and that between Sunite Bactrian camel and Alashan desert Bactrian camel as the second type. The study by Mburu *et al.* (2003) indicates that 6 dromedary populations are divided into 2 isolated subpopulations. Gene exchange exists between the Bactrian resources, which is an important reason for the correlation between clustering result and the ecological area. This study indicated that genetic relationships of Sunite Bactrian camel, Alashan gobi Bactrian camel and Alashan desert Bactrian camel, which were all located in the Inner Mongolia Autonomous Region, were close due to the frequent gene exchange, and located in Gansu Province, Qinghai Bactrian camel had less frequent gene exchange with other 3 Bactrian camel varieties, so its genetic relationships with them were distant.

All camels in China are Bactrian camels which are

dominant livestock resources in desert areas. There are over 280,000 camels in China, where most of them are located in the Inner Mongolia Autonomous Region, accounting for about 67% of the total number, followed by those in Xinjiang Uygur Autonomous Region, accounting for about 20%. The protection of genetic polymorphism of Bactrian camel resources in the Inner Mongolia Autonomous Region not only refers to reasonably managing and utilizing the existing resources but also can maintain a certain resource potential for the future demand.

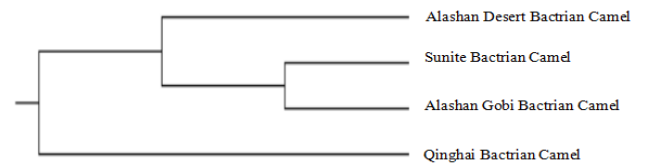


Fig.1. Genetic clustering of four Bactrian camel varieties.

Acknowledgements

Sincere gratitude goes to the sponsor of Alxa League Science and Technology Project in Inner Mongolia (2012-12).

Statement of conflict of interest

The author declares there is no conflict of interest.

References

- Bai, J.Y., Pang, Y.Z., Qi, Y.X., Zhang, X.H. and Yun, Y.X. 2017. *Indian J. Anim. Res.*, **51**: 851-855.
- Bai, J.Y., Pang, Y.Z., Wu, S.J., Yu, M.Q. and Zhang, X.H., 2016a. *Indian J. Anim. Res.*, **50**: 1-7.
- Bai, J.Y., Pang, Y.Z., Zhang, X.H., Yun, Y.X. and Qi, Y.X. 2016b. *Brazilian J. Poult. Sci.*, **18**: 519-524. <https://doi.org/10.1590/1806-9061-2015-0101>
- Bai, J.Y., Pang, Y.Z., Qi, Y.X., Zhang, X.H. and Yun, X.Y. 2016c. *Brazilian J. Poult. Sci.*, **18**: 27-32. <https://doi.org/10.1590/1806-9061-2015-0101>
- Bai, J.Y., Yang, Y.B., Wang, Y.Q., Zhang, X.H. and Pang, Y.Z., 2015. *Indian J. Anim. Res.*, **49**: 585-590.
- Bai, J.Y., Jia, X.P., Yang, Y.B., Zhang, X.H., Pang, Y.Z., Wang, Y.Q. and Qi, Y.X., 2014. *J. Anim. Pl. Sci.*, **24**: 965-968.
- Evdotchenko, D., Han, Y., Bartenschlager, H., Preuss, S. and Geldermann, H., 2003. *Mol. Ecol. Notes*, **3**: 431-434. <https://doi.org/10.1046/j.1471-8286.2003.00477.x>
- Guo, L.L., Guo, D.L., Zhao, W. and Hou, X.G., 2018. *J. Horticult. Sci. Biotechnol.*, **93**: 416-424. <https://doi.org/10.1080/14620316.2017.1373039>
- Hedayat-Evrigh, N., Miraei-Ashtiani, S.R., Shahrehabak,

- M.M., Evrigh, R.K. and Pourasad, K., 2018. *J. Agric. Sci. Technol.*, **20**: 1137-1148.
- Mburu, D.N., Ochieng. J.W., Kuria, S.G., Jianlin, H., Kaufmann, B., Rege, J.E. and Hanotte, O. 2003. *Anim. Genet.*, **34**: 26-32. <https://doi.org/10.1046/j.1365-2052.2003.00937.x>
- Ni, W.W., Jiang, A., Zhang, J., Ei, G.X. and Huang, Y.F., 2018. *Indian J. Anim. Res.*, **52**: 1543-1547.
- Sushma, P., Ali, S., Banerjee, P. and Joshi, J., 2014. *Int. J. Biomed. Life Sci.*, **5**: 286-296.
- Tian, Y.Z., Nuerbiya, W., Wang, L.J., Wu, W.W., Xu, X.M., Zhang, Y.H., Azhi, T., Tian, K.C., 2012. *Anim. Husb. Vet. Med.*, **44**: 38-43.
- Wang, H.J., Ma, K.M., Liu, Z.H., Jin, D.Z. and Wei, W.Q., 2016. *J. Camel Pract. Res.*, **23**: 241-246. <https://doi.org/10.5958/2277-8934.2016.00041.2>
- Wright, S., 1978. In: *Variability within and among natural populations*. University of Chicago Press, Chicago.
- Yang, J., Dai, P.F., Zhou, T.H., Huang, Z.H., Feng, L., Su H.L., Liu, Z.L. and Zhao, G.F. 2013. *Scient. Horticult.*, **150**: 1-10. <https://doi.org/10.1016/j.scienta.2012.11.004>
- Ye, W.L., Wang, F.L., Xie, Z.H., Wang, Y.G., Lin, B. and Wang, J.L., 2014a. *J. Camel Pract. Res.*, **21**: 191-198. <https://doi.org/10.5958/2277-8934.2014.00033.2>
- Ye, W.L., Xie, Z.H., Wang, F.L., Gen, X., Dong, S. and Wang, J.L. 2014b. *J. Camel Pract. Res.*, **21**: 103-109. <https://doi.org/10.5958/2277-8934.2014.00020.4>

Online First Article