



Alleviation of Aflatoxin-B₁ Toxicity by using Clay Adsorbent in Nile Tilapia (*Oreochromis niloticus*) Diets

Dilawar Hussain* and Abdul Mateen

Department of Zoology, Wildlife and Fisheries, University of Agriculture, Faisalabad 38040, Pakistan

ABSTRACT

The present study was designed to alleviate the aflatoxin-B₁ toxicity by the use of 0.5% calcium bentonite clay and to evaluate its effect on the growth performance of Nile tilapia over a period of 10 weeks. Inclusion of AFB₁ at both 2 and 4 ppm levels significantly ($p < 0.05$) decreased specific growth rate (SGR), net weight gain (NWG), average daily gain (ADG), survival, feed intake, feed efficiency ratio (FER) and protein efficiency ratio (PRE), irrespective of the addition of the 4TX in the diets. Among different dietary groups of the fish, % survival was not affected significantly ($p < 0.05$). T₁ showed maximum NWG (45.49±3.85), FER (0.739±0.02) and PER (36.36±1.83) when compared to other dietary treatment groups. The addition of 4TX clay in the diets at both 2 and 4 ppm AFB₁ concentrations have almost the same effect on the growth parameters tested except in the case of PER. T₃ (0.5% 4TX+2 ppm AFB₁) showed better PER (31.42±1.74) when compared to T₄ (0.5% 4TX+ 4 ppm AFB₁) group (27.76±0.67). Interaction between different growth parameters of tilapia fed AFB₁ and 4TX supplemented diets showed that net weight gain (NWG) was significantly ($p < 0.01$) correlated with average daily gain (ADG), specific growth rate (SGR), feed intake (FI) and protein efficiency ratio (PER). In conclusion, aflatoxin-B₁ negatively impacted the growth performance of Nile tilapia regardless the addition of the 4TX clay. Addition of 4TX in the diets has significant effect on some of the parameters tested.

Article Information

Received 09 September 2016

Revised 23 September 2016

Accepted 10 October 2016

Available online 28 January 2017

Authors' Contributions

DH and AM conceived and designed the study. DH collected and analyzed data and wrote the article.

Key words

Nile tilapia, Calcium bentonite clay, Aflatoxin-B₁, Growth performance, Correlation matrices.

INTRODUCTION

Aflatoxins are toxic compounds that are produced mainly by two naturally present mold species (*Aspergillus flavus* and *Aspergillus parasiticus*). These fungal species are abundant in the soil and are a common contaminant of feed crops in warm and humid environments (Fowler *et al.*, 2014). Oilseed crops such as corn, cottonseed and peanut meal are mostly contaminated with aflatoxins. In warm and humid conditions, fish meal, soybean meal, sunflower and other nutritionally complete feeds are also at a risk of aflatoxin contaminations (Hutanasu *et al.*, 2009; Kitya *et al.*, 2010; Kumagai *et al.*, 2008; Tchana *et al.*, 2010). Among all the documented aflatoxins, aflatoxin-B₁ (AFB₁) is the most widespread and toxic for all the animal species including humans due to its strong mutagenic, teratogenic and carcinogenic effects on these species (Han *et al.*, 2009; Santacrose *et al.*, 2008). Being a wide distribution in tropical and sub-tropical areas of the world where a large number of studies reported the frequent prevalence of aflatoxin contamination, tilapia is repeatedly studied to examine the deleterious effects of AFB₁ on its

physiological properties and health (Deng *et al.*, 2010). Chen and Rawlings (2008) collected commercial feeds and raw materials from Asia and reported the fact that aflatoxins could be detected in 96.1% of the 334 tests.

Negative effect of AFB₁ on the health performance of Nile tilapia were observed in many studies as Zychowski *et al.* (2013b) reported the deleterious effects of AFB₁ in tilapia at lower concentrations such as 1.5 ppm. Mehrim and Salem (2013) have revealed serious lethal effects of incorporating 150 ppb AFB₁ in the diets of Nile tilapia with addition to the hepatotoxic effects on the liver than the control group. In another study by El-Banna *et al.* (1992) it is reported that the growth of Nile tilapia decreases significantly when fed 100 µg AFB₁ / kg for a period of 10 weeks, and was observed 16.70% mortality when exposed to a dose of 200 µg / kg.

Extensive research has been designed and carried out to prevent mycotoxicosis in different animal species, including fish that mainly consists of different physical, nutritional chemical or biological means. Vast use of mycotoxin adsorbents that can capture and adsorb the toxin molecules by ion exchange process, thereby delaying their entry into the blood from the gastrointestinal tract has been made considerable attention in the prevention of mycotoxins. Hydrated sodium calcium aluminosilicate (HSCAS), bentonite, zeolite ore-based compound, canola

* Corresponding author: sirdilawar@yahoo.com

0030-9923/2017/0002-0425 \$ 9.00/0

Copyright 2017 Zoological Society of Pakistan

oil, activated carbon, spent bleaching clay, inorganic adsorbents, some organic acids and alumina-silicates have been disclosed in the prevention of aflatoxicosis (Devegowda and Murthy, 2005).

Deleterious effects of aflatoxins in many animal species has alleviated by the use of clay based, non-nutritive adsorbents. Several studies reported the efficacy of these bentonites and aluminosilicate clays to adsorb and mitigate the harmful effects of aflatoxins by *in vitro* binding of these toxins in the interlayers and on the edge of the clay structure with relatively high attraction (Phillips, 1999; Desheng *et al.*, 2005; Kannewischer *et al.*, 2006). As a result of this absorbance, the aflatoxins molecules passes through the gastrointestinal tract unabsorbed, in that way decreasing the lethal effects.

The 4TX calcium bentonite clay was selected on the basis of presence of 81.4% smectite which shows its purity level and strong adsorption capacity with aflatoxin-B₁ (13.9% w/w) (Velazquez, 2011). Considering the fact that commercial bentonites used in feeding industry are dried and ground to fine powders, we obtained commercial bentonite (4TX) from Southern Clay Products Inc. (Gonzales, Texas, USA). The current study is therefore designed to alleviate the AFB₁ induced toxicity by the use of 0.5% 4TX clay against 2 and 4 ppm AFB₁ in the diets and to evaluate the potential effects of the clay on growth parameters of Nile tilapia.

MATERIAL AND METHODS

Experimental diets

Controlled basal diet was formulated having 33.8 g of protein, 8.2 g of lipid, and an estimated 290 KCal of digestible energy 100g⁻¹. This diet fulfils all published nutrient requirements of Nile tilapia (Table I) (Lim and Webster, 2006). Purified aflatoxins B₁ was purchased from Sigma-Aldrich, USA and prior to the addition into the remaining ingredients of the diets it is dispersed in chloroform and thoroughly mixed with Celufil. All dry ingredients accurately weighed and then mixed for 40 min in a mixer (V-mixer) then oil was added slowly to mix thoroughly and at the end 400 mL of water was added and mixed in mixer (Hobart mixer) for 50-60 min. The mixer of all the ingredients then passed through a 3-mm die that is attached with a meat grinder. The diets were dried, labelled and then stored at -20 °C for use in experiment. The four experimental diets consisted of: T₁, 0% 4TX + 0 ppm AFB₁ (Negative control); T₂, 0.5% 4TX + 0 ppm AFB₁ (Positive control); T₃, 0.5% 4TX + 2 ppm AFB₁; T₄, 0.5% 4TX + 4 ppm AFB₁.

Table I.- Basal diet Composition (% dry matter basis) fed tilapia for 10 weeks.

Ingredients	(%) dry matter basis
Fish meal	11
Soybean meal	46.68
Starch	20.25
Vitamin premix ^a	3
Mineral premix	4
Carboxymethyl cellulose	2
Soy oil	4.64
CaPO ₄ , dibasic	1
Glycine	1
DL-methionine	0.15
4TX	0
AFB ₁ (µg)	0
Celufil	6.28
Total	100

^a Contains (as g kg⁻¹): Ca(C₆H₁₀O₆) 5H₂O, 348.49; Ca(H₂PO₄)₂ H₂O, 136.0; FeSO₄ 7H₂O, 5.0; MgSO₄ 7H₂O, 132.0; K₂HPO₄, 240.0; NaH₂PO₄ H₂O, 88.0; NaCl, 45.0; AlCl₃ 6H₂O, 0.15; KI, 0.15; CuSO₄ 5H₂O, 0.5; MnSO₄ H₂O, 0.7; CoCl₂ 6H₂O, 1.0; ZnSO₄ 7H₂O, 3.0; Na₂SeO₃, 0.011.

Stocking and culturing of tilapia

Nile tilapia fingerlings were imported from a local Hatchery operated in Louisiana State, USA. Before the start of the experiment, tilapia were stocked and conditioned with a commercial diet for 15 days in round tanks and conditioned on basal diet for 7 more days in glass aquaria. A closed, recirculating system consisting on glass aquaria 110-L were used for the feeding trial, where salinity was maintained at 5 ppt by the addition of saline water and temperature was controlled at 26 °C. Water quality parameters (Temperature, pH, Ammonia, nitrite, dissolved oxygen and salinity) monitored weekly and keeping below the toxic levels using a biofiltration unit. A total of 144 fish of equal size (4.5±0.4 g) were randomly divided into 4 experimental units (12fish/unit) with three replicates for a period of 10 weeks. Fish were fed at 5% of body weight daily their assigned diet at 8:00 AM and 4:00 PM. The system was monitored daily to check any abnormalities and mortalities and if any, was removed immediately and recorded.

Fish growth response

After terminating the trial, net weight gain (NWG), average daily gain (ADG), % survival, % specific growth rate (SGR), protein efficiency ratio (PER), feed efficiency

ratio (FER), feed intake (FI), and correlation matrices were calculated.

Statistical analysis

All the parameters were computed by analysis of variance (ANOVA) first and then differences among different treatment groups and means were compared by Least Significant Difference (LSD) test. To compute all the statistical data, Statistix, software program version 8.1 (Analytical Software, Tallahassee, FL) was used. The significance level was set at $P \leq 0.05$.

RESULTS

Growth performance

After 10 week exposure of AFB₁ and 4TX clay, there observed a non-significant difference ($P > 0.05$) in % survival among all the treatments. While the growth parameters of tilapia regarding the initial body weight (IBW), final body weight (FBW), average daily gain (ADG), net weight gain (NWG) and specific growth rate (SGR) was significantly ($p < 0.05$) affected after the exposure of AFB₁ and 4TX supplemented diets (Table II). Addition of AFB₁ in the diets at both 2 and 4 ppm negatively affected the health performance of tilapia, irrespective the supplementation of 4TX in the diets. When supplemented alone, 4TX has significantly higher net weight gain (45.03 ± 5.10) and specific growth rate (2.202 ± 0.07) as compared to 2 ppm (31.63 ± 3.03) and (1.998 ± 0.06) and 4 ppm AFB₁ (31.23 ± 5.73) and (1.986 ± 0.11) exposed fish, respectively. The efficacy of 4TX clay in alleviating the AFB₁ toxicity at both 2 and 4 ppm AFB₁ concentrations remains non-significant ($P > 0.05$).

Feed and protein utilization

Feed efficiency ratio (FER) and protein efficiency

ratio (PER) exhibited a significant difference ($p < 0.05$) in all the treatment groups (Table III). T₁ showed maximum (0.739 ± 0.02) while T₄ showed minimum (0.584 ± 0.03) FER. 4TX clay showed better PER (31.42 ± 1.74) at 2 ppm AFB₁ concentration as compared to 4 ppm AFB₁ exposed group (27.76 ± 0.67).

Correlation matrices

Interaction between different growth parameters of tilapia fed AFB₁ and 4TX supplemented diets are shown in Table IV. Net weight gain (NWG) was significantly ($p < 0.01$) correlated with specific growth rate (SGR), protein efficiency ratio (PER), average daily gain (ADG) and feed intake (FI).

Water quality parameters

All the water quality parameters were found within the range (temperature, $25.5-27.2^\circ\text{C}$; pH, $7.89-8.61$; ammonia, $0.11-0.20$ mg/1N NH₃; nitrite, $0.03-0.09$ mg/1N NO₂⁻L; dissolved oxygen, $5.42-6.51$ mg/L and salinity, $0.41-0.48$ ppt) optimum for tilapia growth (Table V).

Table III.- Feed intake, feed efficiency ratio (FER) and protein efficiency ratio (PER) of tilapia AFB₁ and 4TX supplemented diets for 10 weeks.

Treatment	FI (g/fish)	FER	PER
T ₁	61.58±5.56AB	0.739±0.02A	36.36±1.83A
T ₂	65.70±10.15A	0.689±0.09A	34.53±0.51A
T ₃	52.14±3.51B	0.624±0.03B	31.42±1.74B
T ₄	53.00±4.82B	0.584±0.03B	27.76±0.67C

Means sharing similar letters in a column are statistically non-significant ($p > 0.05$). T₁, 0%4TX+0 ppm AFB₁ (Negative control); T₂, 0.5% 4TX+ 0 ppm AFB₁ (Positive control); T₃, 0.5% 4TX+2 ppm AFB₁; T₄, 0.5% 4TX+4 ppm AFB₁; FI, Feed intake; FER, Feed efficiency ratio; PER, Protein efficiency ratio. Feed efficiency ratio (FER) = Live weight gain (g) / Feed intake (g). Protein efficiency ratio (PER) = Live weight gain (g) / Protein intake (g).

Table II.- Means ± SD of growth performance of tilapia fed AFB₁ and 4TX supplemented diets for 10 weeks.

Treatment	Body weight (g/fish)		Body gain		SGR (%)	Survival (%)
	IBW	FBW	NWG (g/fish)	ADG (g/fish/d)		
T ₁	4.657±0.20B	50.14±4.02A	45.49±3.85A	0.607±0.05A	2.209±0.05A	94.44±4.81A
T ₂	5.050±0.15A	50.07±5.24A	45.03±5.10 A	0.600±0.07A	2.202±0.07A	94.44±4.81A
T ₃	4.467±0.11B	36.10±3.13B	31.63±3.03B	0.422±0.04B	1.998±0.06B	97.22±4.81A
T ₄	4.710±0.12B	35.94±5.83B	31.23±5.73B	0.416±0.08B	1.986±0.11B	97.22±4.81A

Means sharing similar letters in a column are statistically non-significant ($p > 0.05$). T₁, 0%4TX+0 ppm AFB₁ (Negative control); T₂, 0.5% 4TX+ 0 ppm AFB₁ (Positive control); T₃, 0.5% 4TX+2 ppm AFB₁; T₄, 0.5% 4TX+4 ppm AFB₁; IBW, Initial body weight; FBW, Final body weight; NWG, Net weight gain; ADG, Average daily gain; SGR, Specific growth rate.

Net weight gain (g/fish) NWG = Average final weight (g) – Average initial weight (g).

Average daily gain, (g/fish/day) ADG = AWG (g)/Experimental period (days).

Specific growth rate (SGR, %/day) = [In final weight – In initial weight] x 100/Experimental period (d).

Survival rate (SR %) = End number of the alive fish/The beginning number of the fish x 100.

Table IV.- Interaction between growth responses of tilapia fed different levels of AFB₁ and 4TX supplemented diets for 10 weeks.

Control Variables	IBW	FBW	NWG	ADG	SGR	Survival	FI	FER
FBW	0.7367** 0.0063							
NWG	0.7222** 0.0080	0.9998** 0.0000						
ADG	0.7222** 0.0080	0.9998** 0.0000	1.0000** 0.0000					
SGR	0.7039* 0.0106	0.9931** 0.0000	0.9937** 0.0000	0.9937** 0.0000				
Survival	-0.1242 0.7005	-0.1255 0.6976	-0.1243 0.7004	-0.1243 0.7004	-0.0555 0.8641			
FI	0.7933** 0.0021	0.9359** 0.0000	0.9324** 0.0000	0.9324** 0.0000	0.9107** 0.0000	-0.2962 0.3500		
FER	-0.1513 0.6389	0.3732 0.2322	0.3865 0.2146	0.3865 0.2146	0.3954 0.2034	0.0224 0.9449	0.1730 0.5907	
PER	0.2993 0.3446	0.7287** 0.0072	0.7359** 0.0064	0.7526** 0.0047	0.7171** 0.0087	-0.4257 0.1676	0.6202 0.0315	0.5487 0.0647

Upper values indicated Pearson's correlation coefficient; lower values indicated level of significance. *, Significant (P<0.05); **, Highly significant (P<0.01).

Table V.- Physico-chemical analysis of control and AFB₁ and 4TX treated aquaria.

Week	Temp. (°C)	pH	Ammonia	Nitrite	Dissolved Oxygen	Salinity
Initial	26.7	8.52	0.14	0.03	6.45	0.42
1.	26.7	7.89	0.11	0.03	6.51	0.43
2.	26.6	8.45	0.15	0.03	5.99	0.41
3.	27.2	8.46	0.12	0.04	6.32	0.48
4.	26.7	7.96	0.12	0.05	6.30	0.46
5.	26.8	8.39	0.13	0.04	6.31	0.47
6.	26.9	8.48	0.18	0.03	6.36	0.42
7.	25.5	8.53	0.14	0.03	6.39	0.44
8.	26.7	8.61	0.13	0.04	5.87	0.46
9.	26.8	8.54	0.20	0.09	5.42	0.42
10.	26.7	8.46	0.12	0.03	6.35	0.41

Ammonia, (Mg/1N NH₃); Nitrite, (Mg/1N NO₂⁻L); Dissolved Oxygen, (Mg/L); Salinity, (Ppt).

DISCUSSION

In tropical and subtropical regions of the world, a high risk of aflatoxin contamination has been observed in many studies due to the higher use of plant based

alternatives in animal diets that ultimately have lethal effects on fish health. Overall, aflatoxin-B₁ negatively affected the tilapia over a course of 10 weeks. The results of current study showed a reduction in specific growth rate (SGR), average daily gain (ADG) and net weight gain (NWG), and at both the supplemented levels of AFB₁. Fish exposed to 4 ppm AFB₁ affected the most comparing with 2 ppm AFB₁ offered fish and control groups. This negative effect of AFB₁ on growth parameters at 2 and 4 ppm proved anti-nutritional nature of AFB₁ as described in previous findings (Al-Faragi, 2014; Ayyat *et al.*, 2013; Chavez-Sanches *et al.*, 1994; Deng *et al.*, 2010; El-Banna *et al.*, 1992; Encarnacao *et al.*, 2009; Salem *et al.*, 2010; Sepahdari *et al.*, 2010; Shehata *et al.*, 2009; Zaki *et al.*, 2008; Zychowski *et al.*, 2013a, b).

In the present study, % survival remained almost the same (p<0.05) among tilapia exposed to 2 and 4 ppm AFB₁ concentrations along with controls. Our results was confirmed by the study of Tuan *et al.* (2002) who concluded that when tilapia exposed to 10 ppm AFB₁/kg or less have not increased the mortality over a period of 8 weeks. In another study by Chavez-Sanchez *et al.* (1994), it is reported that even when exposed to 30 ppm AFB₁/kg did not cause death in tilapia. Our results regarding the survival confirmed this trend.

In the present study, protein efficiency ratio (PER) and feed efficiency ratio (FER) was also decreased significantly

($p < 0.05$) with the increasing AFB₁ concentration. Our results regarding the feed and protein utilization are in line with the previous findings by Abdelhamid *et al.* (2004), Hussein *et al.* (2000), Nguyen *et al.* (2002) and Salem (2002). The possible explanation of this toxicity and deleterious effect of AFB₁ may be because of pathological modifications in the gastro-intestinal tract of the fish (Murjani, 2003). Also, our results was in line with the results of Nguyen *et al.* (2002) who proposed that when fish offered a diet ranging from 10 and 100 ppm AFB₁/kg was expelled out when ingested. When fish administrated a diet having 100 ppm AFB₁/kg diet, only consumed 59 ppm AFB₁/kg of its body weight, the three times than the amount for fish fed the 10 mg AFB₁/kg. Additionally, the results of the study of Salem (2002) reported that a significant reduction ($P < 0.05$) was observed in protein and feed efficiency feed in tilapia when fed with dietary AFB₁. Similar findings regarding the protein and feed utilization were reported by Abdelhamid *et al.* (2002b).

Addition of 4TX clay in the diets did not show any significant difference regarding the growth parameters at both 2 and 4 ppm AFB₁ exposed groups except in protein efficiency ratio in which 0.5% 4TX+2ppm AFB₁ group (T₃) showed higher (31.42 ± 1.74) PER as compared to 0.5%4TX+4ppm AFB₁ (27.76 ± 0.67) group (T₄). The theory behind this effectiveness is that mycotoxin adsorbents such as 4TX strongly binds with the AFB₁ molecules present in the feed that it prevents the absorption of AFB₁ in the digestive tract of the animals. The efficacy of 4TX to adsorb and remove the intoxication of AFB₁ in animal species was also reported by Fowler *et al.* (2014), (2015) and Velazquez (2011). Bentonite clays were found active in minimizing the bioavailability of aflatoxins (Chaturvedi *et al.*, 2002; Desheng *et al.*, 2005; Magnoli *et al.*, 2008). All the water quality parameters were found suitable for the optimum growth of Nile tilapia as described in previous studies (Abdelhamid *et al.*, 2002b).

CONCLUSIONS

The results showed significant difference ($p < 0.05$) regarding the growth parameters among all the dietary treatments. Fish exposed to 4 ppm AFB₁ performed poorly in terms of growth performance when compared to 2 ppm exposed fish and the control groups. Both negative (T₁) and positive (T₂) control groups showed maximum growth performance over AFB₁ offered fish. Supplementation of 4TX in the diets has positive effects on fish growth with better PER in 2 ppm AFB₁ group compared to 4 ppm AFB₁ exposed fish. So, a clay binder such as 4TX if used efficiently have the potential to reduce AFB₁ exposure, thereby preventing bioavailability and consequent

effects, such as decreases in growth parameters and immunosuppression and in Nile tilapia.

ACKNOWLEDGMENT

The authors would like to thank Higher Education Commission of Pakistan for partial support through International Research Support Initiative Program (IRSIP).

Conflict of interest statement

We declare that we have no conflict of interest.

REFERENCES

- Abdelhamid, A.M., Abdelkhalek, A.E., Mehrim, A.I. and Khalil, F.F., 2004. An attempt to alleviate aflatoxicosis on Nile tilapia fish by dietary supplementations with chicken-hatchery by-products (egg shells) and shrimp processing wastes (shrimp shells) 2-On clinical, blood and histological parameters. *J. agric. Sci.*, **29**: 6175-6196.
- Abdelhamid, A.M., Khalil, F.F.M., El-Barbary, M.I., Zaki, V.H. and Hussein, H.S., 2002b. *Feeding Nile tilapia on Biogen® to detoxify aflatoxic diets*. Proc. 1st Confr. Animal & Fish Prod., Sept, 24-25, Mansoura, Egypt, pp. 207-230.
- Al-Faragi, 2014. The efficacy of prebiotic (β -Glucan) as a feed additive against toxicity of hepatocarcinogenicity for four aflatoxins by two exposure routes in rainbow trout. *Mutat. Res.*, **399**: 223-244.
- Ayyat, M.S., Abd-Rhman, G.A., El-Marakby, H.I., Mahmoud, H.K. and Hessian, A.A.A., 2013. Reduction in the aflatoxin toxicity in Nile tilapia fish. *Egypt. J. Nutr. Feeds*, **16**: 469-479.
- Chaturvedi, V.B., Singh, K.S. and Agnihotri, A.K., 2002. *In vitro* aflatoxin adsorption capacity of some indigenous aflatoxin adsorbents. *Indian J. Anim. Sci.*, **72**: 257-260.
- Chavez-Sanchez, M.C., Palacios, C.A.M. and Moreno, I.O., 1994. Pathological effects of feeding young *Oreochromis niloticus* diets supplemented with different levels of aflatoxin B₁. *Aquaculture*, **127**: 49-60. [https://doi.org/10.1016/0044-8486\(94\)90191-0](https://doi.org/10.1016/0044-8486(94)90191-0)
- Chen, H.Y. and Rawlings, R., 2008. The truth of mycotoxin contamination of feed in Asia region. *China Poult.*, **30**: 33-35.
- Deng, S.X., Tian, L.X., Liu, F.J., Jin, S.J., Liang, G.Y., Yang, H.J., Du, Z.Y. and Liu, Y.J., 2010. Toxic effects and residue of aflatoxin B₁ in tilapia (*Oreochromis niloticus*) (*O. aureus*) during long-term dietary exposure. *Aquaculture*, **307**: 233-240.

- <https://doi.org/10.1016/j.aquaculture.2010.07.029>
 Desheng, Q., Fan, L., Yanhu, Y. and Niya, Z., 2005. Adsorption of aflatoxin B₁ on montmorillonite. *Poult. Sci.*, **84**: 959-961. <https://doi.org/10.1093/ps/84.6.959>
- Devegowda, G. and Murthy, T.N.K., 2005. Mycotoxins: their effect in poultry and some practical solutions. In: *The mycotoxin blue book* (ed. D.E. Diaz), Nottingham University Press, Nottingham, England, pp. 25-50.
- El-Banna, R., Teleb, H.M., Hadi, M.M. and Fakhry, F.M., 1992. Performance and tissue residue of tilapias fed dietary aflatoxin. *J. Vet. Med.*, **40**: 17-23.
- Encarnacao, P., Srikhum, B., Rodrigues, I. and Hofstetter, U., 2009. Growth performance of red tilapia (*O. niloticus* x *O. mossambicus*) fed diets contaminated with aflatoxin B₁ and the use of a commercial product to suppress negative effects. *World Aquaculture*, 2009. Veracruz, Mexico, (Abstract).
- Fowler, J., Hashim, M., Velazquez, A.L.B., Deng, Y. and Bailey, C.A., 2014. Utilization of a spray-applied calcium bentonite clay to ameliorate the effects of low-levels of aflatoxin in starter broiler diets containing DDGS. *Nat. Prod. Chem. Res.*, **2**: 127-130.
- Fowler, J., Li, W. and Bailey, C., 2015. Effects of calcium bentonite clay in diets containing aflatoxin when measuring liver residues of aflatoxin B₁ in starter broiler chicks. *Toxins*, **7**: 3455-3464. <https://doi.org/10.3390/toxins7093455>
- Han, D., Xie, S., Zhu, X., Yang, Y. and Guo, Z., 2009. Growth and hepatopancreas in gibel carp fed diets containing low levels of aflatoxin B₁. *Aquacult. Nutr.*, **16**: 335-342. <https://doi.org/10.1111/j.1365-2095.2009.00669.x>
- Hussein, S.Y.I., Mekkawy, A.A., Moktar, Z.Z. and Mubarak, M., 2000. Protective effect of *Nigella sativa* seed against aflatoxicosis in *Oreochromis niloticus*. Proc. Conf. Mycotoxins and Dioxins and the Environment, Bydgoszcz, 25-27 Sept., pp. 109-130.
- Hutanasu, C., Sfarti, C., Trifan, A., Hutanasu, M. and Stanciu, C., 2009. Aflatoxin contamination of food: additional risk factor for chronic liver diseases. *Rev. Med. Chir. Soc. Med. Nat. Iasi.*, **113**: 1061-1065.
- Kannewischer, I., Arvide, M.G.T., White, G.N. and Dixon, J.B., 2006. Smectite clays as adsorbents of aflatoxin B₁ initial steps. *Clay Sci.*, **12**: 199-204.
- Kitya, D., Bbosa, G.S. and Mulogo, E., 2010. Aflatoxin levels in common foods of South Western Uganda: a risk factor to hepatocellular carcinoma. *Eur. J. Cancer Care (Engl.)*, **19**: 516-521. <https://doi.org/10.1111/j.1365-2354.2009.01087.x>
- Kumagai, S., Nakajima, M., Tabata, S., Ishikuro, E., Tanaka, T., Norizuki, H., Itoh, Y., Aoyama, K., Fujita, K., Kai, S., Sato, T., Saito, S., Yoshiike, N. and Sugita-Konishi, Y., 2008. Aflatoxin and ochratoxin A contamination of retail foods and intake of these mycotoxins in Japan. *Food Addit. Contam. Part A Chem. Anal. Contr. Expo. Risk Assess.*, **25**: 1101-1106. <https://doi.org/10.1080/02652030802226187>
- Lim, C. and Webster, C.D., 2006. *Tilapia biology, culture and nutrition*. The Haworth Press, Inc., Binghamton, New York, USA, pp. 469-501.
- Magnoli, A.P., Cabaglieri, L.R., Magnoli, C.E., Monge, J.C., Miazzi, R.D., Peralta, M.F., Salvano, M.A., Rosa, C.A.R., Dalcerro, A.M. and Chiacchiera, S.M., 2008. Bentonite performance on broiler chickens fed with diets containing natural levels of aflatoxin B₁. *Rev. Brasil. Med. Vet.*, **30**: 55-60.
- Mehrim, A.I. and Salem, M.F., 2013. Medicinal herbs against aflatoxicosis in Nile tilapia (*Oreochromis niloticus*): Clinical signs, postmortem lesions and liver histopathological changes. *Egypt. J. Aquacult.*, **3**: 13-25
- Murjani, G., 2003. Chronic aflatoxicosis in fish and its relevance to human health. *J. Am. Vet. med. Assoc.*, **176**: 719-724.
- Nguyen, A.T., Grizzle, J.M., Lovell, R.T., Manning, B.B. and Rottinghaus, E.G., 2002. Growth and hepatic lesions of Nile tilapia *Oreochromis niloticus* fed diets containing aflatoxin B₁. *Aquaculture*, **212**: 311-319. [https://doi.org/10.1016/S0044-8486\(02\)00021-2](https://doi.org/10.1016/S0044-8486(02)00021-2)
- Phillips, T.D., 1999. Dietary clay in the chemoprevention of aflatoxin induced disease. *Toxicol. Sci.*, **52**: 118-126. <https://doi.org/10.1093/toxsci/52.2.118>
- Salem, M.F.E., 2002. *Effect of dietary graded levels of aflatoxin B₁ on growth performance and chromosomal behaviour of Nile tilapia Oreochromis niloticus*. Ph.D. Thesis, Kafu El-Sheikh, Tanta University, Egypt.
- Salem, M.F.I., Shehab, E.M.T., Khalafallah, M.M.M.A., Sayed, S.H. and Amal, S.H., 2010. Nutritional attempts to detoxify aflatoxic effects in diets of tilapia fish (*Oreochromis niloticus*). *J. World Aquacult. Soc.*, **5**: 195-206.
- Santacroce, M.P., Conversano, M.C., Casalino, E., Lai, O., Zizzadoro, C., Centoducati, G. and Crescenzo, G., 2008. Aflatoxins in aquatic species: metabolism, toxicity and perspectives. *Rev. Fish Biol. Fish.*, **18**: 99-130. <https://doi.org/10.1007/s11160-007-9064-8>
- Sepahdari, A., Ebrahimzadeh, M., Sharifpour, H.A.I.,

- Khosravi, A., Motallebi, A.A., Mohseni, M., Kakoolaki, S., Pourali, H.R. and Hallajian, A., 2010. Effects of different dietary levels of Aflatoxin B₁ on survival rate and growth factors of Beluga (*Huso huso*). *Iran. J. Fish. Sci.*, **9**: 141-150.
- Shehata, S.A., El-Melegy, Kh. M. and Ebrahim, M.S., 2009. Toxicity reduction of aflatoxin B₁ by vitamin C in fish. *J. World Aquacult. Soc.*, **4**: 73-86.
- Tchana, A.N., Moundipa, P.F. and Tchouanguép, F.M., 2010. Aflatoxin contamination in food and body fluids in relation to malnutrition and cancer status in Cameroon. *Int. J. Environ. Res. Publ. Hlth.*, **7**: 178-188. <https://doi.org/10.3390/ijerph7010178>
- Tuan, N.A., Grizzle, J.M., Lovell, R.T., Manning, B.B. and Rottinghaus, G.E., 2002. Growth and hepatic lesions of Nile tilapia (*Oreochromis niloticus*) fed diets containing aflatoxin B₁. *Aquaculture*, **212**: 311-319. [https://doi.org/10.1016/S0044-8486\(02\)00021-2](https://doi.org/10.1016/S0044-8486(02)00021-2)
- Velazquez, A.L.B., 2011. *Texas bentonites as amendments of aflatoxin-contaminated poultry feed*. Master's thesis, Texas A&M University, USA.
- Zaki, M.S., Sharaf, N.E. Rashad, H. Mastala, S.O. and Fawzy, Q.M., 2008. Diminution of aflatoxicosis in tilapia nilotica fish by dietary supplementation with fix in toxin and Nigella sativa oil. *Am. Eurasian J. Agric. environ. Sci.*, **3**: 211-215.
- Zychowski, K.E., Hoffman, A.R., Ly, H.J., Pohlenz, C., Buentello, A., Romoser, A., Gatlin D.M., Philips, T.D., 2013a. The effect of aflatoxin-B₁ on red drum (*Sciaenops ocellatus*) and assessment of dietary supplementation of NovaSil for the prevention of aflatoxicosis. *Toxins*, **5**: 1555-1573. <https://doi.org/10.3390/toxins5091555>
- Zychowski, K.E., Pohlenz, C., Mays, T., Romoser, A., Hume, M., Buentello, A., Gatlin, D.M. and Phillips, T.D., 2013b. The effect of NovaSil dietary supplementation on the growth and health performance of Nile tilapia (*Oreochromis niloticus*) fed aflatoxin-B₁ contaminated feed. *Aquaculture*, **76**: 117-123. <https://doi.org/10.1016/j.aquaculture.2012.11.020>