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In Silico Analysis of Cu-Zn Superoxide Dismutase and Mn Superoxide Dismutase Genes in Fugu (*Takifugu rubripes*)

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ABSTRACT

Superoxide dismutases are the best-known enzymatic antioxidants because of their central role in the antioxidant defense system. In this study, the phylogeny, gene structure, and conserved gene synteny of *sod1* and *sod2* genes in fugu—a model organism—were determined. Maximum amino acid similarity identity was found between putative fugu Sod1 and Sod2 proteins and their orthologs from teleost fish and tetrapods. Phylogenetic clustering was seen between *sod* genes in fugu and their orthologs. Finally, highly conserved gene synteny was determined between fugu *sod* genes and their orthologs from teleost fish and human.

INTRODUCTION

Fugu (Japanese pufferfish, *Takifugu rubripes*) is an ideal model organism for vertebrate genome research and developmental biology studies (Uji et al., 2011). Japanese pufferfish has more advantages for genomic studies compared with other vertebrates. Its 400-Mb genome size is quite small compared with the 3,000-Mb human genome size (Watabe and Ikeda, 2006). It is also known that the pufferfish genome has more genes than those of coelacanths and air-breathing fish due to teleost-specific whole genome duplication (tsWGD) event (Van de Peer, 2004). The small size of the genome, which is a factor that facilitates the detection and analysis of genes, has regular sequences like those of the other vertebrates, requiring less work to obtain comparable data. Green spotted pufferfish (Tetraodon nigroviridis) can also be used as model organisms for vertebrate genomes (Close et al., 2016). Many human genes have been uncovered by comparing these two pufferfish genomes with the human genomes (Brenner et al., 1993).

Aerobic biological systems generate reactive oxygen species (ROS), including hydrogen peroxide (H_2O_2) and superoxide anion (O_2^{--}) . ROS attacking essential cellular components may result in irreversible damage in their structure. However, organisms can initiate ROS removal and restrict generation of ROS via their well-developed antioxidant defense system (ADS).



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The superoxide dismutases (SODs) have an important role in ADS due to their ability to prevent the generation of hydroxyl radical (OH) and removing O₂⁻ and catalyzing the dismutation of O_2^{-1} into H_2O_2 and molecular oxygen. SODs using metal ions in their activities have two forms of eukaryotic systems, namely, Cu⁺²/Zn⁺² and Mn⁺² SOD. While the first isoform (Cu⁺²/Zn⁺² SOD) is found in cytosols and its activity is not affected by oxidative stress, the second isoform (Mn⁺² SOD) is located in mitochondria, and its activity is increased in proportion to oxidative stress (Babior, 1997; Davies, 2000). Several studies have been carried out on SOD enzymes in aerobic organisms (Lopes et al., 2001; Fink et al., 2002; van der Oost et al., 2003; Ken et al., 2003; Farombi et al., 2007; Cho et al., 2009). Limited work, however, is available on the genomic organization and gene structure of teleost sods (Cho et al., 2009) and regulatory region of the sod genes in teleost fish (Maehara et al., 1999; Mao et al., 2006). It is known that transcription of the Mn⁺² SOD gene can be regulated by environmental factors (Valavanidis et al., 2006; Kim et al., 2007; Cho et al., 2009), and the function and structure of SOD2 are well conserved in variant organisms (Fink et al., 2002). Therefore, the goals of the current study are determining sod genes in the fugu genome using bioinformatic tools, leading to future molecular works on antioxidant enzyme genes in teleost fishes.

MATERIALS AND METHODS

Fugu *sod1* and *sod2* gene sequences were obtained by performing BLAST (http://useast.ensembl.org/ Multi/blastview) searches with an identical orthologous

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zebrafish (*Danio rerio*) Sod protein sequence as an inquiry to the Ensembl genome sequence database (http://useast. ensembl.org/index.html). Zebrafish *sod* cDNA sequences provided from the Ensembl genome database were used to identify ESTs coded by specific fugu *sod1* and *sod2* genes in a BLAST search of the NCBI database (http://blast. ncbi.nlm.nih.gov).

Teleost fish exhibits strict evolutionary conservation for the gene structure in the same gene family. Therefore, the exon-intron junctions of the *sod1* and *sod2* genes were determined using zebrafish *sod1* and *sod2* gene structures as a reference. To confirm whether *sod1* and *sod2* genes were transcriptionally active or served as pseudogenes or non-functional genes, tBLASTn searches in the NCBI database were conducted to identify their ESTs using their Ensembl-derived amino acid sequences as queries (Table I). Superoxide dismutase genes in fugu with their corresponding zebrafish query sequence IDs, NCBI cDNA IDs, chromosomal locations and length of Sod polypeptides were also determined using Ensembl and the NCBI genome databases (Table II).

Table I. Ensembl gene ID and expressed sequence tags (EST) coded by fugu and zebrafish *sod1* and *sod2* genes.

Gene	Ensembl gene ID	EST
Fugu sod1	ENSTRUG00000021322	Not detected
Fugu sod2	ENSTRUG0000005242	Not detected
Zebrafish sod1	ENSDARG00000043848	EH544614.1
Zebrafish sod2	ENSDART00000062556.4	EH537787.1

Phylogenetic analysis

CLUSTALW (Thompson et al., 1994) at BioEdit software (http://www.mbio.ncsu.edu/bioedit/page2.html) was used for sequence alignment of the sod1 and sod2 genes. The protein sequence of fugu Sod1 and Sod2 was aligned with Sod/SOD protein sequences from fugu, tetraodon, medaka, zebrafish, human, and mice. The pairwise alignment of the BLOSUM62 matrix (Gromiha, 2010) was used for sequence identity and similarity. A maximum-likelihood tree with the Poisson correction distance model based on amino acid substitution per site was built using MEGA6 (Tamura et al., 2013) to determine the phylogenetic relationships of the fugu, tetraodon, medaka, zebrafish, human, and mouse Sod sequences. A bootstrapped neighbor-joining tree was also constructed before construction of the maximum-likelihood tree to confirm the phylogeny of sod genes (data not shown). As in a previous study, the protein sequence of human

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lymphocyte cytosolic protein LCP2) was used as an external group (Kell *et al.*, 2018).

Conserved gene synteny

The conserved gene synteny of fugu sod genes with the sod/SOD of zebrafish, medaka and human was arranged manually using the region conceptus selection of Ensembl database to recognise co-localized gene (Thirumaran and Wright, 2014).

Fugu(Takifugu rubripes) sod1

Fugu (Takifugu rubripes) sodi	
5'gtttactgaataaccaaacttaagatcgaacaaaaggtttttcattaagatatgaaag	-239
ttttgtttctatcataacactgttcatgttgcccatgcttggatacgtcattgtttccac	-179
attttacgcaatatagtccaatatttggtaagaaatcgattgtgtgaaatcatctattgc	-119
tacactaaaatcccaactccgaataagtaactataaattattCATttatcgattcattta	-59
agcataataagtacgactttccggtccgcctcggaccacgtTAATTAAcctgtcatcgct	-1
+1	
CCTGACCACTTAAAATCGAGCTCGCGGTTTCTCTCTCTTCTGATTGGCTGGC	60
TAAGCCCGCCCACCTACCTAAGTTTGGTTGTCTCACCAGCGCGATGCTTGGCGCGTAAGT	120
GGTTTTCCAGGGGACAACTGCGACG ATGGCGATGAAGGCTGTGTGCGTGTTAAAAGGCGC	180
-MAMKAVCVLKGA	
GGGCGACACCAGCGGAACCGTGTATTTTGAGCAGGAGgtgagctgtcaggcgtcgcagga GDTSGTVYFEQE-	240
gacgettteettettaacatttaeggaegettgteeaataaeetegtgeaagettetge	300
gtgtcggtgctaaagttgctagctgagttgtttggaaccaagccgagcatcaaattatgg	360
	420
ttggtccggttccccggaggctcgttaacgccgtgtaatcacctgcaggcaattagcctt	420
cattttaccaagacatttagacattgacaaaactacaagcactttcaaactaataacctt	480 540
tgcacgatcaatgctaacatgttaatctataaccgggtggctgccgcgcctgttgcgcat	540 600
gttcaattgtgtacataaagtatataatgtcaaggttgtccgtgtaatttgggtaataaa	
cgcatgtttcagacgggtgtatgtgtccaaatatctgtcgttggaacgaac	660
aaagaactgaattaacccggaccgagttgcgttttattccgttatttttgctgagtcatc	720
gctggtcgcttccctcatctgtcgccttaaatcgcgtcttaaagtaatttttcccagtaa	780
agcgacatattaaagaaatggaacacctgtgtttcaactcgaacctttagtttctaacca	840
ttattttaccattatttaggctgttaggtgagccagaaatgcctgtttaatcctgctagt	900
ttgeteateteetag AACGAGTCTGCTCCTGTGAAGCT GACCGGGGAGATTAAAGGGCTG	960
	1000
ACCCCCGGAGAACACGGCTTCCACGTCCACGCTTTTGGAGACAATACGAATG -TPGEHGFHVHAFGDNTN	1020
cccaattgcgcacttaaatgaaccttttttgagtgttttaatggtcatttgttctgtaat	1080
cctttcccaqGTTGCATCAGTGCAGGTCCTCACTACAACCCCCACAACAAGACCCACGCT	1140
GCI-S-AGPHYNPHNKTHA-	1140
GGGCCCACTGATGCCGACAGgtaggtccaactaggactgggaggaaacgtcctggcgtga	1200
-GPTDR	
tgcagggcgccccaaatgtgtctccccttcatagttgagtcctaaactggaacaaaaata	1260
taattaatagtgctgttgggctcttgaaggggggggggg	1320
ggaaacactgaacgatgatgcagcctcattatccacgtggaaggtgaggctgtagggctg	1380
cagccctctgaggtagcaccagaggaggctctgaatgctggagtgagaagtgtcacatgc	1440
agccgtgcccctgctgctctgcacgtgcgtgctcgcacacttctacacatcccactgtgg	1500
tgctctcaatacaataaacaatataaatagtagctttgggtaggacttctgtatcccctc	1560
tgtcactaaaatgcctccaattgtcatttttaaaagtatttgtgggtttagttgtacctt	1620
aaaaatctctaagttctggcttcaagtgtggaaaaatgcactttcaagagttacaccttc	1680
ctaagtttgttctaaatctaacaagGCATCTTGGAGACCTGGGAAATGTGACTGCAGGAG	1740
	1000
CAGACAACATCGCCAAGATTGACATAAAAGACTCCATGTTGACCCTCACTGGCCCCTATT ADNIAKIDIKDSMLTLTGPY	1800
CTATAATTGGCAGGACCATGGTGgtgagccaacgtttccttctttatgatttatttata	1860
SIGRTMV-	1900
	1920
tttaggcctgaagtgaacggcgtgaatttgtgtgtttctaagATCCACGAGAAGGCCGAC	1920
GACCTGGGAAAAGGAGGCAACGAGGAGAGCCTGAAAACGGGGAACGCCGGGGGACGTTTG	1980
-DLGKGGNEESLKTGNAGGRL-	1000
GCCTGCGGGGTCATCGGCATCACTCAGTAAtcagcccgtaaataaagttgtgtaaactat	2040
-ACGVIGITQ*-	2010
tettgtaagcacttaacgagaccagtetagtagttgttcaacettgtggetteacteetg	2100
gacccagtcgggcgtgcaggagggttctgctgtttgtgacagtgtttccaaggtttccgt	2160
ggctgctgtttagatttggtcccaagtaattggaaacgcacaagtgacgtccatgtagac	2220
aatettAATAAAtgttacgtteteageeaaataagtetgaateeattattgatettt 3'	2280

Fig. 1. The nucleotide sequence of the fugu *sod1* gene. Exons of the *sod1* gene are shown in capital letters and nucleotide position is indicated the numbers end the rows. The start site of transcription is indicated by +1. The 5' upstream sequence, 3'downstream sequence and introns are shown in lower-case letters. A putative CAAT box, TATA box and polyadenylation signal (AATAAAA) are shown capital letters highlighted in yellow. The stop codon (TAA) is indicated by an asterisk.

Table II. Superoxide dismutase genes of fugu with their corresponding zebrafish query sequence ID, NCBI cDNA
ID, locations of chromosome and length of <i>sod</i> polypeptides.

Gene	Zebrafish query sequence ID	NCBI cDNA ID	Location	Number of amino acids
sod1	NP_571369.1	XP_003971372.1	Chromosome 10: 25 694 395-25 699 454	154
sod2	NP_956270.1	XP_003971923.1	Chromosome 16: 9 495 906-9 498 317	227

Fugu(Takifugu rubripes) sod1

Fu	sod1	1	GAGDTSGTVYFEQENESAPVKLTGEIKGLTP
Zf	sod1	1	WN
Me	sod1	1	T.E.N.V.NSDV.
Mo	sod1	1	D.PVQIHKASGEV.S.Q.TE
Hu	SOD1	1	D.PVQ.IINKESNGVW.SE
Fu	sod2	1	MSTKKRLCRVGQIHKC.ASLSQAVR-QVGASRHKHTLPDLTYDYGALEPHISAMQ.HH
Me	sod2	1	MLCKVWQMRSC.SILHQTVSWKVGSSRQKHTLPDLTYDYGALEPHICAMQ.HH
Zf	sod2	1	MLCRVGYVRRC.ATFNPL.GAVTSRQKHALPDLTYDYGALEPHICAMQ.HH
Hu	SOD2	1	MLSRAVCGTSRQLAPGYL.SRQKHSLPDLPYDYGALEPHINAQ.MQ.HH
Мо	sod2	1	MLCRAACSTGRRLGP.AGAA.SRHKHSLPDLPYDYGALEPHINAQ.MQ.HH
Fu	sod1	42	G-EHGFHVHAFGDNTNGCI SAGPHYNPHNKTHAGPTDAD
Zf	sod1	42	KFDGSV
Me	sod1	42	KI.VYE.E
Мо	sod1	42	QQYQTFS.K.GA.EE
Hu	SOD1	42	LEATFLSRK.GK.EE
Fu	sod2	60	SKH.ATY.NNLNVTEEKYQEALAKRDVT.QVALQ.ALRFNG.GHINHTIFWTNLS.NGGG
Me	sod2	56	SKH.ATY.NNLNVTEEKYQEALAK.DVT.QVTLQ.AL.FNG.GHINHTIFWTNLS.NGGG
Zf	sod2	54	SKH.ATY.NNLNVTEEKYQEALAK.DVTTQVSLQ.AL.FNG.GHINHTIFWTNLS.NGGG
Hu	SOD2	52	SKH. AAY. NNLNVTEEKYQEALAK. DVT. QIALQ. AL. FNG. GHINHSIFWTNLS. NGGG
Mo	sod2	52	SKH. AAY.NNLNATEEKYHEALAK.DVTTQVALQ.AL.FNG.GHINHTIFWTNLS.KGGG
Fu	sod1	80	RHLGDLGNVTAGADNIAKIDIKDSMLTLTGPYSIIGRTMVIHEKADDLGK
Zf	sod1	80	VD.SGVE.E.AS.QHE.
Me	sod1	80	VDNVT.KLIR.SDVV.VV.
Мо	sod1	80	VK.GV.NVS.E.RVIS.S.EHVQ
Hu	SOD1	80	VDK.GV.DVS.EVIS.S.DHCL.V.
Fu	sod2	120	EPQ.E.MEAIKRDFGSFQKMKEKMSA.TVAVQG.GWGWL.YSKET.LCIAACGNQ.PLQ
Me	sod2	116	EPQ.E.MEAIKRDFGSFQKMQEKLSA.TVAVQG.GWGWL.YDKES.LR.AACANQ.PLQ
Zf	sod2	114	EPQ.E.LEAIKRDFGSFQKMKEKISA.TVAVQG.GWGWL.FEKES.LRIAACANQ.PLQ
Hu	SOD2	112	EPK.E.LEAIKRDFGSFDKFKEKLTA.SVGVQG.GWGWL.FNKER.HLQIAACPNQ.PLQ
Мо	sod2	112	EPK.E.LEAIKRDFGSFEKFKEKLTAVSVGVQG.GWGWL.FNKEQ.LQIAACSNQ.PLQ

Identity% Similarity%

Fu	sod1	130	GGNEESLKTGNAGGRLACGVIGITQ	100	100
Zf	sod1	130		80	89
Me	sod1	130	AA.	74	89
Mo	sod1	130	AA.	68	81
Hu	SOD1	130	A	66	79
Fu	sod2	180	. TTGLIPLLGIDVWEHAYY. QYK. VRPDYVKAIWNVINWENVSERLQTA	9	19
Me	sod2	176	. TTGLIPLLGIDVWEHAYY. QYK. VRPDYVKAIWNVVNWENVSERLQIAKK	10	18
Zf	sod2	174	. TTGLIPLLGIDVWEHAYY.QYK.VRPDYVKAIWNVVNWENVSERFQAAKK	10	18
Hu	SOD2	172	. TTGLIPLLGIDVWEHAYY. QYK. VRPDYLKAIWNVINWENVTERYMACKK	10	18
Мо	sod2	172	. TTGLIPLLGIDVWEHAYY. QYK. VRPDYLKAIWNVINWENVTERYTACKK	9	18

Fig. 2. The percentage of sequence identity and similarity among fugu *sod1* gene and human, mouse, zebrafish, medaka *sod1* and *sod2* gene. The dots and dashes in the alignment represent sequence identity and missing amino acids respectively.

RESULTS AND DISCUSSION

Bioinformatics and computational analysis of fugu sod genes

As a known procedure in bioinformatics studies, first, statistical knowledge is collected using biological data; then, a model is generated for solving any computational modeling problem. Finally, testing and evaluation of a computational algorithm for solving a bioinformatics problem are conducted (Can, 2014). In this study, I retrieved some statistics using the Ensembl genomic database, NCBI database, BioEdit software, pairwise alignment of the BLOSUM62 matrix program and MEGA6 program.

Fugu (<i>Takifugu rubripes</i>) <i>sod2</i>	
	-239
5' tagaaatgtttcgaaaacattgaggcactaaatgacgcattgttttaaaacaacaaga	-239
tcgcaacgcaaagcctaacgtgttcgaatgtatgtgaataaaccctatttatt	
gtgccgcgacagaaaagctaaattgcagctgttcgaagtcatttgcttaatttctccgcg	-119
atcgtacaccccctca <mark>CAAT</mark> ccacaatggtacgatcctcgccgtgtcacgttg <mark>AAATT</mark> g	-59
cacatttcaagggcaatcgccactatcgagcacgactgtcccgtctgccaaacgagcact	
<u>+1</u>	
ATGAACACGCTTTGCAGAGTTGGTCAGATACACAGgtaaaagctcatatcagctcagcct	60
-MNTLCRVGQIHR	
${\tt ttcgcttctttcggcctgtgttaagctctgattgggggatttggggcagctgacttgtttc}$	120
ttgcaacatttgctcatgaacttgagacagttgccgggagttaatgccagctagct	180
tgctaacatatcaatggaagcgggtcagcccggctagtctgtggcgcaccctcaaatttt	240
${\tt gttcaactgtggctttttgggactctttagttcaagtggcaacacttgttttagctgccc}$	300
taacattcacacaaagtaagttcgtaactctcgcttcaatgcatgtgtaagctttgtatt	360
atttcttttattcgtttaaatcattggtacctcagccgacgagcttatgcaacagccggt	420
${\tt gtttatctgtctgattgttagcaggataattcgagttatcaacagactcgtggttacgca}$	480
atactttagtttgccaggtgaatttgaagtcgtcaccggtatttgcactactgccaaatg	540
agcactaagaagaggetttgcagagttggtcagatacacaagtaaagetcatattagete	600
agcatttagcctctttcggcctggatgaacctctgattagtattaaagaagtttctgaat	660
ttttttgacggggggagaacagaatgtaaattcgtgaaaagtgtagacggctacaccttgtg	720
accaacaatgtattctgcgagactctgcagatgaaatgtctggaacccagagctctgcat	780
aggcttaatttatttaaaaaaaaaaaagtaaaaattatatgaacggaaactgagcgagtc	840
cagtggggttgagactgcaggttgttgaaaatacacacatccctgttcgatgatccaagt	900
ccttcatgttctgaaatgagtccctttgctttatttcagATGTGCAGCCAGCCTGAGCCA	960
CASLS-Q GGCCGTGAGGCAGGTGGGGGGCGTCGAGACACAAGCACACGCTCCCTGACCTGACCTACGA	1020
AVRQVGASRHKHTLPDLTYD	1020
CTACGGAGCCTTGGAGCCCCATATCAGTGCAGAGATCATGCAGCTGCACCACAGCAAGCA	1080
YGALEPHISAEIMQLHHSKH	1000
CCACGCCACATATGTCAATAATCTTAACGTCACAGAGGAGAAATATCAGGAAGCATTAGC	1140
AAAGAgtatggatgcaatcatgcattatgggtgtcatggtggcaggctcaccgtctgaca	1200
K	
${\tt ttggcgcgcgattgtctttattttag} \underline{{\tt GAGATGTGACTGCACAAGTTGCTCTCCAGCCTGC}.}$	1260
R-D-V-T-A-Q-V-A-L-Q-P-A	
GCTGAGGTTTAACGGAGGTGGCCACATTAACCATACCAT	1320
$ \mathtt{L} - \mathtt{R} - \mathtt{F} - \mathtt{N} - \mathtt{G} - \mathtt{G} - \mathtt{G} - \mathtt{H} - \mathtt{I} - \mathtt{N} - \mathtt{H} - \mathtt{T} - \mathtt{I} - \mathtt{F} - \mathtt{W} - \mathtt{T} - \mathtt{N} - \mathtt{L} - \mathtt{S} - \mathtt{P}$	
AAACGGCGGAGGCGAGCCTCAGGgtaatggggccgcagctgagcatcggcctttggcgaa	1380
NGGEPQ	1440
acctccatgtctaacgtccacatgtcacttcctgtagGGGGGCTGATGGAGGCCATTAAG	1440
	1500
CGGGACTTTGGCTCATTCCAGAAGATGAAGGAGAAGATGTCCGCCGCTACGGTTGCAGTG -RDFGSFQKMKEKMSAAT-VAV-	1200
_RDF_G_S_F_G_Q_R_MRR_R_R_S_R_R_R_R_R_R_R_R_R_R_R_R	1560
-QGSGWGWLGYSKETGRLCIA-	1000
GCCTGTGGCAACCAGGACCCCCTCCAAGGAACTACAGgtgggttcaacagcttttgtttc	1620
-ACGNQDPLQGTT	1010
atcgttttctcttcccgtgtaacatcggcgccatatgaataattttctctttttcccagG	1680
TCTCATCCCGCTCCTTGGCATCGACGTATGGGAGCACGCCTACTATCTTCAGTACAAAAA	1740
LIPLLGIDVWEHAYYLQYKN	
TGTGCGGCCAGACTATGTTAAGGCCATCTGGAATGTGATCAACTGGGAGAATGTGAGCGA	1800
VRPDYVKAIWNVINWENVSE	
ACGTCTCCAAACTGCCAAGAAGTAGgagcccaacccgtcaccccagaactgaccctcagt	1860
RLQTAKK*-	
tatgtggetgetgegtatgttgtaategtteeattagetatteaaaeatttteagegttt	1900
gttgcagaataaaccaccttgttgggaaaaacaccatgttacacctgcagatcgaggcac	1960
attttgagctgtatttatctgattccgcttcccagatttaaaatgtaaatttacatagat	2020
ctgaatctgatgattctacacaaatatggccatgtttcagccagattttttccgtttgtc	2080
taaataattetacatgagegtttgetttgtgttttgatgtattat <mark>AATAAA</mark> atgatacaa	2140
ctgcagttgaactggagtgaaagctgttgctcactactgtgtgctgtgaaagatttaata	2200
atggtgggaaataaaatattcatgttaaatggtgtgtattaatgaaacaatgacaacttt	2260
tataattattagtcttaccagaacaataattagcttaaagtgtataagtgtattcgtcca	2320
agtgcaaataataaagaacctggagaaaaaagcattttaacgtcctgaagggcacctgct	2380
ctgcgcaccatg 3'	2440

Fig. 3. The nucleotide sequence of the fugu *sod2* gene. Exons of the *sod2* gene are shown in capital letters and nucleotide position is indicated the numbers end the rows. The start site of transcription is indicated by +1. The 5' upstream sequence, 3'downstream sequence and introns are shown in lower-case letters. A putative CAAT box, TATA box and polyadenylation signal (AATAAAA) are shown capital letters highlighted in yellow. The stop codon (TAG) is indicated by an asterisk.

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Fugu (Takifugu rubripes)sod2

Fug	gu (Ta	kifu	ıgu rubripes)sod2		
Fu	sod2	1	MNTLCRVGQIHRCAASLSQAVR-QVGASRHKHTLPDLTYDYGALEPHISA	EIMOLE	HSKH
Te	sod2	1	м		
Me	sod2	1	MK.W.MRSSI.H.T.SWKSQ		
Zf	sod2	1	MYVRTFNPLLGAVTQAC.		
Hu	SOD2	1	M.S.AVCGTSRQ.APVLGYLG.Q.SPN.	Q	
Мо	sod2	1	M AACSTGRR . GPVAGAAG S P	Q	
Те	sod1	1	MFGFPAS.V.PCVSFLE.TTAKMVIKAVCVLKGA.ETSGTVYF		
Hu	SOD1	1	MATKAVCVLKGD. PVQGI.NF	. QKESN	IGPVK
Mo	sod1	1	MAMKAVCVLKGD . PVQGT . HF	. QKASO	EPVV
Me	sod1	1	MVLKAVCVLKGT.ETNGVVNF	.QESDS	APVK
Fu	sod1	1	MAMKAVCVLKGA.DTSGTVYF	. QENES	APVK
Zf	sod1	1	MVNKAVCVLKGT. EVTGTVYF	NQEGER	KPVK
-	sod2	60			
	sodz sod2	58	HATYVNNLNVTEEKYQEALAKRDVTAQVALQPALRFNGGGHINHTIFWTN 		
	sod2	58			
	sod2	57			
	SOD2	55			
	sod2	55			
	sodi	55	LTGEIKG. TAG. HGFHVHAFGD. TN. C. SAGPHYNF		
	SOD1	32	VWGSIKG.TEGLHGFHVHEFGD.TA.CTSAGPHINP		
	sod1	32	LSGOITG. TEGOHGFHVHOYGD. TO. CTSAGPHFNF		
	sod1	32	VTGEIKG.TPGKHGFHIHVYGD.TV.CVSAGPHFNF		
	sodi	32	LTGEIKG.TPG.HGFHVHAFGD.TN.C.SAGPHYNF		
	sodi	32	VTGEITG. TPGKHGFHVHAFGD. TN. C. SAGPHFNF		
	Jour				
Fu	sod2	120	GELMEAIKRDFGSFQKMKEKMSAATVAVQGSGWGWLGYSKETGRLCIAAC	GNQDPI	QGTT
Те	sod2	118	T		
Me	sod2	119	DSRV	A	
Zf	sod2	117	L	A	
Hu	SOD2	115	L	P	
Mo	sod2	115	L	s	
Te	sod1	100	ENSLKRHVG.L.NVTAEADQI.KIDITD.VISLH.KFSIITMVIHE	KAD . LO	K.GN
Hu	SOD1	78	ERHVG. L. NVTAD. DGV. D. SIED. VISLS. DHCII TLVVHE	KAD . LO	K.GN
Mo	sod1	78	ERHVG. L. NVTAG. DGV. N. SIEDRVISLS. EHSII TMVVHE	KQD.LC	K.GN
Me	sod1	78	ARHVG.L.NVTAGDNNV.KIDITDKLIRLS.PDSIVTVVVHE		
Fu	sod1	78	ADRHLG.L.NVTAGADNI.KIDIKD.MLTLT.PYSIITMVIHE	KAD.LO	K.GN
Zf	sod1	78	SVRHVG.L.NVTADASGV.KIEIEDAMLTLS.QHSIITMVIHE	KED.LO	K.GN
E.	sod2	100	Ident GLIPLLGIDVWEHAYYLQYKNVRPDYVKAIWNVINWENVSERLQTAKK	ity% \$ 100	imilarity%
	sod2		GLIPLIGIDVWEHAIILQIKNVRPDIVKAIWNVINWENVSERLQIAKK	95	91
	sod2			86	89
	sod2		VF.A	85	85
	SOD2			76	82
	sod2			76	19
	sod1		EES.KTG.AGGRLACGVIGITO	9	17
	SOD1		EES.KTG.AGGKLACGVIGITQ	10	18
	sod1		EESTRIG.AGSRIACGVIGIAQ	9	18
	sod1		DES.KTG.AGARLACGVIGIAQ	8	17
me	3041	1.72	DED. KIG. AGAMIACOVIGIAG	0	17
Fu	sod1	132	EES.KTG.AGGRLACGVIGITO	8	16
				-	
Zf	sod1	132	EES.KTG.AGGRLACGVIGITQ	7	15

Fig. 4. The percentage of sequence identity and similarity among fugu *sod2* gene and human, mouse, zebrafish, medaka *sod1* and *sod2* gene. The dots and dashes in the alignment represent sequence identity and missing amino acids respectively.

The cDNA sequence was obtained from the Ensembl genomic database to determine the exon-intron structure of the fugu *sod* genes, and it was found that these genes had five exons separated by four introns. It was determined that introns of both genes followed the gt-ag rule. Moreover, they have putative TATA and CAAT boxes and polyadenylation signals. The results clearly show that *sod* genes in fugu exhibit a highly conserved gene structure (Figs. 1 and 3).

I searched the ESTs of *sod1* and *sod2* by BLAST searches in the NCBI genomic database using cDNA nucleotide sequences obtained from the Ensembl genomic database (Table I). However, as shown in Table I, there was no EST for these genes. The sequence identity and similarity among the fugu and human, mouse, zebrafish, and medaka *sod* genes are given in Figures 2 and 4. The

highest identity and similarity rates for fugu sod1 and sod2 genes were determined with their orthologs (tetraodon, medaka, and zebrafish). After that, a maximum-likelihood phylogenetic tree was built based on identity and similarity results (Fig. 5). At the end of the phylogeny analysis, high phylogenetic clustering was seen between the sod genes in fugu and their orthologs. Conserved gene synteny evidence was determined for sod genes of fugu and Sod/sod genes of other teleost fishes and human (Fig. 6). The syntenic genes of fugu sod1 and sod2, located on chromosomes 15 and 16, exhibit conserved gene synteny with human SOD1 and SOD2 located on chromosomes 21 and 6, medaka sod1 and sod2 located on chromosomes 14 and 24, and zebrafish sod1 and sod2 located on chromosomes 10 and 20. The results of conserved gene synteny between sod genes in fugu and their orthologs in medaka, zebrafish, and human show that teleost fish lost duplicated copies of the sod1 and sod2 genes after tsWGD because of mutations. This would be an interesting result for future studies on whether all teleost species have single copies of sod genes and their transcriptional controls.

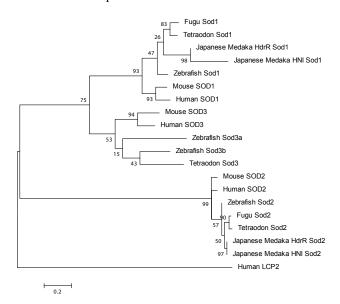


Fig. 5. Phylogenetic relationships between *sod1* and *sod2* sequence from fugu and *sod* sequences from other vertebrates. Accession numbers of the sequences used for phylogenetic tree are the following: Fugu Sod1/Sod2: HE602549, HE602550, Tetraodon Sod1/Sod2/Sod3: H3CX53, Q4S6V6, H3D497; Japanese medaka HDrR Sod1/Sod2: XP_004076261, GCA_002234675, Zebrafish Sod1/Sod2/Sod3: NP_571369, NP_956270, XP_001332758, Mouse Sod1/Sod2/Sod3: NP_035564, NP_038699.2, NP_035565, Human Sod1/Sod2/Sod3: NP_000445, NG_008729, NP_003093.

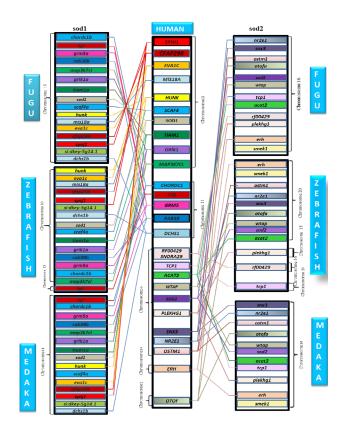


Fig. 6. Conserved gene synteny of fugu sod genes.

CONCLUSION

Antioxidant enzymes are gaining more importance, and they have been studied more in recent years. When antioxidant levels and production of ROS are in balance in normal conditions, the harmful effect of free radicals can be observed with oxidative stress-induced diseases (Scandalios, 1993). Stress responses in fishes can manifest multifaceted levels involving the actions of different sets of genes and their products. Understanding the genetic characteristics associated with model organisms that exhibit a stress response is important for molecular studies. Identification and characterization of the stress genes that are differentially expressed between stress-tolerant and intolerant fish will provide important genetic markers that may be used in aquaculture selection programs to help improve stress tolerance, as well as serving as a model for other vertebrates, including human (Iwama et al., 1999). However, it is known that oxidative stress has a big role in more than 100 diseases in human as their reason or effect (Halliwell et al., 1992; Gutteridge, 1993; Poljsak et al., 2013). For this purpose, I identified sod genes in fuguserving as a model organism—using bioinformatic tools;

the results can illuminate the path for future studies on molecular stress responses in fish.

Statement of conflict of interest

The authors declare there is no conflict of interest.

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