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Interaction between MTHFR Polymorphisms and Maternal Age Increases the Risk of **Congenital Heart Defects in Down Syndrome**

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ABSTRACT

Congenital heart disease (CHD) is responsible for one-third of all congenital anomalies in newborns and is the most frequent cause of infant deaths. Several cohort studies show that down syndrome (DS) and CHD are associated, and maternal hyperhomocysteinemia is an independent risk factor for CHD. In Saudi Arabia CHD represents one of the most important health problems. Here we examined the association between methylenetetrahydrofolate reductase (MTHFR) gene polymorphisms and CHD and DS coinheritance in patients from Al Madinah, Saudi Arabia. MTHFR rs1801133 and rs1801131 polymorphisms were genotyped in 99 CHD patients with or without DS and 126 ethnically matched controls by allelic discrimination. Of 99 patients with CHD, 26 had DS. MTHFR rs1801133 and rs1801131 genotypes and alleles were not significantly different between controls and CHD patients. Further, in CHD individuals, these genotypes failed to show any significant association with DS. However, maternal age increased the risk of CHD in DS (OR=5.32; 95% CIs 1.43-19.82; p=0.013). Mantel-Haenszel analysis showed that MTHFR polymorphisms confounded the effect of maternal age CHD in DS. MTHFR polymorphisms appear to be risk factors for CHD in DS.

INTRODUCTION

ongenital heart disease (CHD) comprises one-third of all congenital anomalies in newborns and is the most frequent cause of infant deaths. Long-term morbidity and mortality from CHD pose a substantial challenge to healthcare systems globally (Dolk et al., 2011). The exact etiology of CHD remains uncertain, the disease is heterogeneous, but it is believed to be multifactorial: environmental factors contribute (Gladki et al., 2015; Gorini et al., 2014; Huhta and Linask, 2013); and twin studies suggest familial aggregation of CHD (Kuo et al., 2017). Specific recurrence patterns and phenotypes also suggest that genetically-determined developmental mechanisms exert a strong influence in CHD etiology (Ellesøe et al., 2018).

Several cohort studies have established a relationship between Down syndrome (DS) and CHD (Ferencz et al., 1989; Kidd et al., 1993), with 4%-10% of all CHD cases



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Authors' Contribution

SJC and KMAH presented the concept of the study. SJC and AMA designed methodology. AHAM and MKAH did data curation. SJC wrote the manuscript. KMAH reviewed and edited the manuscript.

Key words

Congenital heart disease, Down syndrome, Methylenetetrahydrofolate reductase, Polymorphism, Maternal age, Saudi Arabia.

associated with DS and 40%-60% of DS patients having CHD (de Rubens-Figueroa et al., 2003). Several lines of evidence indicate that maternal hyperhomocysteinemia is an independent risk factor for CHD (Malik et al., 2017), and recent population studies in humans have indicated that folic acid supplementation before and/or during pregnancy can decrease CHD risk (Czeizel et al., 2015; Leirgul et al., 2015; Li et al., 2013). This evidence prompted several follow-on studies examining the role of folic acid in CHD risk (Parnell and Correa, 2017; Xu et al., 2016; Feng et al., 2015).

Folates serve as cofactors for nucleotide synthesis and co-substrates of DNA methyltransferases (Stover and Field, 2011). DNA methylation plays a key role in embryonic development, and abnormally methylated genes have been detected in fetuses with both isolated and syndromic heart malformations (Serra-Juhe et al., 2015). Methylenetetrahydrofolate reductase (MTHFR; EC 1.5.1.20) is a key enzyme in folate and homocysteine (Hcy) metabolism (Bhaskar et al., 2011). The gene encoding human *MTHFR* is known to have two important functional polymorphisms, C677T/A222V/rs1801133 and A1298C/ Glu429Ala/rs1801131, which generate a thermolabile form

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of MTHFR that subsequently increases total plasma Hcy and decreases serum folate levels (Goyette *et al.*, 1994). A recent computational modeling study demonstrated that the rs1801133 polymorphism causes conformational changes in the tertiary structure of MTHFR by reducing its FAD-binding ability (Abhinand *et al.*, 2016). However, results from studies examining the relationship between *MTHFR* rs1801133 and rs1801131 polymorphisms and CHD risk have been inconsistent (Zhang *et al.*, 2018; Asim *et al.*, 2017; Yu *et al.*, 2017; Guo *et al.*, 2017). We recently highlighted the high burden of DS and CHD in the Al Madinah region of Saudi Arabia (Abdulhadi *et al.*, 2016). Here we investigated associations between *MTHFR* polymorphisms (rs1801133 and rs1801131) and the risk of CHD and DS co-occurrence in a Saudi population.

PATIENTS AND METHODS

Ethical statement

The Institutional Ethics Committees of Maternity and Children Hospital and Centre for Genetics and Inherited Diseases (CGID), Taibah University, Al Madinah, Kingdom of Saudi Arabia granted ethical approval. As many of the subjects in the CHD group were minors (<18 years old), written informed consent was obtained from the parents or legal guardians of the children enrolled in this study as necessary.

Study population

Ninety-nine CHD patients with (n=26) or without (n=73) DS attended the Pediatric Cardiology Clinic at the Maternity and Children Hospital, Al Madinah, Kingdom of Saudi Arabia were enrolled in this study.

Participants with mosaic, translocation of chromosome 21, non-cardiac congenital anomalies were excluded from this study. The main inclusion criteria for DS was cases had full trisomy 21 confirmed by karyotype and/or Fluorescent *in situ* Hybridization (FISH) for chromosome 21. CHD and CHD in DS samples were collected after checking the family history, physical and clinical examination, electrocardiograms (ECG), chest x-rays, 2D-echocardiography, Karyotypes or FISH data from the patient's case sheet. One hundred and twenty-six healthy subjects without CHD or a family history of CHD were recruited as controls. All participants were of Saudi Arabian ethnicity.

Genotyping

Two ml of whole peripheral blood was collected from each participant after obtaining informed consent. DNA was extracted using the QIAamp DNA Mini Kit (Qiagen, Hilden, Germany). Two MTHFR SNPs [rs1801133: C 1202883 20 and rs1801131: C 850486 20] were genotyped using primers and probes purchased from Applied Biosystems, Foster City, CA. Reactions were carried out in a final volume of 5 µL (2.5 µL TaqMan PCR Master Mix, 0.125 µL TaqMan Genotyping Assay Mix, 1.375 μ L nuclease free water, and 1 μ L of 10 ng/ μ L DNA) in a 96-well optical microplate (Applied Biosystems). At least two no-template controls without any DNA were included on each plate. The PCR conditions were initial denaturation at 95°C for 10 min followed by 40 denaturation cycles (95°C for 15 s) and annealing/extension (60°C for 1 min). PCR amplification was performed using Applied Biosystems' StepOnePlus Real-Time PCR system, and fluorescence was also measured using its

Table I.- The distribution of MTHFR genotypes in control and CHD subjects.

SNP	Genotype	Control (n=126)	CHD (n=99)	OR (95% CI)	<i>p</i> -value
rs1801133	CC	88 (69.8)	69 (69.7)	Reference	1.0*
	СТ	33 (26.2)	26 (26.3)	0.90 (0.48-1.75)	
	TT	5 (4.0)	4 (4.0)	0.85 (0.21-3.46)	
	CT+TT Vs. CC	38 (30.2)	30 (30.3)	0.93 (0.52-1.67)	0.810
	TT Vs. CC+CT	5 (4.0)	4 (4.0)	1.01 (0.26-3.97)	0.990
	MAF	17.1	17.2		
	HWp	0.402	0.445		
rs1801131	AA	57 (45.2)	51 (51.5)	Reference	0.618*
	AC	52 (41.3)	35 (35.4)	0.73 (0.40-1.33)	
	CC	17 (13.5)	13 (13.1)	0.82 (0.34-1.94)	
	AC+CC Vs. AA	69 (54.7)	48 (48.5)	1.29 (0.75-2.20)	0.360
	CC Vs. AA+AC	17 (13.5)	13 (13.1)	1.03 (0.44-2.12)	0.920
	MAF	34.1	30.8		
	HWp	0.357	0.089		

OR, odds ratio; CI, confidence interval; MAF, minor allele frequency; ^aHWp, Hardy-Weinberg p value.

Gene	Genotype	СНД		Univariate	<i>p</i> -value
		Without DS (n=73)	With DS (n=26)	OR (95% CI)	-
rs1801133	CC	52	17	Reference	
	СТ	18	8	2.38 (0.58-9.66)	
	TT	3	1	1.03 (0.08-13.24)	0.831
rs1801131	AA	38	13	Reference	
	AC	27	8	1.43 (0.39-5.19)	
	CC	8	5	4.64 (0.86-25.06)	0.542
Sex	Female	35	13	Reference	
	Male	38	13	1.12 (0.42-3.54)	0.707
Paternal age	< 40 y	51	9	Reference	
	\geq 40 y	22	17	2.45 (0.66-9.19)	0.182
Maternal age	< 35 y	61	11	Reference	
-	\geq 35 y	12	15	5.32 (1.43-19.82)	0.013

Table II.- The distribution of *MTHFR* genotypes and other risk factors among CHD patients and their association with Down syndrome.

OR, odds ratio; CI, confidence interval.

Sequence Detection Software (SDS) v. 2.3. A genotype call rate over 99% was considered for the analysis. As a quality control measure, 10% of samples selected at random were included for duplicate genotyping.

Statistical analyses

Allele frequencies of both polymorphisms in CHD and control groups were calculated by the gene counting method. The genotype distribution for each polymorphism was evaluated for Hardy–Weinberg equilibrium. The strength of the association between *MTHFR* gene polymorphisms and CHD was evaluated using the χ^2 test and odds ratios (OR) with 95% confidence intervals (CI). The influence of different genotypes on the relationship between DS and maternal age and their interaction was examined using the Mantel-Haenszel stratified analysis. All statistical analyses were performed in SPSS v. 14.0 (IBM Statistics, Chicago, IL).

RESULTS

Of 99 patients with CHD, 26 (26.3%) had DS. Among DS patients, atrioventricular septal defect (AVSD) was the most common cardiac defect followed by ventricular septal defect (VSD) and atrial septal defect (ASD). The distributions of *MTHFR* rs1801133 and rs1801131 genotypes and alleles in CHD and control groups are shown in Table I. The *MTHFR* rs1801133 (p=0.402) and rs1801131 (p=0.357) genotype distributions in the control group followed HardyWeinberg equilibrium (p=0.359).

Genotypes and alleles were not statistically different between controls and individuals with CHD (Table I). *MTHFR* rs1801133 and rs1801131 polymorphisms were not associated with CHD risk (Table I) and, in CHD individuals, *MTHFR* rs1801133 and rs1801131 genotypes failed to show any significant association with DS (Table II).

Univariate analysis showed that increased maternal age (\geq 35 years) contributed to CHD risk in DS patients (OR: 5.32, 95% CI: 1.43-19.82; *p*=0.013), while increased paternal age (\geq 40 years) had no effect on CHD in DS patients (OR: 2.45, 95% CI: 0.66-9.19; *p*=0.182) (Table II). Significant heterogeneity in the effect of maternal age on DS was observed among different genotypes of *MTHFR* rs1801133 and rs1801131 polymorphisms. The M-H combined OR for maternal age and rs1801133 (*p*<0.001) was 6.33 and for maternal age and rs1801131 was 7.42 (*p*<0.001) (Table III).

Table III.- Association between CHD with or without down syndrome and maternal age stratified by *MTHFR* genotypes.

Gene	Genotype	OR (95% CI)	<i>p</i> -Value*
Maternal a			
rs1801133	CC	11.78 (3.29-42.14)	< 0.001
	СТ	3.50 (0.59-20.68)	
	TT	-	
	M-H combined	6.33 (2.37-16.93)	
rs1801131	AA	9.96 (2.37 -41.86)	< 0.001
	AC	5.75 (1.00-32.95)	
	CC	4.47 (0.30-73.38)	
	M-H combined	7.42 (2.67-20.64)	

CHD, congenital heart disease; M-H, *Mantel-Haenszel.**Homogeneity test *p* value.

DISCUSSION

The pathogenesis of CHD is complex and remains poorly understood due to its multifactorial etiology. Individuals with DS show several common cardiac malformations such as VSDs, atrioventricular canals, and the tetralogy of Fallot (Tandon and Edwards, 1973). As folate serves as a DNA methyltransferase co-substrate, reduced folate causes hypomethylation and consequently abnormal segregation of chromosomes or chromosome nondisjunction (Fenech, 2011; James *et al.*, 1999; Blom and Smulders, 2011). One common factor in these folate abnormalities is *MTHFR* rs1801133 polymorphisms, with decreased enzyme activity causing impaired remethylation of Hcy to methionine and subsequent hyperhomocysteinemia (Yigit *et al.*, 2013).

However, the association between maternal genetic polymorphisms in folate metabolism genes and DS risk is still controversial (Yang et al., 2013; Brandalize et al., 2009, 2010). Significantly elevated Hcy levels in mothers of children with CHD suggest that maternal hyperhomocysteinemia is an independent risk factor for CHD (Malik et al., 2017; Lu et al., 2011; Verkleij-Hagoort et al., 2007). Due to the high prevalence of CHDs in neonates born with DS and DS survivors in Al Madinah, CHDs are considered as an important health problem in Saudi Arabia, a problem further exacerbated by widespread consanguinity in this region (Abdulhadi et al., 2016). Thus, maternal supplementation with folic acid is likely to be associated with a reduced risk of CHD in DS. Given this perspective and to inform management, we considered the association between MTHFR polymorphisms and CHD in DS susceptibility in a Saudi population.

We did not identify any significant association between MTHFR polymorphisms and CHD risk. Both polymorphisms had a confounding effect on the relationship between CHD in DS and maternal age. The presence of the maternal MTHFR rs1801133 "T" allele was associated with an increased risk of DS in a Brazilian population (Brandalize et al., 2009), while another study of a Croatian population reported no association between maternal MTHFR polymorphisms and CHD in DS (Bozovic et al., 2011; Elsayed et al., 2013). MTHFR rs1801131 was over-transmitted in patients with CHD in DS ASDs (Locke et al., 2010). A meta-analysis showed that MTHFR rs1801131 polymorphisms reduced the risk of CHD in patients without DS (Yu et al., 2017), while another suggested that the maternal MTHFR rs1801133 "T" allele was a risk factor for DS-affected pregnancies (Rai et al., 2014). Finally, a recent study showed that the MTHFR gene was twice as likely to be promoter methylated in mothers of children with DS with CHD than

mothers of children with DS without CHD (Asim *et al.*, 2017).

The following strengths and limitations must be taken into consideration when interpreting our results. Significant strengths include the inclusion of subjects with CHD diagnosed by echocardiography and clinical cardiac examination and DS by karyotype or FISH for chromosome 21. However, the study also has several limitations: plasma Hcy levels were not determined and correlated with DS-related CHD or *MTHFR* variants, and maternal *MTHFR* gene polymorphisms were not tested. Further, the study is relatively small and retrospective, which may impact on the validity of the results.

CONCLUSION

In conclusion, polymorphisms in *MTHFR* may interact with other confounding variables and contribute to the increased risk of CHD in DS.

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Statement of conflict of interests

The authors have no conflicts of interest.

REFERENCES

- Abdulhadi, H.A., Sahar, A.F.H., Lama, M.E.S., Justin, C., Yousef, A. and Khalid, M.A., 2016. Congenital heart disease in Saudi Arabia: the role of molecular genetics with a focus on down syndrome. *Aust. J. Basic appl. Sci.*, **10**: 98-109.
- Abhinand, P.A., Shaikh, F., Bhakat, S., Radadiya, A., Bhaskar, L.V., Shah, A. and Ragunath, P.K., 2016. Insights on the structural perturbations in human MTHFR Ala222Val mutant by protein modeling and molecular dynamics. *J. Biomol. Struct. Dynam.*, 34: 892-905. https://doi.org/10.1080/07391102.201 5.1057866
- Asim, A., Agarwal, S., Panigrahi, I., Saiyed, N. and Bakshi, S., 2017. MTHFR promoter hypermethylation may lead to congenital heart defects in down syndrome. *Intractable Rare Dis. Res.*, 6: 295-298. https://doi.org/10.5582/ irdr.2017.01068
- Bhaskar, L.V., Murthy, J. and Venkatesh Babu, G., 2011. Polymorphisms in genes involved in folate metabolism and orofacial clefts. *Arch. Oral*

Biol., **56**: 723-737. https://doi.org/10.1016/j. archoralbio.2011.01.007

- Blom, H.J. and Smulders, Y., 2011. Overview of homocysteine and folate metabolism. With special references to cardiovascular disease and neural tube defects. J. Inherit. Metab. Dis., 34: 75-81. https:// doi.org/10.1007/s10545-010-9177-4
- Bozovic, I.B., Vranekovic, J., Cizmarevic, N.S., Mahulja-Stamenkovic, V., Prpic, I. and Brajenovic-Milic, B., 2011. MTHFR C677T and A1298C polymorphisms as a risk factor for congenital heart defects in down syndrome. *Pediat. Int. Off. J. Japan Pediat. Soc.*, **53**: 546-550. https://doi. org/10.1111/j.1442-200X.2010.03310.x
- Brandalize, A.P., Bandinelli, E., dos Santos, P.A., Roisenberg, I. and Schuler-Faccini, L., 2009. Evaluation of C677T and A1298C polymorphisms of the MTHFR gene as maternal risk factors for down syndrome and congenital heart defects. *Am. J. med. Genet. A*, **149A**: 2080-2087. https://doi. org/10.1002/ajmg.a.32989
- Brandalize, A.P., Bandinelli, E., dos Santos, P.A. and Schuler-Faccini, L., 2010. Maternal gene polymorphisms involved in folate metabolism as risk factors for down syndrome offspring in Southern Brazil. *Dis. Mark.*, 29: 95-101. https:// doi.org/10.1155/2010/250324
- Czeizel, A.E., Vereczkey, A. and Szabo, I., 2015. Folic acid in pregnant women associated with reduced prevalence of severe congenital heart defects in their children: A national population-based casecontrol study. *Eur. J. Obstet. Gynecol.Reprod. Biol.*, **193**: 34-39. https://doi.org/10.1016/j. ejogrb.2015.06.024
- de Rubens-Figueroa, J., del Pozzo-Magana, B., Pablos-Hach, J.L., Calderon-Jimenez, C. and Castrejon-Urbina, R., 2003. Heart malformations in children with Down syndrome. *Rev. Esp. Cardiol.*, 56: 894-899.
- Dolk, H., Loane, M. and Garne, E., 2011. Congenital heart defects in Europe: Prevalence and perinatal mortality, 2000 to 2005. *Circulation*, **123**: 841-849. https://doi.org/10.1161/ CIRCULATIONAHA.110.958405
- Ellesøe, S.G., Workman, C.T., Bouvagnet, P., Loffredo, C.A., McBride, K.L., Hinton, R.B., van Engelen, K., Gertsen, E.C., Mulder, B.J.M., Postma, A.V., Anderson, R.H., Hjortdal, V.E., Brunak, S. and Larsen, L.A., 2018. Familial co-occurrence of congenital heart defects follows distinct patterns. *Eur: Heart J.*, **39**: 1015-1022. https://doi. org/10.1093/eurheartj/ehx314

- Elsayed, G.M., Elsayed, S.M. and Ezz-Elarab, S.S., 2013. Maternal MTHFR C677T genotype and septal defects in offspring with Down syndrome: A pilot study. *Egyptian J. med. Hum. Genet.*, **15**: 39-44. https://doi.org/10.1016/j.ejmhg.2013.09.003
- Fenech, M., 2011. Micronuclei and their association with sperm abnormalities, infertility, pregnancy loss, pre-eclampsia and intra-uterine growth restriction in humans. *Mutagenesis*, 26: 63-67. https://doi.org/10.1093/mutage/geq084
- Feng, Y., Wang, S., Chen, R., Tong, X., Wu, Z. and Mo, X., 2015. Maternal folic acid supplementation and the risk of congenital heart defects in offspring: A meta-analysis of epidemiological observational studies. *Scient. Rep.*, **5**: 8506. https://doi. org/10.1038/srep08506
- Ferencz, C., Neill, C.A., Boughman, J.A., Rubin, J.D., Brenner, J.I. and Perry, L.W., 1989. Congenital cardiovascular malformations associated with chromosome abnormalities: An epidemiologic study. J. Pediatr., 114: 79-86. https://doi. org/10.1016/S0022-3476(89)80605-5
- Gladki, M.M., Skladzien, T. and Skalski, J.H., 2015. The impact of environmental factors on the occurrence of congenital heart disease in the form of hypoplastic left heart syndrome. *Kardiochir Torakochirur. Pol.*, **12**: 204-207.
- Gorini, F., Chiappa, E., Gargani, L. and Picano, E., 2014. Potential effects of environmental chemical contamination in congenital heart disease. *Pediatr: Cardiol.*, **35**: 559-568. https://doi.org/10.1007/ s00246-014-0870-1
- Goyette, P., Sumner, J.S., Milos, R., Duncan, A.M., Rosenblatt, D.S., Matthews, R.G. and Rozen, R., 1994. Human methylenetetrahydrofolate reductase: Isolation of cDNA, mapping and mutation identification. *Nat. Genet.*, 7: 195-200. https://doi. org/10.1038/ng0694-195
- Guo, Q.N., Wang, H.D., Tie, L.Z., Li, T., Xiao, H., Long, J.G. and Liao, S.X., 2017. Parental genetic variants, MTHFR 677C>T and MTRR 66A>G, associated differently with fetal congenital heart defect. *Biomed. Res. Int.*, 2017: 3043476. https:// doi.org/10.1155/2017/3043476
- Huhta, J. and Linask, K.K., 2013. Environmental origins of congenital heart disease: The heart-placenta connection. *Semin. Fetal Neonatal. Med.*, 18: 245-250. https://doi.org/10.1016/j.siny.2013.05.003
- James, S.J., Pogribna, M., Pogribny, I.P., Melnyk, S., Hine, R.J., Gibson, J.B., Yi, P., Tafoya, D.L., Swenson, D.H., Wilson, V.L. and Gaylor, D.W., 1999. Abnormal folate metabolism and mutation in

the methylenetetrahydrofolate reductase gene may be maternal risk factors for down syndrome. *Am. J. clin. Nutr.*, **70**: 495-501. https://doi.org/10.1093/ ajcn/70.4.495

- Kidd, S.A., Lancaster, P.A. and McCredie, R.M., 1993. The incidence of congenital heart defects in the first year of life. *J. Paediat. Child Hlth.*, **29**: 344-349. https://doi.org/10.1111/j.1440-1754.1993. tb00531.x
- Kuo, C.F., Lin, Y.S., Chang, S.H., Chou, I.J., Luo, S.F., See, L.C., Yu, K.H., Huang, L.S. and Chu, P.H., 2017. Familial aggregation and heritability of congenital heart defects. *Circulation J.*, 82: 232-238. https://doi.org/10.1253/circj.CJ-17-0250
- Leirgul, E., Gildestad, T., Nilsen, R.M., Fomina, T., Brodwall, K., Greve, G., Vollset, S.E., Holmstrom, H., Tell, G.S. and Oyen, N., 2015. Periconceptional folic acid supplementation and infant risk of congenital heart defects in Norway 1999-2009. *Paediat. Perinat. Epidemiol.*, 29: 391-400. https:// doi.org/10.1111/ppe.12212
- Li, X., Li, S., Mu, D., Liu, Z., Li, Y., Lin, Y., Chen, X., You, F., Li, N., Deng, K., Deng, Y., Wang, Y. and Zhu, J., 2013. The association between periconceptional folic acid supplementation and congenital heart defects: A case-control study in China. *Prevent. Med.*, **56**: 385-389. https://doi. org/10.1016/j.ypmed.2013.02.019
- Locke, A.E., Dooley, K.J., Tinker, S.W., Cheong, S.Y., Feingold, E., Allen, E.G., Freeman, S.B., Torfs, C.P., Cua, C.L., Epstein, M.P., Wu, M.C., Lin, X., Capone, G., Sherman, S.L. and Bean, L.J., 2010. Variation in folate pathway genes contributes to risk of congenital heart defects among individuals with down syndrome. *Genet. Epidemiol.*, 34: 613-623. https://doi.org/10.1002/gepi.20518
- Lu, Y., Wang, H. and Wang, X., 2011. Relationship of hyperhomocysteinemia in pregnant rats and congenital heart defects in the newborn rats. *Zhong Nan Da Xue Xue Bao Yi Xue Ban*, **36**: 68-73.
- Malik, R.A., Lone, M.R., Ahmed, A., Koul, K.A. and Malla, R.R., 2017. Maternal hyperhomocysteinemia and congenital heart defects: A prospective case control study in Indian population. *Indian Heart J.*, 69: 17-19. https://doi.org/10.1016/j.ihj.2016.07.014
- Parnell, A.S. and Correa, A., 2017. Analyses of trends in prevalence of congenital heart defects and folic acid supplementation. J. Thorac. Dis., 9: 495-500. https://doi.org/10.21037/jtd.2017.03.16
- Rai, V., Yadav, U., Kumar, P., Yadav, S.K. and Mishra, O.P., 2014. Maternal methylenetetrahydrofolate

reductase C677T polymorphism and down syndrome risk: A meta-analysis from 34 studies. *PLoS One*, **9**: e108552. https://doi.org/10.1371/ journal.pone.0108552

- Serra-Juhe, C., Cusco, I., Homs, A., Flores, R., Toran, N. and Perez-Jurado, L.A., 2015. DNA methylation abnormalities in congenital heart disease. *Epigenetics*, **10**: 167-177. https://doi.org/10.1080/ 15592294.2014.998536
- Stover, P.J. and Field, M.S., 2011. Trafficking of intracellular folates. Adv. Nutr., 2: 325-331. https:// doi.org/10.3945/an.111.000596
- Tandon, R. and Edwards, J.E., 1973. Cardiac malformations associated with down's syndrome. *Circulation*, 47: 1349-1355. https://doi. org/10.1161/01.CIR.47.6.1349
- Verkleij-Hagoort, A., Bliek, J., Sayed-Tabatabaei, F., Ursem, N., Steegers, E. and Steegers-Theunissen, R., 2007. Hyperhomocysteinemia and MTHFR polymorphisms in association with orofacial clefts and congenital heart defects: a meta-analysis. *Am. J. med. Genet. A*, **143A**: 952-960. https://doi. org/10.1002/ajmg.a.31684
- Xu, A., Cao, X., Lu, Y., Li, H., Zhu, Q., Chen, X., Jiang, H. and Li, X., 2016. A meta-analysis of the relationship between maternal folic acid supplementation and the risk of congenital heart defects. *Int. Heart J.*, 57: 725-728. https://doi. org/10.1536/ihj.16-054
- Yang, M., Gong, T., Lin, X., Qi, L., Guo, Y., Cao, Z., Shen, M. and Du, Y., 2013. Maternal gene polymorphisms involved in folate metabolism and the risk of having a down syndrome offspring: A meta-analysis. *Mutagenesis*, 28: 661-671. https:// doi.org/10.1093/mutage/get045
- Yigit, S., Karakus, N. and Inanir, A., 2013. Association of MTHFR gene C677T mutation with diabetic peripheral neuropathy and diabetic retinopathy. *Mol. Vis.*, **19**: 1626-1630.
- Yu, D., Zhuang, Z., Wen, Z., Zang, X. and Mo, X., 2017. MTHFR A1298C polymorphisms reduce the risk of congenital heart defects: A meta-analysis from 16 case-control studies. *Italian J. Pediat.*, 43: 108. https://doi.org/10.1186/s13052-017-0425-1
- Zhang, R., Huo, C., Wang, X., Dang, B., Mu, Y. and Wang, Y., 2018. Two common MTHFR gene polymorphisms (C677T and A1298C) and fetal congenital heart disease risk: An Updated metaanalysis with trial sequential analysis. *Cell Physiol. Biochem.*, **45**: 2483-2496. https://doi. org/10.1159/000488267