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# Deltamethrin Induced Changes in the Activities of Various Esterases in Deltamethrin-Resistant Populations of *Trogoderma granarium*

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## ABSTRACT

The present study was aimed to evaluate the toxic effects of deltamethrin on the level of various esterases in deltamethrin-resistant populations of *Trogoderma granarium* collected from some godowns of Punjab. The level of various esterases like total esterases, cholinesterase, acetylcholinesterase, arylesterase and carboxylesterase in 4<sup>th</sup>, 6<sup>th</sup> instar larvae and adult beetles of deltamethrin-resistant populations' *viz.*, Gujranwala, Okara and D.G. Khan was significantly increased as compared to susceptible population of *T. granarium* (population never exposed to any kind of insecticide/fumigant since 2001). Different developmental stages possessed different levels of esterase activities. Based on the level of activities the adult beetles were more susceptible to deltamethrin than 4<sup>th</sup> and 6<sup>th</sup> instar larvae. The increased level of esterases contributes towards resistance against pesticide in stored grain pests.

## **INTRODUCTION**

Theat is a leading food grain in Pakistan (Wajid, 2004; Goyal and Parasad, 2010) and is stored at farms in heaps, pots, baskets and bags covered by either straws, plastered or mud, house type godowns, PASSCO type godowns, hexagonal bins, silos and open bulk heads (Peng et al., 2011). During storage one third of the potential food supply is lost every year (Duveiller et al., 2007) and about 10-20% post-harvest losses are mainly caused by insect pests (Khan et al., 2010). Trogoderma granarium (Everts) is the most important insect species that adversely infest wheat in tropical regions of the world (Lowe et al., 2000; Ahmedani et al., 2011). A number of control methods have been used to control the pest population which include the use of botanical insecticide (Fields, 2006; Prakash and Rao, 2006; Musa and Dike, 2009; Gandhi et al., 2010), synthetic insecticides like organochlorines, carbamates, organophosphates and synthetic pyrethroids (Kljajic and Peric, 2006; Ali et al., 2007; Athanassiou et al., 2007), fumigants (Daglish, 2004), biological insecticides (Nayak et al., 2005) and application of physical agents like heat, temperature, pressure, aeration and relative humidity (Ofuya and Reichmuth, 2002; Mbata et al., 2004). Synthetic insecticides are most commonly



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Authors' Contribution FRS designed and supervised the research project. AH conducted the experimental work. AH, TR and FRS analyzed the data and wrote the article.

Key words Khapra beetle, Total esterases, Insecticide resistance, Carboxyl esterases, Pyrethroid insecticide.

used to control agricultural insect pests all over the world (Mathewes, 1993) but excessive and unplanned use of these pesticides results in the development of resistance (Fragoso *et al.*, 2003; Ribeiro *et al.*, 2003). Dletamethrin a synthetic pyrethroid is widely used in grain godowns before the fumigation process but many researchers have reported resistance in *T. granarium* against deltamethrin (Irshad and Iqbal, 1994; Tarakanov *et al.*, 1994; Saxena and Sinha, 1995; Kumar *et al.*, 2010).

Esterases cause the hydrolysis of ester containing pyrethroids into their corresponding alcohol and acid. They can also sequester insecticides by the formation of stable compounds so toxic insecticidal molecules may not be available for chemical reactions within insect body (Devonshire and Moores 1982; Oakeshott et al., 2005; Wheelock et al., 2005). Cholinesterase, acetylcholinesterase, carboxylesterase and arylestearse are important esterases involved in the process of detoxification and causing resistance for particular insecticide (Ellman et al., 1961; Fournier and Mutero, 1994) and elevated levels of esterases was studied in resistant strains of insect pests (Casida, 1973; Riaz et al., 2017). Also different developmental stages exhibit different levels of esterases as larval stages are found to be more resistant than adults (Nakakita and Winks, 1981; Riaz et al., 2017). The present study was aimed to investigate the abnormalities developed in the level of esterases in deltamethrin-resistant populations of T. granarium and to correlate the level of esterase activities with resistance.

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## **MATERIALS AND METHODS**

Deltamethrin resistant and susceptible populations of T. granarium used in this study were those used in Hafiz et al. (2017). The master culture of susceptible and resistant populations of Khapra beetle was reared according to Riaz et al. (2014), Shakoori et al. (2016) and Hafiz et al. (2017). The culture was maintained in 300ml sterilized jam jar covered with muslin cloth at 35±2°C with 60±5% relative humidity (Riaz et al., 2014). Whole wheat grains, crushed wheat and wheat floor was used as feed of different larvae (FAO, 1974). From homogeneous stock of each population 4th, 6th instar larvae and adult beetles were used to record toxicological data. To calculate  $LC_{50}$  values of 4th, 6th instar larvae and adult beetles of T. granarium mortality data was subjected to Probit analysis by Minitab 16 software (Finney, 1971) and were expressed in ppm. The LC<sub>50</sub> values of deltamethrin for these populations are presented in Hafiz et al. (2017).

## Biochemical analysis

Biochemical analysis for a number of esterase activities including carboxylesterase (CE), acetylcholine esterase (AchE), cholinesterase (ChE), total esterase (TE) and arylesterase (AE) were carried out. Using their respective standard curves, absorbance of various enzyme activities like CE, TE and ChE were converted into activity/quantity.

For estimation of various esterases, twenty larvae

(4<sup>th</sup> and 6<sup>th</sup> instar larvae) and twenty adult beetles of Khapra beetle from each population were taken in five replicates, each containing three test tubes. They were weighed and homogenized in their respective buffer (pH 7.0) by using motor driven Teflon glass homogenizer with consistent cooling in squashed ice. The activities of acetylcholinesterase and arylesteras were measured according to Devonshire (1975a) and Junge and Klees (1981), respectively. While the activities of carboxylesterase and Total esterases were determined according to the method of Devonshire (1975b). The activity of Choline esterase was measured according to Rappapot *et al.* (1959).

#### Statistical analysis

The biochemical data was subjected to one way ANOVA and Tukey's test to compare the significance difference between means of susceptible and resistant populations at P < 0.05 using Minitab 16 software. Values <0.05 were considered statistically significant.

## RESULTS

The activities of various esterases *viz.*, Total esterases, Cholinesterase, Acetylcholine esterases, Aryl esterases and Carboxyl esterases in three developmental stages (4<sup>th</sup> and 6<sup>th</sup> instar larvae and adult beetles) of deltamethrinsusceptible and three deltamethrin-resistant populations (Gujranwala, Okara and D.G. Khan) of *T. granarium* are presented in Table I. The activities of TE, ChE, AChE, AE

Table I.- Activities of various esterases (IU/mg body weight) of 4<sup>th</sup> and 6<sup>th</sup> instar larvae and adult beetles of susceptible and resistant populations of *T. granarium*.

Populations	ТЕ	ChE	AChE	AE	CE
4 <sup>th</sup> instar larvae					
Susceptible	*232.62±3.83 <sup>d</sup>	2.84±0.03 <sup>d</sup>	220.49±1.03 <sup>d</sup>	92.98±0.36 <sup>d</sup>	112.37±0.79°
Gujranwala	437.09±2.89ª	3.70±0.01ª	257.88±1.04ª	133.43±0.18 <sup>b</sup>	210.71±0.47ª
Okara	365.37±3.35 <sup>b</sup>	3.30±0.01 <sup>b</sup>	225.30±1.04b	101.77±0.16°	175.14±0.74 <sup>b</sup>
D.G. Khan	283.11±3.75°	3.08±0.06°	203.77±0.72°	140.20±0.17ª	129.85±0.66°
6 <sup>th</sup> instar larvae					
Susceptible	$274.64 \pm .46^{d}$	$1.86 \pm 0.07^{d}$	$124.64 \pm 0.54^{d}$	68.13±0.06°	$108.10 \pm 0.26^{d}$
Gujranwala	361.34±0.35ª	2.38±0.09ª	179.63±0.32ª	155.48±0.01 <sup>d</sup>	176.80±0.73ª
Okara	326.47±0.21b	2.24±0.01b	157.34±0.28 <sup>b</sup>	84.68±0.04ª	139.49±0.47 <sup>b</sup>
D.G. Khan	366.53±1.53°	1.97±0.02°	138.27±0.42°	69.46±0.07 <sup>b</sup>	121.50±0.33°
Adult beetles					
Susceptible	229.19±3.4°	8.32±0.13°	52.79±0.19 <sup>d</sup>	24.86±0.92 <sup>d</sup>	$48.91 \pm 0.34^{d}$
Gujranwala	275.73±3.1ª	$11.07{\pm}0.08^{a}$	70.94±0.21ª	81.04±0.92ª	90.97±0.23ª
Okara	265.12±3.14 <sup>ab</sup>	9.88±0.130 <sup>b</sup>	63.19±0.27 <sup>b</sup>	72.68±1.35 <sup>b</sup>	72.51±0.21b
D.G. Khan	256.27±1.98b	8.32±0.138°	57.16±0.17°	56.33±1.03°	64.58±0.27°

\*Mean $\pm$ SEM; n= 5 (Mean values in each single assay derived from five replicates and each replicate contains 20 beetles). Mean values with super script letters<sup>a,b,c,d</sup> within each column represents the significant differences among the means of various populations while mean values with common superscript letters indicates non-significant differences with others except susceptible population at *P* <0.05 according to Tukey's post hoc test.

and CE in deltamethrin-resistant populations were increased significantly as compared to deltamethrinsusceptible population at P<0.05 except the activity of ChE which are not significantly different in adult beetles of Gujranwala and susceptible populations. Similarly the TE activity in adult beetles of (D.G. Khan and Okara Population) and (Okara and Gujranwala populations) were not significantly different from each other at P<0.05. Different developmental stages (4<sup>th</sup> and 6<sup>th</sup> instar larvae and adult beetles) in all resistant populations possessed significantly different levels of esterases at P<0.05. Percent change in the activities of these esterases in deltamethrinresistant populations with reference to susceptible population is presented in Figure 1.

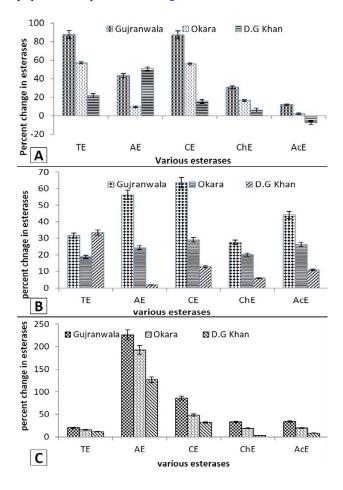


Fig. 1. Percent (%) change in various esterases of  $4^{th}$  instar (A),  $6^{th}$  instar (B) and adult beetles (C) of resistant population compared with susceptible population.

A decreasing trend was found in the activities of esterases from 4<sup>th</sup> instar larvae to 6<sup>th</sup> instar larvae and adult beetles except 6<sup>th</sup> instar larvae of D.G. Khan Population which possessed higher TE activity and 6<sup>th</sup> instar of

Gujranwala Populations which possessed higher AE activity than 4<sup>th</sup> instar larvae. Similarly adult beetles of Gujranwala population exhibited higher ChE activity than 6<sup>th</sup> and 4<sup>th</sup> instar larvae.

Among resistant populations the 4<sup>th</sup> instar larvae and adult beetles of Guiranwala population possessed highest T.E activity (87.90 and 20.31%) while 4th instar larvae and adult beetles of DG Khan population possessed lowest T.E activity (21.70 and 11.82%), respectively as compared to 4<sup>th</sup> instar larvae and adult beetles of susceptible population. On other hand, 6th instar larvae of D.G. Khan population possessed highest TE activity (33.45%) and 6th instar larvae of Okara population possessed lowest TE activity (18.87%) as compared to 6<sup>th</sup> instar larvae of susceptible population. The 4<sup>th</sup> and 6<sup>th</sup> instar larvae and adult beetles of Gujranwala population possessed highest CE activity (87.51, 63.55 and 86.01%) among resistant populations and the 4<sup>th</sup> instar larvae of DG Khan population possessed lowest activity (15.55, 13.00 and 31.99%), respectively as compared to 4<sup>th</sup>, 6<sup>th</sup> instar larvae and adult beetles of susceptible population.

The AChE activity was also found significantly increased in all populations with respect to susceptible population except the 4th instar larvae of D.G. Khan population. Among resistant populations, 4th and 6th instar larvae and adult beetles of Gujranwala population possessed highest AChE (17.00, 44.13 and 34.93%) and ChE activities (36.68, 27.60 and 33.15%) while the 4th and 6<sup>th</sup> instar larvae and adult beetles of D.G. Khan population possessed lowest AChE (-7.58, 10.94 and 8.28%) and ChE activities (6.07, 6.02 and 0.02%), respectively as compared to 4th and 6th instar larvae and adult beetles of susceptible population. The 4<sup>th</sup> instar larvae of D.G. Khan population possessed highest AE activity (50.78) and the 4<sup>th</sup> instar larvae of Okara population possessed lowest AE activity (9.45) as compared to 4<sup>th</sup> instar larvae of susceptible population. The 6th instar larvae and adult beetles of Guiranwala populations possessed highest AE activity (56.17 and 226.32%) while the 6<sup>th</sup> instar larvae and adult beetles of D.G. Khan population possessed lowest activity (1.94 and 126.80%), respectively as compared to 6<sup>th</sup> instar larvae and adult beetles of susceptible population. On the basis of increased levels of activities of various esterases, the pest populations can be graded as Gujranwala > Okara > D.G. Khan > Susceptible.

## DISCUSSION

The resistant populations of *T. granarium* have been collected from some stored grain godowns of the Punjab where deltamethrin has been applied on the grains prior to Phosphine application. The doses of the deltamethrin

have not been calculated periodically according to level of pest resistance, so as a result of indiscriminate exposure of deltamethrin to *T. granarium* in the stored grain houses the pest has developed resistance against deltamethrin. The resistance can be measured using different resistance indicators like TE, ChE, AChE, AE, and CE activities. It was investigated that the level of all esterases tested was found significantly increased in 4<sup>th</sup> and 6<sup>th</sup> instar larvae and adult beetles of all deltamethrin-resistant populations as compared to susceptible population.

In the present study, the elevated levels of TE are in accordance to the findings of Sher et al. (2004) who reported that TE activity was increased in 4th instar larvae of T. granarium after 10 h exposure to Phosphine in Haroonabad population. Lewis and Medge (1984) investigated higher levels of TE in foliar spray resistant strains of aphid as compared to susceptible strains. Riaz et al. (2017) also reported increase in TE in various Phosphine tolerant populations of T. granarium. Cholinesterases and Acetylcholine esterase belongs to important group of enzymes that play key role in nervous system and involve in conduction of nerve impulse at neuromuscular junction (Ollis et al., 1992; Walsh et al., 2001). Due to increased ChE and AchE activities, the acetylcholine may efficiently be converted into choline and various systems of the pests may coordinate timely so insect gain protection against insecticide and develop resistance (Riaz et al. 2017). The increased activities of ChE and AChE in present study are in favour of the findings of Sher et al. (2004) who reported increased activity of ChE in 4th instar larvae of T. granarium after Phosphine exposure. Riaz et al. (2017) also reported increase in ChE and AChE in various Phosphine tolerant populations of T. granarium. It is also reported in literature that the role of AChE in development of resistance is correlated with the alteration in AChE binding sites in insecticide resistant pests which leads to the insensitivity of the enzyme to insecticides inhibition as studied by Siegfried and Scott (1992). Dvir et al. (2010) reported that there are two sub sites in active site of AChE named as esteratic and anionic. The esteratic sub site is responsible for catalytic process while anionic sub site is responsible for binding of choline. The esteratic sub site comprises of catalytic triad that consist of three amino acids as serine, histidine and glutamate. At this catalytic site, hydrolysis of acetylcholine into choline and acetic acid takes place (Soreg and Seidman, 2001). AChE insensitivity is wellknown principal feature of resistant insects (Carpentier and Founeir, 2001). Karoly et al. (1996) reported that in resistant apple bud moths, the activity of AChE increased in each developmental stage when compared to susceptible

population. Bourguet *et al.* (1996) studied that AChE gene duplication may causes overproduction of AChE and results in insecticidal resistance. Guedes *et al.* (1997) find out that higher level of AChE activity in resistant populations of *R. Dominica* were less sensitive to malaoxon inhibition than the susceptible populations. Levitin and Cohen (1998) also reported that enhanced levels of ChE activity in *Aonidiella aurantii* is due to organophosphate resistance. Zhu and Gao (1999) evaluated that resistance to organophosphates in green bugs was due to elevated levels of AChE. Similarly Charpentier and Fournier (2001) and Rumpet *et al.* (1997) also reported that increased level of ChE and AChE are responsible for development of resistance against insecticides.

Carboxyl esterases are enzymes that are involved in the hydrolysis of carboxylic esters into alcohol and free acid anion (Krisch, 1971; Junge and Krisch, 1975; Cygler et al., 1993; Satoh and Hosokawa, 2006; Hosokawa et al., 2007). The enzymes play important role in detoxification and metabolism of many compounds (Potter and Wadkins, 2006) including carbamates (Sogorb and Vilanova, 2002), organophosphates (Casida and Quistad, 2004) and pyrethroids (Stok et al., 2004). Byrne et al. (2000), Oakeshott et al. (2005) and Cui et al. (2007) investigated that elevated levels of CE are involved in the development of resistance to agrochemicals, fumigants and pesticides. Riaz et al. (2017) also reported an increase in CE in five Phosphine tolerant populations of T. granarium. Likewise all deltramethrin-resistant populations showed significant increase in AE activity. Zhu and He (2000) investigated that higher level of AE activity in S. graminum as compared to susceptible population. Riaz et al. (2017) reported increased AE activity in Phosphine tolerant populations of T. granarium. Sher et al. (2004) investigated that the level of AE activity in 4th instar larvae of T. granarium in Khanewal population was decreased after exposure to 0.8ppm of phosphine while elevated level of AE activity was noticed in Haroonabad population after their exposure to phosphine for 80 h. This suggests that longer periods of exposure to insecticides with their sub lethal doses leads to insecticide tolerance at first which later on results in insecticidal resistance.

In current study, it was also found that Adult beetles have significantly low TE, ChE, AChE, AE, and CE activity than 4<sup>th</sup> and 6<sup>th</sup> instar larvae. Nakakita and Winks (1981) and Riaz *et al.* (2017) reported that the level of esterase changed throughout the life cycle in different developmental stages as larvae are more tolerant to pesticides than adult beetles. Kim *et al.* (1988) suggest that it is the developmental stage of insect which determines the resistance or susceptibility of insect to particular insecticide.

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#### Ethical standard

This article does not contain any studies with human participants or animals performed by any of the authors.

#### Statement of conflict of interest

The authors AH, TR and FRS stated no conflicts of interest.

## REFERENCES

- Ahmedani, M.S., Haque, M.I., Afzal, S.N., Naeem, M., Hussain T. and Naz. S., 2011. Quantitative losses and physical damage caused to wheat kernel (*Triticum aestivum*) by khapra beetle infestation. *Pak. J. Bot.*, **43**: 659-668.
- Ali, N.S., Munir, M., Ali, S.S. and Shakoori, A.R., 2007. Efficacy of mixtures of an organophosphate, malathion and a synthetic pyrethroid, deltamethrin against lesser grain borer, *Rhyzopertha dominica*. *Pakistan J. Zool.*, **39**: 179-184.
- Anand, P. and Jagadiswari, R., 2006. Exploitation of newer botanicals as rice grain protectants against Angoumois grain moth, *Sitotroga cerealella* Oliv. *Entomon*, **31**: 1-8.
- Athanassiou, C.G., Kavallieratos, N.G., Peteinatos, G.G., Petrou, S.E., Boukouvala, M.C. and Tomanovic, Z., 2007. Influence of temperature and humidity on insecticidal effect of three diatomaceous earth formulations against larger grain borer (Coleoptera: Bostrichidae). J. econ. Ent., 100: 599-603. https:// doi.org/10.1093/jee/100.2.599
- Bourguet, D., Raymond, M., Bisset, J., Pasteur, N. and Arpagaus, M., 1996. Duplication of the Ace.1 locus in *Culex pipiens* mosquitoes from the Caribbean. *Biochem. Genet.*, 34: 351-362. https://doi. org/10.1007/BF00554410
- Byrne, F.J., Gorman, K.J., Cahill, M., Denholm, I. and Devonshire, A.L., 2000. The role of B-type esterases in conferring insecticide resistance in the tobacco whitefly, *Bemisiatabaci* (Genn). *Pestic. Manage. Sci.*, 56: 867-874. https://doi.org/10.1002/1526-4998(200010)56:10<867::AID-PS218>3.0.CO;2-P
- Casida, J. (Ed.), 1973. *Pyrethrum: The natural insecticide*. Academic Press, USA.

- Casida, J.E. and Quistad, G.B., 2004. Organophosphate toxicology: safety aspects of non acetylcholine esterase secondary targets. *Chem. Res. Toxicol.*, 17: 983-998. https://doi.org/10.1021/tx0499259
- Charpentier, A. and Fournier, D., 2001. Levels of total acetylcholinesterase in *Drosophila melanogaster* in relation to insecticide resistance. *Pestic. Biochem. Physiol.*, **70**: 100-107. https://doi.org/10.1006/ pest.2001.2549
- Cui, F., Weill, M., Berthomieu, A., Raymond, M. and Qiao, C.L., 2007. Characterization of novel esterases in insecticide resistant mosquitoes. *Insect Biochem. mol. Biol.*, **37**: 1131-1137. https://doi. org/10.1016/j.ibmb.2007.07.002
- Cygler, M., Schrag, J.D., Sussman, J.L., Harel, M., Silman, I., Gentry, M.K. and Doctor, B.P., 1993. Relationship between sequence conservation and three dimensional structure in a large family of esterases, lipases and related proteins. *Protein Sci.*, 2: 366-382. https://doi.org/10.1002/ pro.5560020309
- Daglish, G.J., 2004. Effect of exposure period on degree of dominance of phosphine resistance in adults of *Rhyzopertha dominica* (Coleoptera: Bostrichidae) and *Sitophilus oryzae* (Coleoptera: Curculionidae). *Pest Manage. Sci.*, 60: 822-826. https://doi. org/10.1002/ps.866
- Devonshire, A.L. and Moores, G.D., 1982. A carboxylesterase with broad substrate specificity causes organophosphorus, carbamate and pyrethroid resistance in peach-potato aphids (*Myzuspersicae*). *Pestic. Biochem. Physiol.*, 18: 235-246. https://doi. org/10.1016/0048-3575(82)90110-9
- Devonshire, A.L., 1975a. Studies of the acetylcholinesterase from the house fly resistant and susceptible organophosphorus insecticides. *Biochem. J.*, 149: 463-469. https://doi.org/10.1042/ bj1490463
- Devonshire, A.L., 1975b. Studies of the carboxylesterase of *Myzuspersicae* resistant and susceptible to organophosphorus insecticides. *Proc. Br. Insect. Fungi Conf.*, **8**: 67-73.
- DuVeiller, E., Ravi, A.E., Singh, P., Julie, A.E. and Nicol, M., 2007. The challenges of maintaining wheat productivity: pests, diseases and potential epidemics. *Euphytica*, **157**: 417-430. https://doi. org/10.1007/s10681-007-9380-z
- Dvir, H., Silman, I., Harel, M., Rosenberry, T.L. and Sussman, J.L., 2010. Acetylcholinesterase: from 3D structure to function. *Chem. Boil. Interactions*, 187: 10-22. https://doi.org/10.1016/j.cbi.2010.01.042
- Ellman, G.L., Courtney, K.D. and Featherstone, R.M.,

1961. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.*, 7: 88-95. https://doi.org/10.1016/0006-2952(61)90145-9

- FAO, 1974. Recommended methods for the detection and measurement of resistance of agricultural pests to pesticides. Tentative method for adults of some major beetle pests of stored cereals with malathion or lindane. *FAO Pl. Prot. Bull.*, **22**: 127-137.
- Fields, P.G., 2006. Effect of *Pisumsativum* fractions on the mortality and progeny production of nine stored-grain beetles. *J. Stored Prod. Res.*, **42**: 86-96. https://doi.org/10.1016/j.jspr.2004.11.005
- Finney, D.J., 1971. *Probit analysis*, 3<sup>rd</sup> Ed., Cambridge University Press, London, pp. 333.
- Fournier, D. and Mutero, A., 1994. Modification of acetylcholinesterase as a mechanism of resistance to insecticides. *Comp. Biochem. Physiol. Part C: Pharmacol. Toxicol. Endocrinol.*, **108**: 19-31. https://doi.org/10.1016/1367-8280(94)90084-1
- Fragoso, D.B., Guedes, R.N.C. and Rezende, S.T., 2003.Glutathione-S-transferase detoxification as a potential pyrethroid resistance mechanism in the maize weevil, *Sitophiluszeamais. Ent. Exp. et Appl.*, **109**: 21–29. https://doi.org/10.1046/j.1570-7458.2003.00085.x
- Gandhi, N., Pillai, S. and Patel, P., 2010. Efficacy of pulverized *Punicagranatum* (Lythraceae) and *Murrayakoenigii* (Rutaceae) leaves against stored grain pest *Triboliumcastaneum* (Coleoptera: Tenebrionidae). *Int. J. agric. Biol.*, **12**: 616-620.
- Goyal, A. and Prasad, R., 2010. Some important fungal diseases and their impact on wheat production.
  In: *Management of fungal plant pathogens* (eds. A. Arya and A.E.V. Perelló). CABI (H ISBN 9781845936037), pp. 362.
- Guedes, R.N.C., Kambhampati, S., Dover, B.A. and Zhu, K.Y., 1997. Biochemical mechanisms of organophosphate resistance in *Rhyzopertha dominica* (Coleoptera: Bostrichidae) populations from the United States and Brazil. *Bull. Ent. Res.*, 87: 581-586. https://doi.org/10.1017/ S0007485300038670
- Hafiz, A., Riaz, T. and Shakoori, F.R., 2017. Metabolic profile of a stored grain pest *Trogoderma granarium* exposed to deltamethrin. *Pakistan J. Zool.*, **49**: 8-12.
- Hosokawa, M., Furihata, T., Yaginuma, Y., Yamamoto, N., Koyano, N., Fujii, A., Nagahara, Y., Satoh, T. and Chiba, K., 2007. Genomic structure and transcriptional regulation of the rat, mouse, and humancarboxylesterasegenes. *Drug Metab. Rev.*, 39:

1-15. https://doi.org/10.1080/03602530600952164

- Irshad, M. and Iqbal, J., 1994. Phosphine resistance in important stored grain insect pests in *Pakistan*. *Pakistan J. Zool.*, **26**: 347-350.
- Junge, W. and Krisch, K., 1975. The carboxylesterase/ amidases of mammalian liver and their possible significance. *CRC. Crit. Rev. Toxicol.*, **3**: 371-434. https://doi.org/10.3109/10408447509079864
- Junge, W. and Klees, H., 1981. Arylesterase. In: *Method* of enzyme analysis, 3<sup>rd</sup> Ed., Vol. 4, Enzyme 2 Esterases, glycosidases, ligases. Verlag Chemic, Florida, pp. 8-14.
- Karoly, E.D., Rose, R., Thompson, D.M., Hodgson, E., Rock, G.C. and Roe, R.M., 1996. Monooxygenase esterase and glutathione S-transferase activity associated with azinphosmethyl resistance in the tufted apple bud moth, *PlatynotaIdae usalis*. *Pestic. Biochem. Physiol.*, **55**: 109-121. https://doi. org/10.1006/pest.1996.0040
- Khan, I., Afsheen, S., Din, N., Khattak, S., Khalil, S.K. and Lou, Y.H.Y., 2010. Appraisal of different wheat genotypes against angoumois grain moth, *Sitotroga ceralella* (Oliv). *Pakistan J. Zool.*, **42**: 161-168.
- Kijajic, P. and Peric, I., 2006. Susceptibilty to contact insecticides of granary weevil *Stophilus granaries* (L.) (Coleoptera: Curculionidae) originating from different locations in the former Yugoslavia. *J. Stored Prod. Res.*, 42: 149-161. https://doi.org/10.1016/j. jspr.2005.01.002
- Kim, G.H., Ahn, Y.J. and Cho, K.Y., 1988. Susceptibility of insecticides to the developmental stages in the bean bug (*Riptortus clavatus*). *Korean J. Ent.*, 18: 269-274.
- Krisch, K., 1971. Carboxylesterase. In: *The enzyme* (ed. P.D. Boyer), Vol. V. Academic Press, New York, pp. 43-69.
- Kumar, M.K., Srivastava, C. and Garg, A.K., 2010. In vitro selection of deltamethrin resistant strain of Trogoderma granarium and its susceptibility to insecticides. Annls. Pl. Prot. Sci., 18: 26-30.
- Levitin, E. and Cohen, E., 1998. The involvement of acetylcholinesterase in resistance of the California red scale *Aonidiella aurantii* to organophosphorus pesticides. *Ent. exp. Appl.*, **88**: 115-121.
- Lewis, G.A. and Madge, D.S., 1984. Esterase activity and associated insecticide resistance in the damson hop aphid, *Phorodon humuli* (Schrank) (Hemiptera: Aphididae). *Bull. Ent. Res.*, 74: 227-238. https:// doi.org/10.1017/S0007485300011366
- Lowe, S., Browne, M., Boudjelas, S. and Depoorter, M., 2000. 100 of the World's worst invasive alien species: A selection from the global invasive species

*database*. Invasive Species Specialist Group, World Conservation Union (IUCN). Available online at http://www.issg.org/booklet.pdf. Accessed 27 September 2005

- Matthews, G.A., 1993. Developments in the application of pesticides. In: *Modern crop protection: Developments and perspectives* (ed. J.C. Zadoks). Wageningen Academic Publishers, The Netherlands, pp. 61-68.
- Mbata, G.N., Phillips, T.W. and Payton, M., 2004. Mortality of eggs of stored-product insects held under vacuum: effects of pressure, temperature, and exposure time. *J. econ. Ent.*, **97**: 695-702. https://doi.org/10.1093/jee/97.2.695
- Mehlhorn, H., 2011. Nature helps. In: Parasitology research monographs. Springer, Heidelberg, Berlin. https://doi.org/10.1007/978-3-642-19382-8
- Musa, A.K. and Dike, M.C., 2009. Life cycle, morphometrics and damage assessment of the khapra beetle, *Trogoderma granarium* everts (Coleoptera: Dermestidae) on stored groundnut. *J. agric. Sci.*, **52**: 135-142. https://doi.org/10.2298/ JAS0902135M
- Nakakita, H. and Winks, R.G., 1981. Phosphine resistance in immature stages of a laboratory selected strain of *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). J. Stored Prod. Res., 17: 43–52. https://doi.org/10.1016/0022-474X(81)90016-3
- Nayak, M.K., Daglish, G.J. and Byrne, V.S., 2005. Effectiveness of spinosad as a grain protectant against resistant beetle and psocid pests of stored grain in Australia. J. Stored Prod. Res., 41: 455-467. https://doi.org/10.1016/j.jspr.2004.07.002
- Oakeshott, J.G., Devonshire, A.L., Claudianos, C., Sutherland, T.D., Horne, I., Campbell, P.M., Ollis, D.L. and Russell, R.J., 2005. Comparing the organophosphorus and carbamate insecticide resistance mutations in choline and carboxylesterases. *Chem. Biol. Interact.*, **157-158**: 269-275. https://doi.org/10.1016/j.cbi.2005.10.041
- Ofuya, T.I. and Reichmuth, C., 2002. Effect of relative humidity on the susceptibility of *Callosobruchus maculates* (Fabricius) (Coleoptera: Bruchidae) to two modified atmospheres. J. Stored Prod. Res., **38**: 139-146. https://doi.org/10.1016/S0022-474X(01)00009-1
- Ollis, D.L., Cheah, E., Cygler, M., Dijkstra, B., Frolow, F., Franken, S.M., Harel, M., Remington, S.J., Silman, I., Schrag, J., Sussman, J.L., Verschueren, K.H.G. and Goldman, A., 1992. The α/β hydrolase fold. *Protein Engg.*, **5**: 197-211. https://doi.

org/10.1093/protein/5.3.197

- Peng, J., Sun, D. and Nevo, E., 2011. Wild emmer wheat, *Triticum dicoccoides*, occupies a pivotal position in wheat domestication. *Australian J. Crop Sci.*, 5: 1127-1143.
- Potter, P.M. and Wadkins, R.M., 2006. Carboxylesterases detoxifying enzymes and targets for drug therapy. *Curr. Med. Chem.*, **13**: 1045–1054. https://doi. org/10.2174/092986706776360969
- Rappaport, F., Fischil, J. and Pinto, N., 1959. An improved method for the determination of cholinesterase activity in serum. *Clin. Chem. Acta*, 4: 227-230. https://doi.org/10.1016/0009-8981(59)90134-2
- Riaz, T., Shakoori, F.R. and Ali, S.S., 2013. Effect of temperature on the development, survival, fecundity and longevity of stored grain pest, *Trogoderma* granarium. Pakistan J. Zool., 46: 1485-1489.
- Riaz, T., Shakoori, F.R. and Ali, S.S., 2014. Effect of temperature on the development, survival, fecundity and longevity of stored grain pest, *Trogoderma* granarium. Pakistan J. Zool., 46: 1485-1489.
- Riaz, T., Shakoori, F.R. and Ali, S.S., 2017. Effect of Phosphine on esterases of larvae and adult beetles of phosphine-exposed populations of stored grain pest, *Trogoderma granarium* collected from different Godowns of Punjab. *Pakistan J. Zool.*, **49**: 819-824.
- Ribeiro, B.M., Guedes, R.N.C., Oliveira, E.E. and Santos, J.P., 2003. Insecticide resistance and synergism in Brazilian populations of *Sitophiluszeamais* (Coleoptera: Curculionidae). *J. Stored Prod. Res.*, **39**: 21–31. https://doi.org/10.1016/S0022-474X(02)00014-0
- Rumpet, S., Hetzel, F. and Frampton, C., 1997. Lacewings (Neuroptera: Hemerobiidae: Chrysopidae) and integrated pest management enzyme activity as biomarker of sublethal insecticide exposure. J. econ. Ent., 90: 102-108.
- Satoh, T. and Hosokawa, M., 2006. Structure, function and regulation of carboxylesterases. *Chem. Biol. Interact.*, **162**: 195-211. https://doi.org/10.1016/j. cbi.2006.07.001
- Saxena, J.D. and Sinha, S.R., 1995. Evaluation of some insecticides against malathion resistant strains of red flour beetle, *Tribalium castaneum* (Herbst). *Indian J. Ent.*, **75**: 401-405.
- Shakoori, F.R., Feroze, A. And Riaz, T., 2016. Effect of sub-lethal doses of phosphine on macromolecular concentrations and metabolites of adult beetles of stored grain pest, *Trogoderma granarium*, previously exposed to phosphine. *Pakistan J. Zool.*,

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**48**: 583-588.

- Sher, F., Ali, S.S. and Shakoori, A.R., 2004. Phosphine induced changes in various esterase levels in 4<sup>th</sup> instar larvae of *Trogoderma granarium*. *Pakistan J. Zool.*, **36**: 257-260.
- Siegfried, B.D. and Scott, J.G., 1992. Biochemical charecterization of hydrolytic and oxidative enzymes associated chlorpyrifos and propoxes resistance in German cockroach, *Blatella germanica* (L). *J. econ. Ent.*, **85**: 1892-1098.
- Sogorb, M.A. and Vilanova, E., 2002. Enzymes involved in the detoxification of organophosphorus, carbamate and pyrethroid insecticides through hydrolysis. *Toxicol. Lett.*, **128**: 215-228. https://doi.org/10.1016/S0378-4274(01)00543-4
- Soreq, H. and Seidman, S., 2001. Acetylcholinesterase new roles for an old actor. *Nat. Rev. Neurosci.*, 2: 294-302. https://doi.org/10.1038/35067589
- Stok, J., Huang, H., Jones, P.J., Wheelock, C.E., Morisseau, C. and Hammock, B.D., 2004. Identification, expression and purification of a pyrethroid hydrolysing carboxylesterase from mouse liver microsomes. *J. biol. Chem.*, 279: 29863-29869. https://doi.org/10.1074/jbc.M403673200
- Tarakanov, I.A., Kurambaev, Y., Khusinov, A.A. and Safonov, V.A., 1994. Respiratory and circulatory disorders in experimental poisoning with an organophosphorus pesticide. *Bull. exp. Biol. Med.*, 117: 466-471. https://doi.org/10.1007/BF02444326
- Wajid, S.A., 2004. Modelling development, growth

and yield of wheat under different sowing dates, plant populations and irrigation levels. Ph.D. thesis Department of Agronomy, University of Agriculture, Faisalabad, Pakistan.

- Walsh, S., Dolden, T., Moores, G., Kristensen, M., Lewis, T., Devonshire, A.L. and Williamson, M., 2001. Identification and characterization of mutations in housefly (*Muscadomestica*) acetylcholinesterase involved in insecticide resistance. *Biochem. J.*, 359: 175-181. https://doi.org/10.1042/bj3590175
- Wheelock, C.E., Miller, J.L., Miller, M.G., Shan, G., Gee, S.J. and Hammock, B.D., 2005. Development of toxicity identification evaluation (TIE) procedures for pyrethroid detection using esterase activity. *Environ. Toxicol. Chem.*, 23: 2699-2708. https://doi.org/10.1897/03-544
- Zhu, K.Y. and Gao, J.R., 1999. Increased activity associated with reduced sensitivity of acetylcholine esterase in organophosphate resistant green bug, *Schizaphis graminum* (Homoptera: Aphididae). *Pestic. Sci.*, **55**: 11-17. https://doi.org/10.1002/ (SICI)1096-9063(199901)55:1<11::AID-PS850>3.3.CO;2-W
- Zhu, K.Y. and He, F., 2000. Elevated esterases exhibiting arylesterase like ch.aracteristics in an organophosphate resistant clone of the greenbug, *Schizaphis graminum* (Homoptera: Aphididae). *Pestic. Biochem. Physiol.*, **67**: 155-167. https://doi. org/10.1006/pest.2000.2488