Short Communication

Supercooling Capacity of the Sweet Potato Leaf Folder, *Brachmia macroscopa* Meyrick (Lepidoptera: Gelechiidae)

Xia-Lin Zheng¹, Pan Wang², Jun-Yan Liu¹, Yu-Jing Zhang¹, Jun Li¹ and Wen Lu^{1,*}

¹Guangxi Key Laboratory of Agric-Environment and Agric-Products Safety, College of Agriculture, Guangxi University, Nanning 530004, China ²Institute of Vegetable Wuhan Academy of Agricultural Science and Technology, Wuhan 430345, China

Xia-Lin Zheng and Pan Wang contributed equally to this work.

ABSTRACT

The supercooling point is a good predictor of cold hardiness. In this study, supercooling capacity of *Brachmia macroscopa* was determined. The 4th instar larvae, pupae and adults survived in January, in which their supercooling points were -12.79 ± 0.67 °C, -19.92 ± 0.66 °C and -21.23 ± 0.99 °C, respectively. Body water contents of these stages in *B. macroscopa* have no negative or positive correlation with the supercooling points. Supercooling points of these stages are lower than the air temperature during the winter, which suggested that these stages are the potential overwintering stages of this species in Wuhan City.

Temperature plays a key role in the seasonal adaptation of insects. During seasonal cycles, many insect species are frequently exposed to stressful low temperatures that present a major challenge to survival (Hance *et al.*, 2007). Insects can adopt behavioral and physiological strategies to adapt to these stressful climatic conditions (Bale, 2002; Sinclair *et al.*, 2003).

Cold hardiness is an essential component of winter survival for most insects. Cold hardiness strategies of insects have generally been divided into two major categories: freeze tolerance insects tolerate the formation of extracellular ice within the body, whereas freeze avoidance insects cannot tolerate the formation of ice within their bodily fluid by depressing the temperature at which the spontaneous freezing of body water occurs (Zachariassen, 1985; Denlinger and Lee, 2010). This value is termed "supercooling point" and is experimentally determined by detecting the released latent heat of fusion as body water freezes (Duman, 2001; Denlinger and Lee, 2010). Understanding the cold hardiness is critical to explore the adaptive evolution between insects and their environment. The sweet potato leaf folder, Brachmia macroscopa Meyrick, 1932 (Lepidoptera: Gelechiidae) is one of the



Article Information Received 22 February 2017 Revised 23 March 2017 Accepted 01 April 2017 Available online 15 March 2018

Authors' Contribution

XLZ and PW conceived and designed the study. XLZ, JYL and YJZ wrote the article. JL and WL analyzed the data.

Key words

Brachmia macroscopa, Overwintering, Supercooling point, Cold hardiness, Body water content.

most dangerous herbivores of economic crops (e.g., Ipomoea batatas L., I. aquatica Forsk. and Dioscorea opposita Thunb.) in India, Philippines, Burma, Vietnam, China, Korea and Japan (Nakayama, 1939; Khurana et al., 1974; Hirano and Muramoto, 1976a, b; Hirano et al., 1976; Muramoto et al., 1976; Wu, 1982; Zhang et al., 2003; Zheng and Wang, 2009; Jiang, 2010; Wen and Wang, 2010; Wang and Tan, 2011). Previous studies of this species have focused on morphology (Nakayama, 1939; Zheng and Wang, 2009), biology (Hirano and Muramoto, 1976a; Wu, 1982; Zhang et al., 2003; Zheng and Wang, 2009; Jiang, 2010; Wen and Wang, 2010; Wang and Tan, 2011), and chemical and biological control (Khurana et al., 1974; Hirano et al., 1976; Hirano and Muramoto, 1976b; Muramoto et al., 1976; Chen and Chen, 1982; Huang, 1983). The distribution of *B. macroscopa* in China ranges from the Guangxi Zhuang Autonomous Region in the south to Heilongjiang Province in the north (Zhang, 1981; Wang and Tan, 2011). The number of generations per year that this species can complete 3-4 generations in Beijing and Shandong Provinces (Chi et al., 2000), 4-5 generations in Hubei and Zhejiang Provinces (Zheng and Wang, 2009), 5-7 generations in Jiangxi and Hunan Provinces (Zhang, 1981; Xiong, 2007; Jiang, 2010; Wen and Wang, 2010), and 8-9 generations in Guangxi and Guangdong Provinces (Zheng and Wang, 2009). Although B. macroscopa is an occasional pest in Asia, it causes considerable damage to economic

Corresponding author: luwenlwen@163.com
0030-9923/2018/0002-0779 \$ 9.00/0
Copyright 2018 Zoological Society of Pakistan

crops in summer and autumn. As for the outbreak population, which source whether come from overwintering population at local or migrate from southern regions is still unclear. Study on the supercooling capacity is useful for us to judge the possibility that *B. macroscopa* can overwinter at local based on the meteorological data.

The objective of the present study was to gain insight into the cold hardiness of *B. macroscopa* in temperate climatic zones. We assessed the supercooling capacity of *B. macroscopa* larvae, pupae and adults in Wuhan City.

Materials and methods

Brachmia macroscopa larvae were collected from sweet potato fields (114°20′E, 30°28′N) on *Ipomoea* batatas L. (Solanales: Convolvulaceae) in Wuhan suburb, Hubei Province, P. R. China in October, 2010. *Ipomoea* batatas was also collected from the field and cultured in the incubator (SPX-250IC, Boxun Medical Instrument Manufacturer of Shanghai, Shanghai, P. R. China) at 26 \pm 1 °C with a 16L: 8D photoperiod and 70-80% humidity. Larvae were reared with leaves of the host plant in a Petri dish ($\Phi = 9$ cm) at room temperature to allow pupation or eclosion. Leaves were replaced daily. In January 2011 (air temperature was the lowest during the winter in this month; Fig. 1), all stages, including 4th instar larvae, pupae and adults (3 days old), were sampled for the following experiment.

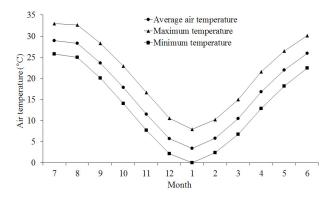


Fig. 1. Seasonal changes in air temperature (average, maximum and minimum) during 1951-2011 periods in Wuhan City (Hubei Province, China). Air temperature data provided by Climate Data Centre, China Meteorological Administration.

Supercooling point was measured using surfacecontact thermometry following the approach developed by Zheng *et al.* (2011). The larvae were weighed for fresh weight, and then their supercooling point was determined. Each larva was weighed again after oven drying for 24 h at 60 °C to determine dry weight. For the supercooling point determination, a larva (or pupa or adult) was fixed in a 1-mL polyethylene pipette tip to a copper-constantan thermocouple linked to an automatic multichannel temperature recorder (Jiangsu Senvi Economic Development Co., Ltd., Jiangsu Province, China). The pipette tip opening was plugged with cotton wool, which anchored the sensing junction of the thermocouple against the larval (or pupal or adult's) body. The thermocouple, together with the specimens protected by the pipette tips, was placed into a water bath (DW-40L188, Qingdao Haier Medical and Low Temperature Technology Co., Ltd., Shandong Province, China) that resulted in a highly repeatable temperature decline rate that corresponded to an approximate 1.5 °C min⁻¹ decrease from room temperature to 0 °C that slowed to approximately 0.5 °C min⁻¹ decrease sub-zero. Larval body temperatures were recorded at 1 s intervals. We ended each observation after 20 min at the minimum freeze temperature. A total of 30, 35 and 16 individuals for 4th instar larvae, pupae and adults were respectively observed. Body water content was measured according to the following formula:

Body water content (%) = [(fresh weight – dry weight) / fresh weight] \times 100

Statistical analysis was used SPSS 16.0 (SPSS Inc., Chicago, Illinois, U.S.A.). Supercooling points and body water contents were compared using one-way analysis of variance (ANOVA), followed by Tukey HSD test for multiple comparisons. Regression analysis was used to determine the relationship between supercooling points and body water contents. A level of $P \le 0.05$ was accepted as statistically significant.

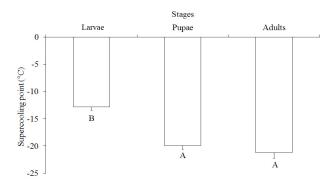


Fig. 2. Supercooling points of larvae, pupae, and adults of *Brachmia macroscopa* in January.

Results

Supercooling capacities of 4th instar larvae, pupae and adults in *B. macroscopa* have significant difference ($F_{2,80}$ = 36.85, P < 0.001). Although supercooling capacities of 4th

instar larvae were lower than pupae and adults, there is no statistical difference between the latter two stages (Fig. 2).

Body water contents of pupae and adults were significantly lower than 4th instar larvae ($F_{2,80} = 98.50$, P < 0.001). However, there is no statistical difference both pupae and adults (Fig. 3). Moreover, supercooling points of 4th instar larvae, pupae and adults have no negative or positive correlation with body water contents (Fig. 4).

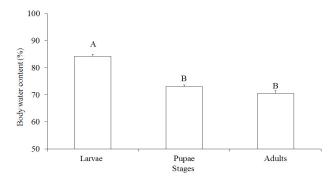


Fig. 3. Body water contents of larvae, pupae, and adults of *Brachmia macroscopa* in January.

Discussion

Our data illustrated that supercooling capacity of larvae, pupae and adults in *B. macroscopa* was obviously lower than the air temperature during the winter. Results suggested that larvae, pupae and adults are the potential overwintering stages, which correspond to the data of the field investigation (unpublished data). This information is helpful to understand the local population sources, establish an accurate forecasting system and draw up an effective plan against this species (Bale and Hayward, 2010).

Survival of insects during winter is often predicted by laboratory experiments, like freezing body temperature measurements (*i.e.*, supercooling point). Supercooling point, which is the temperature at which their body water freezes, regard as the lower limit of survival (Zachariassen, 1985). So, the supercooling capacity was one of the important predictors of cold hardiness in insects. In this study, supercooling capacity of overwintering larvae, pupae and adults in *B. macroscopa* was obviously lower than the air temperature during the winter. This result hints that the sweet potato leaf folder has potential ability to resist the extreme cold temperature in Wuhan City.

Bale (1996) indicated that the body water content was an essential component for the development and growth of insects and played an important role in cold hardiness due to its effects on the concentration of low-molecularweight substances and the activity of ice-nucleating agents. Generally, a high percentage of body water content negatively affects the cold hardiness and survival of insects (Zachariassen, 1991; Chapman, 1998; Holmstrup *et al.*, 2002; Sinclair *et al.*, 2013).

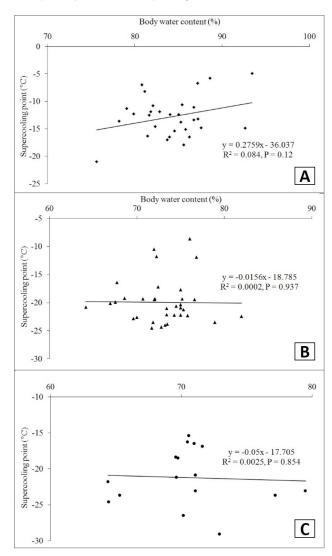


Fig. 4. Regression analysis between supercooling points and body water contents of larvae (A), pupae (B) and adults (C) in *Brachmia macroscopa*.

In our experiment, the body water contents of 4^{th} instar larvae, pupae and adults in *B. macroscopa* have no negative or positive correlation with the supercooling points. We speculated that the cause as a consequence of it's physiologically complex in their body. It is well known that the relationship between supercooling point and body water content is very complex and is influenced by a variety of other physical and chemical factors within and among various species, such as the level and combination of biochemical antifreeze compounds, the presence of ice

nucleating bacteria, antifreeze proteins, and the presence or absence of nucleating agents. Thus, further systematic studies are required to explore this issue.

It is well known that supercooling point is not the only predictor for survival of insects during the winter. Exposure time to low temperature (non-lethal temperature) couldn't be neglected when survival of insects under natural conditions was accurately evaluated. Many studies confirmed that long-term exposure to subfreezing nonlethal lower temperature can also decrease the survival of insects. For example, indirect chilling injury, which is caused by the long-term exposure to the low temperature above freezing point, found in the beet armyworm, Spodoptera exigua. Results indicated that several dayexposures to 5°C gave significant nonfreezing injuries such as low egg hatchability, retarded larval development, and low pupation rates (Kim and Song, 2000). In Wuhan City, air temperature was the lowest in January, in which days below 0 and 5°C was 11 and 30 days, respectively (Supplementary Table I). In this study, the survival of B. macroscopa exposure to non-lethal low temperature was not tested. From the perspective of cold hardiness of insects, this experiment will need to be further studied.

Acknowledgments

We are grateful to the anonymous referees and editor for their valuable comments on an earlier version.

Supplementary material

There is supplementary material associated with this article. Access the material online at: http://dx.doi. org/10.17582/journal.pjz/2018.50.2.sc5

Statement of conflict of interest

Authors have declared no conflict of interest. .

References

- Bale, J.S., 1996. I. Eur. J. Ent., 93: 369-382.
- Bale, J.S., 2002. *Phil. Trans. R. Soc. Lond. B*, **357**: 849– 862. https://doi.org/10.1098/rstb.2002.1074
- Bale, J.S. and Hayward, S.A.L., 2010. *J. exp. Biol.*, **213**: 980–994. https://doi.org/10.1242/jeb.037911
- Chapman, R.F., 1998. The insects: structure and function (4th edition). Cambridge University Press, New York, pp. 788. https://doi.org/10.1017/ CBO9780511818202
- Chen, Y.H. and Chen, Y.M., 1982. Fujian Agric. Sci. Technol., 6: 25–28.
- Chi, M.F., Jiang, H.S., Qin, S.M., Wu, H.X. and Leng, D.X., 2000. B. Agric. Sci. Technol., 8: 25.
- Denlinger, D.L. and Lee Jr., R.E., 2010. Low temperature

biology of insects. Cambridge University Press, UK, pp. 406. https://doi.org/10.1017/ CBO9780511675997

- Duman, J.G., 2001. Annu. Rev. Physiol., 63: 327–357. https://doi.org/10.1146/annurev.physiol.63.1.327
- Hance, T., van Baaren, J., Vernon, P. and Boivin, G., 2007. *Annu. Rev. Ent.*, **52**: 107–126. https://doi. org/10.1146/annurev.ento.52.110405.091333
- Hirano, C. and Muramoto, H., 1976a. *Appl. Ent. Zool.*, **11**: 154–159.
- Hirano, C. and Muramoto, H., 1976b. *Kontyû, Tokyo*, **44**: 214–216.
- Hirano, C., Muramoto, H. and Horiike, M., 1976. *Naturwissenschaften*, 63: 439. https://doi. org/10.1007/BF00599423
- Holmstrup, M., Bayley, M. and Ramløv, H., 2002. *Proc. natl. Acad. Sci. U.S.A.*, **99**: 5716–5720. https://doi. org/10.1073/pnas.082580699
- Huang, M., 1983. Chin. J. appl. Ent., 20: 129-130.
- Jiang, H.M., 2010. J. Res., 4: 173-175.
- Kim, Y. And Song, W., 2000. J. Asia-Pac. Ent., **3**: 49– 53.
- Muramoto, H., Souda, E. and Hirano, C., 1976. *Japan. J. appl. Ent. Zool.*, **20**: 77–80. https://doi. org/10.1303/jjaez.20.77
- Nakayama, S., 1939. J. Pl. Prot. (Tokyo), 26: 159-161.
- Sinclair, B.J., Ferguson, L.V., Salehipour-Shirazi, G. and Macmillan, H.A., 2013. *Integr. Comp. Biol.*, 53: 545–556. https://doi.org/10.1093/icb/ict004
- Sinclair, B.J., Vernon, P., Klok, C.J. and Chown, S.L., 2003. *Trends Ecol. Evol.*, 18: 257–262. https://doi. org/10.1016/S0169-5347(03)00014-4
- Wang, J.L. and Tan, R.R., 2011. *J. Changjiang Vegetab.*, **4**: 75–77.
- Wen, L. and Wang, C.C., 2010. *China Vegetab.*, **13**: 28–29.
- Wu, Z.Q., 1982. J. Fujian Agric. Coll., 2: 53-57.
- Xiong, S.G., 2007. Biol. Dis. Sci., 30: 32.
- Zachariassen, K.E., 1985. Physiol. Rev., 65: 799-832.
- Zachariassen, K.E., 1991. *Insects at low temperature* (eds. R.E. Lee Jr. and D.L. Denlinger), Chapman and Hall, New York, pp. 47–63. https://doi. org/10.1007/978-1-4757-0190-6_3
- Zhang, J.C., Cong, Z.F., Song, Y.Y., Zhang, F. and Sun, M., 2003. . J. Shandong Agric. Univ., **34**: 206–208.
- Zhang, S.M., 1981. Sci. Technol., 11: 10-11.
- Zheng, X.L., Cheng, W.J., Wang, X.P. and Lei, C.L., 2011. Cryobiology, 63: 164–169. https://doi. org/10.1016/j.cryobiol.2011.07.005
- Zheng, X.L. and Wang, P., 2009. *J. Changjiang Vegetab.*, **21**: 35–36.